Breeding Conditions Induce Rapid and Sequential Growth in Adult Avian Song Control Circuits: A Model of Seasonal Plasticity in the Brain

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In adult songbirds, seasonal changes in photoperiod and circulating testosterone (T) stimulate structural changes within the neural song control circuitry. The mechanisms that control this natural plasticity are poorly understood. To determine how quickly and in what sequence the song nuclei respond to changing daylength and circulating T, we captured 18 adult male white-crowned sparrows and kept them on short days for 12 weeks. We killed five of these birds and exposed the rest to long days (LD) and elevated T. We killed these birds either 7 or 20 d after LD + T exposure. We measured song nuclei volumes and cellular attributes, the mass of the vocal production organ (the syrinx), and song behavior. The neostriatal song control nucleus HVC (also known as “high vocal center”), added 50,000 neurons and increased in size within 7 d of exposure to LD + T. Efferent targets of HVC, the robust nucleus of the archistriatum (RA), and area X of the parolfactory lobe grew more slowly and were not significantly larger until day 20 of the study. The tracheosyringeal portion of the hypoglossal nucleus (nXIIts), which receives projections from RA and normally grows in response to seasonal cues, did not grow over the time course of this study. Syringleal mass increased within 7 d of LD + T treatment. The anatomical changes in the brain were accompanied by behavioral changes in song production. On day 7 when the song circuitry was incompletely developed, male sparrows sang less stereotyped songs than males at day 20 with more completely developed song circuits. These results suggest that the song circuitry responds rapidly and sequentially to breeding-typical conditions (long days and elevated T), and that song stereotypy increases as nuclei within this circuitry grow.

Key words: plasticity; season; songbird; song system; testosterone; white-crowned sparrow

During perinatal ontogeny, brain regions that compose neural circuits develop in a given sequence with a specific time course (e.g., Garey, 1984; Robinson and Dreher, 1990; Blakemore, 1991; Bottjer and Arnold, 1997). Many of these brain regions (or nuclei) continue to exhibit morphological plasticity in adult animals (Nottebohm, 1989; Wade and Crews, 1991; Hofman and Swaab, 1992; Chino, 1995; Woolley, 1998), but little is known regarding the sequence and the timing of plastic changes within and among these regions. A related question is whether the events that underlie adult plasticity match those observed during perinatal development (Kandel and O’Dell, 1992; Bailey and Kandel, 1993). Furthermore, there is scant information about behavioral changes that accompany adult brain changes. The avian song control system offers excellent opportunities to address these fundamental issues.

Song is a learned behavior that plays a critical role in reproduction (Catchpole and Slater, 1995). Song behavior, and the circuits that control it, are seasonally plastic in seasonally breeding species (Nottebohm, 1981; Nottebohm et al., 1986; Kroodsma and Miller, 1996; Smith et al., 1997a; Brenowitz et al., 1998). For example, white-crowned sparrows’ (Zonotrichia leucophrys gambelii) songs are more stereotyped during the breeding season, and several of the song control nuclei can be up to 80% larger in breeding males than in nonbreeding males (Smith et al., 1995, 1997b; Soma et al., 1999). Several song nuclei contain gonadal steroid receptors (Arnold et al., 1976; Gahr, 1990; Balthazart et al., 1992; Brenowitz and Arnold, 1992; Smith et al., 1996; Soma et al., 1999) (Fig. 1), and the cyclical changes in the adult anatomy of these nuclei are mediated primarily by seasonal patterns of photoperiod and testosterone (T) secretion (Nottebohm, 1981; Rasika et al., 1994; Smith, 1996; Bernard et al., 1997; Smith et al., 1997a; Brenowitz et al., 1998; Soma et al., 1999; Tramontin and Brenowitz, 1999).

The song control system is a leading model of anatomical and functional plasticity in the adult brain. The nuclei within the song circuitry are easy to identify in histological sections, and their connectivity is well known (for review, see Brenowitz et al., 1997) (Fig. 1). The natural seasonal plasticity observed in wild songbird brains can be easily reproduced in captive animals by experimentally increasing day length and plasma sex steroid levels (Smith et al., 1995). Much has been learned by manipulating these factors and then analyzing brain structures several weeks or more later (Brenowitz et al., 1991; Bernard and Ball, 1995; Smith et al., 1995, 1997b; Bernard et al., 1997; Tramontin et al., 1998b). This approach, however, misses critical early events of dynamic song system growth. To understand the mechanisms that underlie adult brain plasticity, we must investigate changes as they occur rather than after they are completed. Another limitation of the tradi-
syrinx, vocal production organ; parolfactory lobe. RA, of the hypoglossal nucleus; considerably between individual birds (Wingfield and Farner, 1978; increases slowly over several days, and the timing of this increase varies song nuclei (Tramontin et al., 1998b; Soma et al., 1999). This growth is six males on day 20 (D20). implantation, we killed seven males on day 7 (D7) of the experiment and into the range typical of wild breeding birds (4 –10 ng/ml). After T crystalline T to rapidly increase plasmas concentrations of this hormone would experience on their Alaskan breeding grounds. On day 2, we fully developed in this species within 40 – 45 d of exposure to long days kept on SD indefinitely maintain regressed testes, basal or nondetectable levels of circulating T, and regressed song control nuclei that are typical of considering the song control circuitry as an integrated whole. For example, we do not yet know the sequence of seasonal changes among the various nuclei of the song control circuitry or the degree to which changes in one nucleus may depend on changes in afferent or efferent targets. We have therefore taken a circuit analysis approach to examine the time course over which the song control system responds to long, spring-like days and increased circulating T. MATERIALS AND METHODS Collection of animals and experimental treatments. We collected adult male Gambel’s white-crowned sparrows in eastern Washington during their annual migration in September and October 1997. Eighteen males were housed indoors for 12 weeks on short days (SD = 8 hr light) at 20°C to mimic the photoperiod and temperature on their wintering grounds and to ensure that all birds were sensitive to the stimulatory effects of breeding photoperiod and elevated T. White-crowned sparrows kept on SD indefinitely maintain regressed testes, basal or nondetectable levels of circulating T, and regressed song control nuclei that are typical of the nonbreeding season (Middleton, 1965; Sansum and King, 1976; Smith et al., 1995, 1997c). Food and water were available ad libitum throughout the experiment.

On day 0 (D0) of the experiment, we killed five birds and placed the remaining 13 into two rooms (seven birds in one room and six in another). Previous experiments demonstrated that the song system was fully developed in this species within 40 – 45 d of exposure to long days and T (Smith et al., 1997b). On day 1 of the current experiment, 13 birds were photoshifted to long days (LD = 20 hr light), typical of what they would experience on their Alaskan breeding grounds. On day 2, we implanted these 13 males subcutaneously with a SILASTIC capsule (1.47 mm inner diameter × 1.96 mm outer diameter) containing 12 mm of crystalline T to rapidly increase plasma concentrations of this hormone into the range typical of wild breeding birds (4 –10 ng/ml). After T implantation, we killed seven males on day 7 (D7) of the experiment and six males on day 20 (D20).

White-crowned sparrows exposed to spring environmental conditions in the field or to long days in the laboratory respond with growth of the song nuclei (Tramontin et al., 1998b; Soma et al., 1999). This growth is controlled primarily by the photo-induced increase in circulating T (Smith et al., 1997b). In both the field and the laboratory, circulating T increases slowly over several days, and the timing of this increase varies considerably between individual birds (Wingfield and Farner, 1978; Tramontin et al., 1998b). The minimum concentration of circulating T that is required to induce song system growth is not yet known. These different sources of variability make it difficult to conduct a study of the time course of song system growth in response to gradually changing seasonal cues. We therefore designed our study so that all birds were exposed simultaneously to a photoperiod and T level known to be adequate to induce growth of the song system in this species. Smith et al. (1995, 1997b) showed that exposing white-crowned sparrows to long days and T induced song system growth in captive sparrows that was similar to that observed in wild sparrows (Soma et al., 1999). Exposing all birds to an overnight shift from 8 to 20 hr of light and implanting them with T pellets to produce physiological plasma T levels provided us with a discrete starting point against which to measure the time course of plastic growth in the song system. The results of this study therefore demonstrate the rapidity with which the song circuits can respond to conditions that are sufficient to induce growth during the breeding season.

Hormone assay and measurement of reproductive characteristics. On the morning that each male was killed, we collected 300 μl of whole blood by alar venepuncture into heparinized capillary tubes. The blood was immediately centrifuged, and the plasma was removed and stored at −20°C until assay. We measured plasma T in a single radioimmunoassay using the Coat-A-Count Total Testosterone radioimmunoassay kit (Diagnostic Hormone assay and measurement of reproductive characteristics. Birds were deeply anesthetized by methoxyflurane inhalation and perfused through the heart with heparinized saline followed by 10% neutral-buffered formalin. Brains were post-fixed in NBF for at least 2 weeks, embedded in gelatin, cryoprotected in a 20% sucrose–NBF solution, and sectioned in the coronal plane at 50 μm on a freezing microtome. Every other section was mounted and stained with thionin. The Nissl-defined borders of song nuclei coincide with the borders as defined by other labels (Johnson and Bottjer, 1993a; Raskia et al., 1994; Bernard and Ball, 1995; Smith et al., 1997b; Soma et al., 1999; Tramontin and Brenowitz, 1999).

We projected a magnified (46×) image of each mounted section that contained a song nucleus profile (100 μm sampling interval). We traced onto paper the Nissl-defined borders of three nuclei in the main descending motor pathway of the song control circuitry: HVC (also known as “high vocal center”), the robust nucleus of the archistriatum (RA), and the tracheosyringeal portion of the hypoglossal nucleus (nXIIts) (Fig. 1). We also traced two nuclei of the anterior forebrain circuit: area X of the parolfactory lobe, and the lateral portion of the magnocellular nucleus of the anterior neostriatum (IMAN). These tracings were scanned into a computer, and the cross-sectional area of each song nucleus profile was calculated using NIH Image software (version 1.57; Wayne Rasband, National Institutes of Health, Bethesda, MD). We estimated the volume of each nucleus using the formula for a cone frustum over each measured profile area (Smith et al., 1995). All brain measurements were made blind to the treatment group of each bird.

We expressed song nucleus volume in absolute terms (cubic millimeters) and as a percentage of the entire telencephalon. This second method of expression controlled for differences among groups in overall brain size and/or histological preparation. (Nucleus nXIIts is located in the brainstem and so was only expressed in absolute terms.) We estimated the volume of each nucleus using the formula for a cone frustum over each measured profile area (Smith et al., 1995). All brain measurements were made blind to the treatment group of each bird.

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RESULTS

Plasma testosterone levels and reproductive characteristics

Increased daylength and T implants significantly increased circulating T in our implanted male birds (Table 1). Males killed either on day 7 or day 20 had significantly higher plasma T levels than males killed on day 0. This circulating T was within the physiological range for breeding male sparrows (4–10 ng/ml) and induced significant growth in two peripheral androgen-sensitive tissues. The length of the cloacal protuberance and the mass of the syrinx both increased significantly within 7 d of the onset of treatment (Table 1).

As expected, all males in this study had fully regressed testes that did not grow during the course of the study (Table 1). White-crowned sparrows recrudesce their gonads each year in preparation for breeding, when mean gonadal volume becomes greater than 100 mm$^3$ (Wingfield et al., 1996). This annual gonadal growth is under the control of hypothalamic gonadotropin-releasing hormone, which induces follicle-stimulating hormone secretion from the anterior pituitary. Exogenous T administration inhibits the secretion of gonadotropins through endogenous negative feedback mechanisms (Matt, 1980).

Song system morphometry

The song control system showed significant growth in response to LD + T treatment. The size of the telencephalon also differed slightly, but significantly among treatment groups (Table 2). To control for these group differences in overall telencephalon size, we divided song nucleus volume by telencephalon volume (see Materials and Methods). By expressing song nucleus volume as a percentage of the telencephalon, we confirmed that HVC, RA, and area X grew specifically in response to LD + T (data and statistics in Table 2). The time course of growth, however, was not parallel among these three nuclei. HVC grew more rapidly than either of its two efferent targets, RA and area X (Table 2, Fig. 3). Note in Figure 3 that when expressed as a percentage of their maximum size, HVC was relatively smaller than RA and area X at day 0, but was relatively larger than both of these nuclei at day 7. On day 7, HVC was 94% of its full breeding size, suggesting that most of the growth of HVC was accomplished by this time. On day 20, HVC was fully grown and was not different from HVC size in white-crowned sparrows similarly treated with LD + T for 44 d by Smith et al. (1997b) (t = 1.119; df = 11; p = 0.29).

The efferent targets of HVC, RA, and area X, also grew in response to LD + T, but the growth of these regions was delayed with respect to HVC (Table 2, Fig. 3). Between days 0 and 7, RA did not grow significantly. Between days 7 and 20, however, RA did grow significantly and was fully developed by day 20. On day 20, RA size was the same as that reported for white-crowned sparrows similarly treated with LD + T for 44 d by Smith et al. (1997b) (t = 0.151; df = 11; p = 0.883). Area X also responded to LD + T (Table 2, Fig. 3). Between days 0 and 7, area X did not grow significantly. By day 20, however, area X reached its full breeding size and was similar to area X size in sparrows exposed to LD + T for 44 d by Smith et al. (1997b) (t = 0.499; df = 11; p = 0.63).

Two song nuclei did not show significant growth over the time course of this study. The sizes of IMAN and nXIIIs did not increase at any time in response to LD + T treatment (Table 2).
HVC volume (mm$^3$) 0.554
Cloxin protuberance length (mm) 2.36 ± 0.14
Syinx mass (mg) 12.30 ± 0.49
Gonad volume (mm$^3$) 0.51 ± 0.12

Telencephalon volume (mm$^3$) 596.20
IMAN/Telencephalon (%) 0.024
Area X/Telencephalon (%) 0.243
IMAN volume (mm$^3$) 0.144

*p between days 7 and 20 (D7 vs D20, *p*(D0 vs D7, *p*(LD and killing.

IMAN/Telencephalon (%) 0.037 between days 7 and 20 (D7 vs D20, *p*(D0 vs D7, *p*(LD and killing.

Table 2. Brain morphometry (mean ± SEM)

<table>
<thead>
<tr>
<th></th>
<th>Day 0 (n = 5)</th>
<th>Day 7* (n = 7)</th>
<th>Day 20* (n = 6)</th>
<th>ANOVA F</th>
<th>ANOVA p</th>
</tr>
</thead>
<tbody>
<tr>
<td>HVC volume (mm$^3$)</td>
<td>0.554 ± 0.039</td>
<td>0.937 ± 0.082</td>
<td>1.012 ± 0.121</td>
<td>6.450</td>
<td>0.010</td>
</tr>
<tr>
<td>RA volume (mm$^3$)</td>
<td>0.339 ± 0.040</td>
<td>0.405 ± 0.037</td>
<td>0.494 ± 0.030</td>
<td>4.357</td>
<td>0.032</td>
</tr>
<tr>
<td>Area X volume (mm$^3$)</td>
<td>1.441 ± 0.166</td>
<td>1.734 ± 0.188</td>
<td>2.159 ± 0.204</td>
<td>3.309</td>
<td>0.065</td>
</tr>
<tr>
<td>IMAN volume (mm$^3$)</td>
<td>0.144 ± 0.010</td>
<td>0.144 ± 0.007</td>
<td>0.161 ± 0.024</td>
<td>0.398</td>
<td>0.678</td>
</tr>
<tr>
<td>nXIIIs volume (mm$^3$)</td>
<td>0.074 ± 0.006</td>
<td>0.071 ± 0.006</td>
<td>0.092 ± 0.013</td>
<td>1.621</td>
<td>0.230</td>
</tr>
<tr>
<td>Telencephalon volume (mm$^3$)</td>
<td>596.20 ± 16.35</td>
<td>554.39 ± 13.84</td>
<td>542.50 ± 9.61</td>
<td>3.960</td>
<td>0.042</td>
</tr>
<tr>
<td>HVC/Telencephalon (%)</td>
<td>0.092 ± 0.008</td>
<td>0.167 ± 0.016</td>
<td>0.183 ± 0.019</td>
<td>8.477</td>
<td>0.003</td>
</tr>
<tr>
<td>RA/Telencephalon (%)</td>
<td>0.057 ± 0.006</td>
<td>0.073 ± 0.006</td>
<td>0.091 ± 0.006</td>
<td>7.546</td>
<td>0.005</td>
</tr>
<tr>
<td>Area X/Telencephalon (%)</td>
<td>0.243 ± 0.031</td>
<td>0.314 ± 0.032</td>
<td>0.314 ± 0.032</td>
<td>4.159</td>
<td>0.037</td>
</tr>
<tr>
<td>IMAN/Telencephalon (%)</td>
<td>0.024 ± 0.002</td>
<td>0.026 ± 0.002</td>
<td>0.030 ± 0.005</td>
<td>0.799</td>
<td>0.468</td>
</tr>
</tbody>
</table>

*p Interval between exposure to LD + T and killing.

** Song system neuronal attributes **

To investigate the time course of cellular changes in the song system, we measured neuron size, density, and number in HVC, RA, and nXIIIs. LD + T treatment rapidly increased two of these measures within HVC (Fig. 4). LD + T significantly increased the size of HVC neurons ($F_{(2,17)} = 17.823; p < 0.001$). Within 7 d of LD + T treatment, HVC neurons were 26% larger than at day 0 (D0 vs D7, $p < 0.001$). Neuron size did not increase further between days 7 and 20 (D7 vs D20, $p = 0.55$). The density of HVC neurons did not change in response to LD + T ($F_{(2,17)} = 0.916; p = 0.421$). We have never observed a seasonal change in HVC neuron density in this species (Smith et al., 1995, 1997b; Tramontin et al., 1998a). HVC neuron number, however, was significantly affected by LD + T treatment ($F_{(2,17)} = 5.007; p = 0.022$). Between days 0 and 7, mean HVC neuron number increased by nearly 50,000 neurons (67%) (D0 vs D7, $p = 0.01$). Neuron number did not increase further between days 7 and 20 (D7 vs D20, $p = 0.90$).

LD + T treatment also significantly affected two cellular attributes within RA (Fig. 4). The cross-sectional soma area of RA neurons increased significantly in response to LD + T ($F_{(2,17)} = 9.552; p = 0.002$). Between days 0 and 7, mean RA neuron size increased by 25% (D0 vs D7, $p = 0.01$). This change in cell size occurred despite the fact that the size of RA did not change significantly until after 20 d of LD + T (Table 2, Fig. 3). Neuron size in RA only increased an additional 10% between days 7 and 20 (D7 vs D20, $p = 0.21$). The density of RA neurons was also significantly affected by LD + T treatment ($F_{(2,17)} = 3.959; p = 0.04$). RA neuron density decreased in response to LD + T, but this was not statistically significant until day 20 when neuron density was 24% lower than at day 0 (D0 vs D20, $p = 0.02$). Thus, the time course of the decrease in neuron density was similar to the time course of the overall growth of this nucleus. Neuron number in RA was not significantly affected by LD + T treatment ($F_{(2,17)} = 0.430; p = 0.66$). We have never observed a seasonal change in RA neuron number in this species (Smith et al., 1995, 1997b; Tramontin et al., 1998a).

LD + T treatment increased the size of nXIIIts motoneurons by 39%, but this difference just failed to reach significance (Table 3). We observed no changes in nXIIIts neuron density or number over the time course of this study (Table 3).
Song behavior

We measured song rate and song stereotypy in birds exposed to long days and T for either 7 or 20 d (short-day birds at day 0 never sang and so were not analyzed). Song rate did not differ between males treated with T for 7 or 20 d ($t = 0.319; df = 11; p = 0.38$). Birds at day 7 sang at a mean rate of 1.11 ± 0.41 songs/min, whereas birds at day 20 sang at a mean rate of 0.93 ± 0.33 songs/min.

The absolute values of the 10 song attributes that we measured were not significantly different between groups (Table 4). Song stereotypy, however, did differ between groups. Table 4 shows the mean coefficients of variability for four temporal and six spectral measures of song variability. For every measure, the coefficient of song variability was lower (i.e., singing was more stereotyped) at 20 d after T treatment. For seven of these measures, the difference between groups reached statistical significance (Table 4).

DISCUSSION

Previous studies in songbirds have clearly demonstrated that prolonged exposure to breeding-typical seasonal cues results in dramatic seasonal song system growth (Nottebohm, 1980; Smith et al., 1995, 1997a,b,c). These studies have been pivotal in identifying environmental and physiological factors that influence adult plasticity in the vertebrate brain. They have not, however, examined the dynamic growth of the song control system while it is occurring. This approach is necessary for an understanding of the cellular and molecular events that mediate the seasonal plasticity of the song control system in adult birds. Toward this end, we examined the time course over which the song control circuitry responds to long spring-like days and elevated circulating T.

HVC grew rapidly in response to LD $1$ T. This treatment increased mean HVC volume from 0.55 ± 0.04 (mean ± SEM) to 0.94 ± 0.22 mm$^3$ within 7 d (69% growth). In a different study, Smith et al. (1997b) exposed castrated white-crowned sparrows to LD $1$ T for 44 d and reported a mean HVC volume of 1.18 ± 0.09 mm$^3$, which corresponded to an increase of 80% compared to short-day birds. Smith et al. (1997b) used the same histological procedures as those used in the present study. Taken together, these results suggest that in the present study, HVC was almost entirely recrudesced by day 7. This result is consistent with an anecdotal report of rapid seasonal HVC growth in American tree sparrows (Spizella arborea) (Ball, 1999). To our knowledge, such rapid structural changes of an entire adult brain region have not been reported in any other vertebrate system.

HVC and RA neurons grew significantly within 7 d of LD $1$ T treatment. In RA, neuronal growth preceded the overall growth...
Table 4. Song structure and variability measures (mean ± SEM)

<table>
<thead>
<tr>
<th></th>
<th>Day 7*</th>
<th>Day 20*</th>
<th>T**</th>
<th>p**</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>(n = 5)</td>
<td>(n = 6)</td>
<td></td>
<td></td>
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<tr>
<td>Whole song duration (sec)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mean</td>
<td>1.80 ± 0.07</td>
<td>1.84 ± 0.09</td>
<td>−0.341</td>
<td>0.37</td>
</tr>
<tr>
<td>CV</td>
<td>12.85 ± 2.88</td>
<td>6.35 ± 2.09</td>
<td>1.872</td>
<td>0.05</td>
</tr>
<tr>
<td>Whistle duration (sec)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mean</td>
<td>0.51 ± 0.01</td>
<td>0.52 ± 0.02</td>
<td>−0.569</td>
<td>0.29</td>
</tr>
<tr>
<td>CV</td>
<td>9.65 ± 1.56</td>
<td>5.45 ± 1.24</td>
<td>2.142</td>
<td>0.03</td>
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<tr>
<td>Warble duration (sec)</td>
<td></td>
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</tr>
<tr>
<td>Mean</td>
<td>0.41 ± 0.03</td>
<td>0.39 ± 0.02</td>
<td>0.479</td>
<td>0.32</td>
</tr>
<tr>
<td>CV</td>
<td>5.42 ± 1.26</td>
<td>3.03 ± 0.36</td>
<td>1.987</td>
<td>0.04</td>
</tr>
<tr>
<td>1st buzz duration (sec)</td>
<td></td>
<td></td>
<td></td>
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</tr>
<tr>
<td>Mean</td>
<td>0.24 ± 0.01</td>
<td>0.24 ± 0.02</td>
<td>−0.252</td>
<td>0.41</td>
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<tr>
<td>CV</td>
<td>9.35 ± 2.32</td>
<td>5.93 ± 1.57</td>
<td>1.566</td>
<td>0.08</td>
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<tr>
<td>Whistle minimum frequency (kHz)</td>
<td></td>
<td></td>
<td></td>
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</tr>
<tr>
<td>Mean</td>
<td>3.72 ± 0.28</td>
<td>3.70 ± 0.25</td>
<td>0.065</td>
<td>0.48</td>
</tr>
<tr>
<td>CV</td>
<td>1.91 ± 0.23</td>
<td>1.02 ± 0.17</td>
<td>3.186</td>
<td>0.01</td>
</tr>
<tr>
<td>Whistle maximum frequency (kHz)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mean</td>
<td>4.18 ± 0.30</td>
<td>3.99 ± 0.23</td>
<td>0.505</td>
<td>0.32</td>
</tr>
<tr>
<td>CV</td>
<td>2.59 ± 0.43</td>
<td>1.28 ± 0.12</td>
<td>3.166</td>
<td>0.01</td>
</tr>
<tr>
<td>Warble minimum frequency (kHz)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mean</td>
<td>2.95 ± 0.17</td>
<td>3.15 ± 0.22</td>
<td>−0.706</td>
<td>0.25</td>
</tr>
<tr>
<td>CV</td>
<td>3.24 ± 0.35</td>
<td>2.66 ± 0.30</td>
<td>1.248</td>
<td>0.12</td>
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<tr>
<td>Warble maximum frequency (kHz)</td>
<td></td>
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<td></td>
</tr>
<tr>
<td>Mean</td>
<td>5.12 ± 0.29</td>
<td>5.46 ± 0.31</td>
<td>−0.789</td>
<td>0.23</td>
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<tr>
<td>CV</td>
<td>3.68 ± 0.33</td>
<td>2.16 ± 0.32</td>
<td>3.308</td>
<td>0.01</td>
</tr>
<tr>
<td>1st buzz minimum frequency (kHz)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mean</td>
<td>4.19 ± 0.14</td>
<td>4.23 ± 0.11</td>
<td>−0.190</td>
<td>0.43</td>
</tr>
<tr>
<td>CV</td>
<td>5.12 ± 1.69</td>
<td>2.27 ± 0.27</td>
<td>1.833</td>
<td>0.05</td>
</tr>
<tr>
<td>1st buzz maximum frequency (kHz)</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Mean</td>
<td>6.08 ± 0.17</td>
<td>5.88 ± 0.08</td>
<td>1.142</td>
<td>0.14</td>
</tr>
<tr>
<td>CV</td>
<td>3.60 ± 0.83</td>
<td>2.26 ± 0.34</td>
<td>1.598</td>
<td>0.08</td>
</tr>
</tbody>
</table>

*Interval between exposure to LD + T and killing.
**Student’s unpaired t test (one-tailed). df = 9 for all comparisons.
*CV = SD/mean × 100.

of the entire nucleus. HVC developed too quickly for us to determine if neuronal growth preceded the overall growth in this brain region as well. A time course study with sampling between days 0 and 7 would provide this information. This is a critical question that might provide insight regarding the functional relevance of neuronal growth in HVC. Perhaps larger neurons are better able to provide trophic support to new neurons that are being incorporated into HVC, or to neurons in area X and RA that may depend on trans-synaptic support from HVC neurons.

Neurons in nXIIIts grew 39% larger between days 0 and 20. This increase in neuron size just missed reaching statistical significance (p = 0.059), but may have done so given more time. Between days 0 and 20, the volume of nXIIIts did not increase significantly. Smith et al. (1997b) reported that 44 d of LD + T treatment increased nXIIIts volume by 50% in white-crowned sparrows. Taken together, these observations suggest that the growth of nXIIIts neurons may precede the volumetric growth of the entire nucleus and that the seasonal development of nXIIIts is delayed with respect to HVC and RA. Interestingly, we found that the mass of the syrinx, the efferent target of nXIIIts, increased within 7 d of LD + T. The syringeal muscles contain androgen receptors (Lieberburg and Nottebohm, 1979; Luine et al., 1980; Smith et al., 1996) and so may respond directly to changes in plasma T or its androgenic metabolites. nXIIIts development may depend on anterograde trophic support from RA, retrograde support from the syrinx, or both.

The addition of ~50,000 neurons to HVC within 7 d of LD + T treatment is particularly interesting. Neuronal addition appears to be the primary means by which HVC volume increases before each annual breeding season. In a recent study investigating the regulation of neuronal survival in HVC, Rasika et al. (1999) injected female canaries (Serinus canaria) with the cell proliferation marker [3H]thymidine and then treated the birds with either empty SILASTIC implants or T implants. They reported that 10 d of T treatment increased the survival of [3H]-labeled HVC neurons and that this treatment also increased overall HVC volume by 18%. These results are consistent with the hypothesis that testosterone increases the volume of HVC by rescuing neurons that would otherwise die. Data from our study also support this hypothesis and further suggest that 7 d of LD + T is nearly sufficient for the full development of HVC. Future studies with temporal sampling between days 0 and 7 will provide valuable insights into the regulation of neuronal proliferation and survival in the adult brain.

HVC sends axonal projections to two efferent targets in the song control system (Fig. 1). These two targets, RA and area X,
responded more slowly than HVC to LD + T treatment (Fig. 3). The delayed growth in RA and area X did not appear to be fully accomplished until day 20 when the sizes of both nuclei were similar to those reported by Smith et al. (1997b) in sparrows exposed to LD + T for 44 d. The delayed growth of RA and area X is consistent with two alternative hypotheses. These two nuclei may simply take longer to either initiate and/or complete growth in response to LD + T. Alternatively, the plastic growth of these two nuclei may depend on afferent input from HVC. This second hypothesis seems especially likely in the case of area X, which does not express gonadal steroid receptors and presumably cannot respond directly to increased circulating T (Gahr and Metzdorf, 1997) (Fig. 1). Preliminary data from our laboratory suggest that afferent input from HVC is required for seasonal growth of RA as well as area X. In male white-crowned sparrows with unilateral HVC lesions, ipsilateral RA and area X did not grow in response to LD + T treatment. Contralateral to the HVC lesion, however, the growth of RA and area X occurred normally (Brenowitz and Lent, 1999). Trans-synaptic factors from HVC might be sufficient to induce seasonal development of RA and area X (Akutagawa and Konishi, 1998; Rasika et al., 1999), or in the case of RA may facilitate the action of T within the nucleus itself.

Early ontogeny and adult plasticity of the song control circuits may exploit similar mechanisms. In young zebra finches (Poephila guttata; between 12 and 53 d after hatching), the volumetric growth of HVC appears to precede that of RA and area X (Bottjer et al., 1985). This developmental sequence within the juvenile song circuitry closely parallels the sequence of growth in the adult brain in response to conditions typical of the breeding season (i.e., LD + T). Also, lesions of HVC in juvenile zebra finches block the full development of RA and area X (Herrmann and Arnold, 1991; Johnson and Bottjer, 1993b; Akutagawa and Konishi, 1994). As stated above, unilateral lesions of HVC in adult Gambel’s white-crowned sparrows prevent LD + T-induced growth of RA and area X ipsilateral to the lesion (Brenowitz and Lent, 1999). These observations suggest that it will be productive for future studies to explore the extent to which plasticity of adult neural circuits is regulated by cellular and molecular mechanisms similar to those that regulate plasticity during early development.

Song behavior
The functional relevance of the seasonal plasticity in the song system remains a critical but elusive problem. In wild, freely ranging white-crowned sparrows, the seasonal anatomical changes in the song control system and the seasonal growth of the syrinx are both correlated with seasonal changes in song stereotypy and song structure (Smith et al., 1995; Brenowitz et al., 1998; A. Tramontin, N. Perfito, J. Wingfield, and E. Brenowitz, unpublished data). During the breeding season when the song system and the syrinx are fully developed, the temporal and spectral structure of males’ songs are highly stereotyped. During the nonbreeding season when the song system and the syrinx are fully regressed, males’ songs are more variable in structure. In the present study, we were unable to measure song stereotypy in males maintained on short days because they did not sing. We were, however, able to measure song stereotypy in males at day 7 that had fully developed syringes, but only partially developed song control circuitry, and males at day 20 with large syringes and more completely developed song control systems. We found no differences in song production rate between the two groups, but males killed after 7 d of LD + T sang less stereotyped songs than males at day 20. These data indicate that there is not a simple relationship between song rate and the developmental state of the song circuitry. These data do suggest, however, that a large HVC and/or a large vocal production organ are not sufficient for stereotyped song production. It seems reasonable to hypothesize that a fully developed song control circuit is required to produce stereotyped songs, but not to sing at a high rate. A direct causal relationship between neural and behavioral changes, however, has yet to be established (for an extensive discussion of this issue, see Smith et al., 1997a).

Conclusion
There are other vertebrate models in which entire brain regions undergo extensive seasonal structural changes (Wade and Crewe, 1991; Hofman and Swaab, 1992; Barnea and Nottebohm, 1994; Galea et al., 1999). It will be interesting to determine whether our observations of sequential growth in the adult song circuitry apply to these systems as well. It is known that plastic anatomical changes in the adult brain can occur rapidly (e.g., Woolley and McEwen, 1992). To what degree do these changes depend on afferent or efferent support? What factors are required for such putative trans-synaptic support? Is cellular growth a first step in changing the size of adult brain regions in other model systems? These questions should be addressed in a wide variety of neural circuits that show adult plasticity.

REFERENCES


