Sex Steroid Regulation of the Inflammatory Response: Sympathoadrenal Dependence in the Female Rat

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To investigate the role of sex steroids in sex differences in the response of rats to the potent inflammatory mediator bradykinin (BK), we evaluated the effect of sex steroid manipulation on the magnitude of BK-induced synovial plasma extravasation (PE). The magnitude of BK-induced PE is markedly less in females. Ovariectomy of female rats increased BK-induced PE, and administration of 17β-estradiol to ovariectomized female rats reconstituted the female phenotype. Castration in male rats decreased BK-induced PE, and administration of testosterone or its nonmetabolizable analog dihydrotestosterone reconstituted the male phenotype. The results of these experiments strongly support the role of both male and female sex steroids in sex differences in the inflammatory response.

Because the stress axes are sexually dimorphic and are important in the regulation of the inflammatory response, we evaluated the contribution of the hypothalamic–pituitary–adrenal and the sympathoadrenal axes to sex differences in BK-induced PE. Neither hypophysectomy nor inhibition of corticosteroid synthesis affected BK-induced PE in female or male rats. Adrenal denervation in females produced the same magnitude increase in BK-induced PE as adrenalectomy or ovariectomy, suggesting that the adrenal medullary factor(s) in females may account for the female sex steroid effect on BK-induced PE. Furthermore, we have demonstrated that in female but not male rats, estrogen receptor α immunoreactivity is present on medullary but not cortical cells in the adrenal gland. These data suggest that regulation of the inflammatory response by female sex steroids is strongly dependent on the sympathoadrenal axis, possibly by its action on estrogen receptors on adrenal medullary cells.

Key words: plasma extravasation; inflammation; sex differences; estrogen; testosterone; estrogen receptor; sympathoadrenal axis; hypothalamic-pituitary-adrenal axis

Most inflammatory rheumatic diseases (e.g., rheumatoid arthritis and systemic lupus erythematosus) are more common in women (Da Silva, 1995). In addition, severity of rheumatoid arthritis (Deighton et al., 1992; Katz and Crisswell, 1996), in particular joint destruction (Weyand et al., 1998), is greater in women. In animal models of inflammatory disease, similar sex differences have been reported; for example, in rodents, the susceptibility to and severity of experimental arthritis is greater in females (MacKenzie et al., 1979; Wilder et al., 1982; Allen et al., 1983; Griffiths et al., 1994; Holmdahl, 1995). Research in both humans and animals has implicated a role for sex steroids in susceptibility to and severity of inflammatory rheumatic diseases (Da Silva et al., 1994; Jemec and Heidenheim, 1995; Josefsson and Tarkowski, 1997). For example, sex differences in the severity of experimental arthritis (number of joints involved, degree of erythema, and swelling) are abolished in male rats following either castration or administration of the principal female sex steroid 17β-estradiol (Allen et al., 1983).

Stress may also contribute to severity of arthritis because the sympathoadrenal axis modulates the inflammatory response (α-adrenergic receptor activation decreases severity whereas β2-adrenergic activity increases severity in models of chronic inflammation) (Coderre et al., 1990, 1991; Miao et al., 1992a; Lundeberg et al., 1993). Of note, the sympathetic and sympathoadrenal stress axes are sexually dimorphic; for example, female rats exhibit higher basal (DeTurck and Vogel, 1980) and stimulated (Livezey et al., 1985; Taylor et al., 1989) plasma epinephrine and norepinephrine levels, muscle sympathetic nerve activity (burst frequency and burst incidence) is higher in women (Ng et al., 1993), and stress-induced increases in plasma norepinephrine in men are enhanced by estradiol treatment (Kirschbaum et al., 1996). In addition, androgens also modulate sympathoadrenal activity in male rats (Le Thu et al., 1984) and men (Del Rio et al., 1995). Interestingly, although epinephrine acts via β2-adrenergic receptors to inhibit synovial plasma extravasation, it also produces, by the same mechanism, exacerbatation of joint damage in complete Freund’s adjuvant-induced arthritis in rats (assessed radiographically) (Coderre et al., 1990, 1991). This inverse relationship between magnitude of synovial plasma extravasation and severity of experimental arthritis occurs with other pharmacological interventions (Green et al., 1991; Miao et al., 1992b), indicating that the net effect of increasing plasma extravasation is to contribute to tissue repair/protection rather than tissue injury. In fact, recently it has been noted that physiological control of inflammation occurring after local generation of bradykinin (BK)
to enhance vascular permeability may be caused by the increased extravasation of plasma proteinase inhibitors (e.g., $\alpha_1$-proteinase inhibitor, $\alpha_1$-anti-chymotrypsin, and $\alpha_2$-macroglobulin). These mediators control excessive proteolytic activity and thereby protect against connective tissue damage (Kozik et al., 1998).

The hypothalamic–pituitary–adrenal (HPA) axis, which also plays an important role in modulating the inflammatory response (Sterneberg et al., 1989; Sweep et al., 1991; Calogero et al., 1992), is also sexually dimorphic in both animals and humans (Da Silva, 1995). For example, (1) testosterone tends to inhibit HPA axis function, whereas estrogen enhances HPA function (Handa et al., 1994; Suescun et al., 1994), (2) female rats have a higher basal plasma corticosterone level than males (Kitay, 1961; Ehlers et al., 1994; Suescun et al., 1994), and (3) in humans there is a greater HPA axis stress response in females (Peskind et al., 1995). It has been hypothesized that the apparent paradox of a both greater arthritic severity/incidence and higher glucocorticoid levels in females is attributable to females being more dependent on glucocorticoids than males to modulate inflammation (Da Silva, 1995).

In this study we have tested the hypothesis that sex differences in magnitude of a critical component of the inflammatory response (i.e., inflammatory mediator-induced plasma protein extravasation) is dependent on sex steroids and that the effect of female sex steroids is mediated through the sympathoadrenal and/or the HPA axes.

We present evidence that 17$\beta$-estradiol suppresses and testosterone enhances inflammatory mediator-induced plasma extravasation (PE), and that the effect of 17$\beta$-estradiol is sympathoadrenal axis-dependent.

**MATERIALS AND METHODS**

**Animals**

The experiments were performed on weight-matched male and female Sprague Dawley rats (Bantin and Kingman, Fremont, CA, except as described below). Rats were used in PE experiments when they weighed 280–380 gm. The rats were housed in a temperature- and humidity-controlled environment and were maintained on a 12 hr light/dark cycle (lights on at 6 A.M.). Food and water were available ad libitum. Experiments were approved by the UCSF Committee on Animal Research.

**Plasma extravasation**

BK-induced plasma extravasation in the knee joint of the rat was assessed as described previously (Coderre et al., 1989; Green et al., 1991). Rats were anesthetized with sodium pentobarbital (Nembutal, 50 mg/ kg). Skin overlying the knee was excised to expose the joint capsule, and rats were then given an intravenous injection of Evans blue dye (50 mg/ kg, in a volume of 2.5 ml/kg). A 30 gauge hypodermic needle was then inserted into the knee joint cavity for the inflow of perfusion fluid (300 µl/min; controlled by a syringe pump. Sage Instruments model 341B), and after infusion of 100–200 µl of fluid, a second needle (25 gauge) was inserted into the joint cavity for outflow of the perfusion fluid (250 µl/min; syringe pump, Sage Instruments model 351). Samples of perfusion fluid were collected over 5 min intervals for a period of 90 min. Samples were analyzed for Evans blue dye concentration by spectrophotometric measurement of absorbance at 620 nm; absorbance is linearly related to dye concentration (Carr and Wilhelm, 1964). After collection of the first three samples (to establish baseline plasma extravasation levels), BK (150 nm) was added to the perfusing fluid and remained present in the fluid for the duration of the experiment.

**Gonadectomy**

Gonadectomy was performed on male and female rats at 3 weeks of age, i.e., before onset of puberty, and animals were used in plasma extravasation experiments when they weighed 280–380 gm.

**Orchiectomy.** Under brief ether anesthesia, a single cutaneous incision was made along the dorsal midline followed by bilateral incisions through the peritoneum. The ovaries were located, and their vascular bundles were tied off with 4-0 silk suture. Ovaries were then excised, and the cutaneous incision was closed with 5-0 silk suture (Waynforth and Flecknell, 1992).

**Administration of sex steroids**

Chronic administration of sex steroids was performed as described previously (Smith et al., 1977). Briefly, 17$\beta$-estradiol and testosterone were administered via implanted SILASTIC tubes. Segments of SILASTIC tubing (1.67 mm inner diameter × 3.18 mm outer diameter) were used to make hormone implants as follows: testosterone implants consisted of 30-mm-long segments filled with testosterone, and estrogen implants consisted of 10-mm-long segments filled with 17$\beta$-estradiol. Sham implants were prepared of each length and filled with cholesterol. The ends of the implants were capped with wooden plugs and sealed with SILASTIC medical adhesive (Dow Corning, Midland, MI). Implants were washed in absolute ethanol and equilibrated in four changes of PBS over a 24 hr period before placement in the rat. Implants, which were placed subcutaneously at the time of gonadectomy, produce physiological levels of sex steroids (Bridges and Russell, 1981; Bridges, 1984).

**Adrenalectomy**

To remove both adrenal glands, rats were anesthetized with pentobarbital. Incisions in the abdominal wall were made to expose the adrenal glands, which were then excised. Adrenalectomy was performed 1 week before PE experiments. During that week, the rats’ drinking water contained 0.5% sodium chloride and 25 µg/ml corticosterone; water was removed 1 hr before animals were used in PE studies. To confirm that adrenalectomies were complete, immediately before being used in PE studies, corticosterone levels were assessed as described below and shown to be <1 µg/dl.

**Adrenal denervation**

The greater splanchnic nerve innervating the adrenal gland was exposed after a lateral incision was made in the abdominal wall; the adrenal innervation region was isolated close to the adrenal gland and cut, as described previously (Celler and Schramm, 1981). Joint perfusion experiments were carried out at least 7 d after adrenal denervation. Bilateral adrenal denervation did not affect the baseline level of BK-induced plasma extravasation (data not shown). To ensure that adrenal denervation did not interfere with the function of the adrenal cortex, plasma samples were collected immediately after induction of anesthesia for knee joint perfusion experiments and assayed for corticosterone levels as described below.

**Hypophysectomy**

Hypophysectomized Sprague Dawley rats were purchased from Charles River (Hollister, CA). Hypophysectomies were performed after ventral midline incision was made through the mandible to expose the ventral surface of the cranium. Trephines were used to excise a section of cranium and expose the hypophysis, which was then aspirated. For the first week after surgery, animals were given 5% sucrose in their drinking water, and animals were used in PE studies 7–10 d after hypophysectomy. Intact rats from Charles River, which produce the same degree of BK-induced PE as those from Bantin and Kingman (Green et al., 1995), were used as control animals.

**Corticosterone assay**

Blood samples (50–100 µl) were collected from pentobarbital-anesthetized animals by venipuncture (tail vein) immediately before the knee joint perfusion experiment. Samples were immediately centrifuged, and plasma was taken and stored at −20°C until assayed.

Total plasma corticosterone was assayed with a double antibody 125I RIA kit (ICN Biomedicals, Costa Mesa, CA) as described previously (Akana et al., 1985). The assay has a sensitivity of detection of 0.5 µg/ml and has a cross-reactivity reported by ICN Biomedicals as 0.34% for desoxycorticosterone and 0.1% for testosterone.

**Estrogen receptor α immunocytochemistry**

Adrenal glands were excised from anesthetized male and female animals and immersed in Zamboni’s fixative for 4 hr at 25°C. Tissue blocks were...
dehydrated, infiltrated, and embedded in paraffin. Histologic sections were prepared at a thickness of 5 μm and collected on glass slides. The sections were processed for estrogen receptor α immunocytochemistry using the Dako estrogen receptor α-labeled streptavidin–biotin kit (Dako Labs, Carpinteria, CA) following the manufacturer’s instructions. Briefly, slides were deparaffinized, rehydrated, and incubated in Dako Target Retrieval Solution. After rinsing, the slides were treated with 3% aqueous hydrogen peroxide to quench endogenous peroxidase activity, rinsed, and exposed to either 1D5 primary antibody or an isotypically matched mouse IgG1 control reagent. The 1D5 monoclonal antibody is a very well characterized reagent, shown to bind to estrogen receptor α in human endometrial and myometrial cells and normal and hyperplastic mammary epithelial cells, as well as some breast carcinoma epithelia. Similarly, this reagent has been used to identify estrogen receptor α-containing cells in neurons, uteri, and growth plates of rats (Greco et al., 1998; Kennedy et al., 1999). After rinsing, the slides were sequentially exposed to biotinylated anti-mouse immunoglobulins, streptavidin conjugated to horseradish peroxidase, and 3,3'-diaminobenzidine chromogen in imidazole-HCl, containing hydrogen peroxide. Hematoxylin counterstain was not used to avoid obscuring nuclear estrogen receptor reactivity. The slides were rinsed and overslimed in AquaMount. Slides from different experimental groups were reacted at the same time. Because this reagent kit is used by the UCSF diagnostic pathology laboratory, the reagents are tested at least once per week on a human breast tumor with demonstrated estrogen receptor α activity. Slides were viewed with a Nikon Microphot-FXA and photographed with a Sony DCR-5000 digital camera.

Materials
Evans blue dye, bradykinin triacetate, 17β-estradiol, testosterone, dihydrotestosterone, and metyrapone were obtained from Sigma (St. Louis, MO). SILASTIC tubing (Dow Corning, Midland, MI) was obtained from Storz Instrument Company (St. Louis, MO). Nembutal was obtained from Abbott Laboratories (North Chicago, IL). Anti-estrogen receptor α antibody was obtained from Dako Labs. Testosterone and 17β-estradiol were used in crystalline form, metyrapone was dissolved in DMSO, and all other drugs were dissolved in 0.9% saline.

Statistical analysis
Plasma extravasation data were analyzed using repeated-measures ANOVA with one between-subjects factor, treatment, with two levels (control and treated) and one within-subjects factor, time, with 10 levels (20–90 min, 5 min intervals). We present the results of the analysis of the main effect and treatment group. Fisher’s least squares difference test (Fisher, 1949) was used for post hoc comparisons. Differences were considered significant when p < 0.05.

RESULTS
Plasma extravasation
Effects of sex
Perfusion of the knee joint with BK produced a sustained increase in PE that reached a plateau within 25–30 min in both male and female rats. The magnitude of BK-induced PE in female rats was ~50% of that produced in males (p < 0.05) (Fig. 1).

Females
We addressed the question of whether 17β-estradiol decreased, and testosterone increased, BK-induced PE in female rats.

Effects of gonadectomy. To determine the role of female sex steroids in sex differences in BK-induced PE, rats were ovariec- tomized. After ovariectomy, the magnitude of BK-induced PE was significantly higher than that produced in intact females (p < 0.05) (Fig. 2).

Effects of hormone administration. To determine whether the difference in magnitude of BK-induced PE after ovariectomy can be reversed by replacement with a single hormone, we evaluated the effect of chronic exposure to 17β-estradiol in ovariectomized female rats. BK-induced PE in ovariectomized rats with 17β-estradiol implants was significantly lower than after ovariectomy alone and not significantly different from that seen in intact females (p > 0.05) (Fig. 2). In contrast, ovariectomized females implanted with testosterone had a BK-induced PE not signifi- cantly different from that produced by ovariectomy alone.

We next addressed the question of whether female sex steroid effects on BK-induced PE were sympathoadrenal or HPA axis dependent.

Effect of lesions of sympathoadrenal axis. We found that adrenal denervation also resulted in BK-induced PE that was significantly higher than that produced in intact female rats (p < 0.05) (Fig. 3A). Similarly, BK-induced PE in adrenal denervation in ovari- ectomized estrogen-replaced rats was higher than in intact fe-
males \( (p < 0.05) \) (Fig. 3A) and not significantly different from that produced by adrenal denervation alone. Plasma corticosterone levels were not significantly different in adrenal-denervated rats \( (34.5 \pm 4.9 \, \mu g/dl) \) compared with pentobarbital-anesthetized controls \( (39.2 \pm 3.5 \, \mu g/dl) \). Sham surgeries did not affect BK-induced PE (data not shown).

**Effect of lesions of HPA axis.** In female rats, surgical hypophysectomy (to evaluate the pituitary contribution) and metyrapone-induced inhibition of glucocorticoid synthesis (to evaluate the adrenal cortical contribution) did not significantly alter the magnitude of BK-induced PE \( (p > 0.05) \) (Fig. 3B). However, adrenalectomy that interrupts the sympathoadrenal axis as well, significantly increased BK-induced PE \( (p < 0.05) \) (Fig. 3A). Plasma corticosterone levels in all adrenalectomized animals were <1 \( \mu g/dl \). Plasma corticosterone levels were also reduced after hypophysectomy \( (0.9 \pm 0.3 \, \mu g/dl) \) and metyrapone treatment \( (1.2 \pm 0.4 \, \mu g/dl) \) compared with control pentobarbital-anesthetized females \( (39.2 \pm 3.5 \, \mu g/dl) \). Sham adrenalectomies did not affect BK-induced PE (data not shown).

**Males**

We next addressed the question of whether testosterone and dihydrotestosterone raise and 17\( \beta \)-estradiol decreases levels of BK-induced PE in male rats.

**Effects of gonadectomy.** After castration in male rats, BK-induced PE was significantly less than that produced in control males \( (p < 0.05) \) (Fig. 4).

**Effects of hormone administration.** We evaluated the effect of chronic exposure to testosterone and dihydrotestosterone in castrated male rats. BK-induced PE in gonadectomized testosterone-treated and gonadectomized dihydrotestosterone-treated males was not significantly different from that produced in males that were only gonadectomized \( (p > 0.05) \) (Fig. 4). In contrast, gonadectomized males implanted with 17\( \beta \)-estradiol had a BK-induced PE not significantly different from that produced in males that were only gonadectomized.

Finally, we addressed the question of whether male sex steroid effects on BK-induced PE were HPA or sympathoadrenal axis dependent.

**Effect of lesions of sympathoadrenal axis.** Sympathoadrenal axis ablation (adrenal denervation) in males had no effect on the magnitude of BK-induced PE compared with intact males \( (p > 0.05) \) (Fig. 5A). Plasma corticosterone levels after adrenal ablation (adrenal denervation) in males that were only gonadectomized. Adrenal denervation \( (\Delta, n = 14) \) significantly enhanced BK-induced PE compared with control males \( (\bigcirc, n = 13) \). Adrenal denervation also enhanced BK-induced PE in ovariectomized rats receiving 17\( \beta \)-estradiol \( (\wedge, n = 7) \), HPA axis.

**Effect of hormone administration.** Testosterone and dihydrotestosterone raise and 17\( \beta \)-estradiol decreases levels of BK-induced PE in male rats castrated at 21 d of age and chronically receiving either dihydrotestosterone \( (\bigcirc, n = 6) \), DHT \( (\square, n = 8) \), or 17\( \beta \)-estradiol \( (\wedge, n = 7) \) was not significantly different from that produced in weight-matched controls \( (\bigcirc, n = 13) \), but was significantly greater than the magnitude of BK-induced PE produced in castrated \( (\bigcirc, n = 9) \) male rats.

**Estrogen receptor \( \alpha \) immunohistochemistry**

Immunoreactivity for estrogen receptor \( \alpha \) was evident in the cytoplasm of adrenal medullary cells in female rats (Fig. 6). No estrogen receptor \( \alpha \) immunoreactivity was apparent in adrenal cortical cells. No estrogen receptor \( \alpha \) immunoreactivity was apparent in adrenal medullary or cortical cells of male rats (data not shown).

**DISCUSSION**

In this study we have shown that PE produced by the potent inflammatory mediator BK is markedly lower in female compared...
with male rats. This gender difference can be accounted for in part by female and male sex steroids. The contribution of the female sex steroid 17β-estradiol is, at least in part, sympathoadrenal axis dependent.

**Plasma extravasation**

**Influence of sex steroids**

Because reports in the literature have shown that females have greater severity and susceptibility to inflammatory disease (Mackenzie et al., 1979; Wilder et al., 1982; Allen et al., 1983; Deighton et al., 1992; Griffiths et al., 1994; Da Silva, 1995; Homdahl, 1995; Katz and Criswell, 1996), we evaluated the role of the female sex steroid 17β-estradiol in BK-induced PE, an integral component of the inflammatory response. The observations that (1) gonadectomy in female rats resulted in an increase in BK-induced PE, whereas administration of 17β-estradiol to gonadectomized female rats reconstituted the female phenotype, and (2) castration in male rats resulted in a decrease in BK-induced PE, whereas administration of testosterone to castrated male rats reconstituted the male phenotype, strongly argues in favor of a major part of sex differences in this component of the inflammatory response being attributable to these sex steroids. Our work extends previous studies that have shown that sex steroids modulate inflammatory disease severity [e.g., estrogens enhance (Allen et al., 1983; Ansar Ahmed et al., 1985) and testosterone reduces severity of inflammatory disease in animal models (Da Silva et al., 1993a; Harbuz et al., 1995; Booji et al., 1996)] to provide a detailed analysis of the influence of sex and sex steroids on a critical component of the acute inflammatory response, BK-induced PE.

**Mechanisms of sex differences**

Analysis of the mechanism by which a major female sex steroid influences BK-induced PE leads us to hypothesize that the female sex steroid 17β-estradiol produces its effect on the inflammatory response via a sympathoadrenal axis-dependent mechanism, because adrenalectomy and adrenal denervation in female rats results in increased BK-induced PE, and estrogen receptor α immunoreactivity is present in adrenal medullary cells. That sympathoadrenal axis ablation did not affect BK-induced PE in males supports the suggestion that these surgical interventions did not have nonspecific effects on PE.

**Sympathoadrenal axis.** In the present study, we tested the hypothesis that sex steroid effects on BK-induced PE are mediated by an action on the sympathoadrenal axis. Although adrenalectomy in female rats changed the PE response induced by BK to make it like that of male rats, adrenalectomy also ablated the adrenal medulla component of the sympathoadrenal axis. Because the magnitude of BK-induced PE was similar after adrenal denervation and adrenalectomy, we suggest that an adrenal medullary factor(s) in females may be sufficient to account for the sex differences in BK-induced PE observed in this study. Furthermore, because adrenal denervation also blocks the effect of 17β-estradiol implants in gonadectomized females, this further supports the hypothesis that 17β-estradiol's effects are dependent on an intact sympathoadrenal axis. In contrast, the adrenal medulla does not appear to mediate male hormone modulation of BK-induced PE. Finally, the presence of estrogen receptor α immunoreactivity in female adrenal medullary cells supports the suggestion that the female sex steroid estrogen acts directly on the adrenal medulla to induce sympathoadrenal-dependent differences in inflammation.

There is a limited literature describing sex differences in the sympathetic activation of the adrenal medulla (Hinojosa-Laborde et al., 1999). For example, it is known that increases in plasma catecholamine in response to stress is much greater in female rats (Lizey et al., 1985; Taylor et al., 1989; Weinstock et al., 1998) and in women (Frankenhaeuser et al., 1976). Although the site of action of sex steroids on the sympathoadrenal system is unknown, and an action in the CNS cannot be excluded (Hinojosa-Laborde et al., 1999), the presence of estrogen receptor on female adrenal medullary cells provides support for the suggestion that these cells are the target for estrogen's action. Interestingly, the epinephrine and norepinephrine content of adrenal medullary cells varies with estrous cycle in female rats (de Miguel et al., 1989). Although this is the first report that the estrogen receptor is present in adrenomedullary cells, it has recently been reported that estrogen receptor protein and estrogen receptor mRNA are also present in closely related (Anderson, 1993; Lachmund et al., 1994) postganglionic sympathetic neurons (Papka et al., 1997). Despite the fact that the functional role for these receptors is yet to be determined, the observation that they are cytosolic rather than nuclear receptors suggests that they may mediate more rapid
stress to produce plasma levels that are vasoconstrictive (Zukowska-Grojec, 1995), and we have provided evidence that they potentiate inhibit BK-induced PE in the knee joint (Green et al., 1993b). Finally, our results support the suggestion that the sympathoadrenal axis may play an important role in gender differences in stress physiology.

**HPA axis.** Because sexual dimorphism is well established in the HPA axis (Patchev and Almeida, 1998) and the HPA axis regulates inflammation (Da Silva et al., 1993b; Da Silva, 1995; Masi et al., 1996), we tested the hypothesis that the sex difference we observed in BK-induced PE may be mediated, at least in part, by the HPA axis. However, we found that the HPA axis did not appear to play a role in sex differences because hypophysectomy did not affect the magnitude of BK-induced PE in females or males. Thus, sex differences in glucocorticoid physiology are also unlikely to mediate these effects [although estrogen enhances glucocorticoid release (Amin et al., 1980) and BK-induced PE is inhibited by noxious stimulation-induced release of glucocorticoid from the adrenal gland (Green et al., 1997)], because inhibition of corticosteroid synthesis with metyrapone had no effect on the magnitude of BK-induced PE in female or male rats. Interestingly, >90% of rheumatoid arthritis patients do not show abnormality in cortisol levels, suggesting a normally functioning HPA axis (Wilder, 1995); however, it is important to note that there are species differences in the role sex steroids play in the severity of arthritis (Wilder, 1996). Thus, it is still possible that the HPA axis regulates other elements of the inflammatory response in a gender-dependent manner (Da Silva et al., 1993b; Da Silva, 1995; Masi et al., 1996).

Hypophysectomy also decreases plasma levels of sex steroids, which might be expected to influence PE levels; however, it should be noted that PE studies were performed only 7–10 d after hypophysectomy, in contrast to ovariectomies, which were performed ~10 weeks before PE studies. Because of large sex steroid stores in fatty tissue (Deslypere et al., 1985), their continued synthesis in corpus lutea (albeit at a reduced rate) for at least 4 weeks after hypophysectomy (Halling, 1992) and the number of weeks required for altered gene expression to be manifested on removal of estrogens [e.g., rat brain 5-hydroxytryptamine2A receptor densities are decreased 3 months but not 2 weeks after ovariectomy (Cyr et al., 1998)], it is not surprising that we failed to observe an effect on PE 1 week after hypophysectomy.

**Chronic inflammatory disease severity**

Although the etiology of inflammatory diseases such as rheumatoid arthritis are not known, the pathogenetic mechanisms are believed, at least in part, to be immunologically determined; therefore, sex differences in inflammatory response are usually ascribed to an action on the immune system (Ansar Ahmed et al., 1985). In addition, in both female and male rheumatoid arthritis patients, gonadal and adrenal androgens (testosterone and dihydrotestosterone) are suppressed, and androgen administration appears to be protective in chronic inflammatory disease in both animal models and clinical trials (for review, see Lahita, 1996). Rheumatoid arthritis and other rheumatic diseases are more severe in females, yet estrogen produces a decrease in PE. Although this may appear paradoxical, we have shown that mediators that decrease BK-induced PE increase the radiological severity of adjuvant-induced arthritis, and mediators that increase BK-induced PE decrease the radiological severity of adjuvant-induced arthritis (Coderre et al., 1991; Green et al., 1991; Miao et al., 1992b). This has led to the suggestion that PE may be a tissue effects of estrogen, such as release of mediators (Wang et al., 1995; Gu and Moss, 1996), rather than estrogen-induced protein synthesis. The adrenal medulla releases several mediators that may affect the inflammatory response. For example, catecholamines (Coderre et al., 1991), enkephalins (Green and Levine, 1992), galanin (Green et al., 1992), neuropeptide Y (Green et al., 1993a), and corticotropin-releasing hormone (Wei et al., 1993) attenuate inflammatory mediator-induced PE. In particular, catecholamines and neuropeptide Y may be released during...
Protective or repressive component of the synovial inflammatory response (Babbaum and Levine, 1991).

In summary, we have shown that there are sex differences in the PE component of the inflammatory response in rats and that this is dependent on sex steroids in both female and male animals. Importantly, we have shown in female rats a sympathoadrenal axis dependence. Further experiments are required to determine whether estrogen acts at the level of the adrenal medulla cell, which sympathoadrenal mediators contribute to the sex differences in the inflammatory response, and whether the sympathoadrenal axis also contributes to sex differences in chronic inflammatory disease models.

REFERENCES


