Developmental Changes in the Subcellular Localization of Calretinin

Nicola J. Hack, Mark C. Wride, Kathleen M. Charters, Stanley B. Kater, and Thomas N. Parks

Department of Neurobiology and Anatomy, University of Utah School of Medicine, Salt Lake City, Utah 84132

Brainstem auditory neurons in the chick nucleus magnocellularis (NM) express high levels of the neuron-specific calcium-binding protein calretinin (CR). CR has heretofore been considered a diffusible calcium buffer that is dispersed uniformly throughout the cytosol. Using high-resolution confocal microscopy and complementary biochemical analyses, we have found that during the development of NM neurons, CR changes from being expressed diffusely at low concentrations to being highly concentrated beneath the plasma membrane. This shift in CR localization occurs at the same time as the onset of spontaneous activity, synaptic transmission, and synapse refinement in NM. In the chick brainstem auditory pathway, this subcellular localization appears to occur only in NM neurons and only with respect to CR, because calmodulin remains diffusely expressed in NM. Biochemical analyses show the association of calretinin with the membrane is detergent-soluble and calcium-independent. Because these are highly active neurons with a large number of $Ca^{2+}$-permeable synaptic AMPA receptors, we hypothesize that localization of CR beneath the plasma membrane is an adaptation to spatially restrict the calcium influxes.

Key words: calcium-binding proteins; auditory neurons; calcium homeostasis; calretinin; subcellular localization; nucleus magnocellularis

One of the most characteristic features of neurons is the degree to which specific functions are localized. The subcellular location of particular molecules within a neuron determines the contributions that these molecules can make to the overall function of the cell (Rosenmund et al., 1994; Steward, 1997; Ziff, 1997). For example, the precise localization of calcium channels at the presynaptic terminal determines many aspects of synaptic function, because calcium levels locally govern exocytic neurotransmitter release (Augustine et al., 1987). Because other proteins also contribute to the regulation of calcium ion concentration, one might predict that such proteins, rather than being uniformly dispersed within a cell, might also be spatially localized. The present study focused on one such protein, the calcium-binding protein calretinin, in a class of auditory neurons that face unique calcium challenges.

Brainstem auditory neurons preserve the relative timing of action potentials transmitted through successive synaptic levels by means of unique morphological and functional adaptations in axons, dendrites, and synapses. These adaptations include rapidly desensitizing, calcium-permeable AMPA receptors, a strong $K^+$ conductance to repolarize the cell rapidly, and large calycine axosomatic endings to facilitate reliable transmission (Oertel, 1999; Trussell, 1999). Because auditory neurons have some of the highest average and maximal discharge rates in the CNS, the presence of $Ca^{2+}$-permeable synaptic receptors subjects them to high calcium influxes and requires them to maintain a variety of mechanisms for maintaining calcium homeostasis (Zirpel et al., 1998). Calcium-binding proteins (CBPs) have been proposed to be particularly important for such calcium regulation because they are abundantly expressed in auditory neurons of both mammals and birds (Takahashi et al., 1987; Rogers, 1989; Parks et al., 1997). The cochlear nucleus magnocellularis (NM) in the chick is composed of neurons with all of the characteristic specializations of brainstem auditory neurons and whose anatomy, physiology, and development have been well characterized (Rubel and Parks, 1988; Oertel, 1999; Trussell, 1999).

Nucleus magnocellularis neurons express a single CBP, calretinin, and not the closely related E-F hand CBPs parvalbumin and calbindin (Parks et al., 1997). Calretinin, like parvalbumin and calbindin, has widely been thought to be a soluble protein that resides primarily in the cytosol, as a soluble buffer (Baimbridge et al., 1992; Andressen et al., 1993). We present evidence here that, during the development of NM neurons, calretinin shifts from being uniformly distributed throughout the cell body to being concentrated in a shell underlying the cell membrane. This change in localization coincides in developmental time with the onset of spontaneous electrical activity in the cochlea (Lippe, 1994), the onset of synaptic transmission from the cochlear nerve.
to NM neurons, and the condensation of cochlear nerve axons into large calycine axosomatic endings (Rubel and Parks, 1988). We propose that the change in calretinin localization allows NM neurons to function effectively under conditions of high cytoplasmic calcium load.

**MATERIALS AND METHODS**

**Animals.** Fertilized White Leghorn chicken eggs were obtained from a local supplier and incubated in forced draft incubators.

**Immunohistochemistry and confocal microscopy.** Embryonic chicks were anesthetized by intraperitoneal injection of sodium pentobarbital and then perfused transcardially with 0.9% saline followed by cold 4% paraformaldehyde (in 0.1 M phosphate buffer, pH 7.4). Brains were removed, post-fixed (4°C) for 2–4 hr, and placed in 30% sucrose in 0.1 M phosphate buffer for 24 hr. Embryos were staged according to the series of Hamburger and Hamilton (1951). Eighteen-micrometer sections were cut on a cryostat and mounted on gelatin-coated slides for immunohistochemistry. Calretinin immunoreactivity was detected with rabbit polyclonal antisera against human calretinin (Schwaller et al., 1993; SWant, Bellinzona, Switzerland) by using a tetramethylrhodamine isothiocyanate-tagged secondary antibody (Jackson ImmunoResearch, West Grove, PA). Because it has been reported that antibody recognition of calretinin can depend on its calcium binding status, some sections were stained with a calcium-insensitive antibody to calretinin (Winsky and Kuznicki, 1996). We saw no difference in staining pattern between the two antibodies. Sections were viewed on a BioRad (Hercules, CA) 1024 confocal microscope with a 60× oil objective. Optical sections every 2 μm through the thickness of a cell were captured at fixed camera gain, pinhole size, and laser intensity.

**Quantitative analysis of subcellular distribution of calretinin.** Analysis of fluorescent intensity was performed on a Macintosh computer using the public domain NIH Image program (developed at the US National Institutes of Health and available on the Internet at http://rsb.info.nih.gov/nih-image/). Fluorescent intensities of a defined area (2 × 2 pixels) were measured at four locations at the perimeter of each cell and two locations within each cell avoiding the nucleus. The average intensity at the perimeter and the center of the cell was calculated and expressed as a ratio (Iperimeter/Icenter). Three animals were analyzed at each embryonic age: embryonic day 10 (E10; 27 cells), E13 (53 cells), E15 (58 cells), and E18 (55 cells).

**Dissociation of nucleus magnocellularis neurons.** NMs dissected from chick brainstem were placed in papain (Worthington Biochemicals, Lakewood, NJ; 40 U/ml) for 30 min at room temperature. Using a fire-polished Pasteur pipette, NMs were triturated, and single cells were plated on poly-L-lysine-coated glass-bottomed dishes in MEM containing 10% fetal bovine serum (HyClone, Logan, UT). Four hours after plating, cells were fixed with 4% paraformaldehyde in 0.1 M phosphate buffer, pH 7.4, containing 0.12 m sucrose. Cells were immunostained for calretinin and viewed under a confocal microscope.

**Immunoblot analysis.** NMs were dissected from chickens at various embryonic ages and homogenized in a harvesting buffer containing 62.5 mM Tris-HCl, pH 6.9, 2.5% SDS, 10% glycerol, and 2 mM EDTA. Seventy micrograms of total protein (estimated by the BCA method) were subjected to SDS-PAGE according to the method of Laemmli (1970). One hundred nanograms of calretinin (in vitro translated and purified using a glutathione S-transferase purification system; Pharmacia Biotech, Uppsala, Sweden) was run as an internal control. Proteins were transferred to nitrocellulose membranes, and immunoblotting was performed in the presence of 0.5% nonfat dry milk and 0.1% Tween 20 in 0.01 M phosphate buffer, pH 7.5. Antisera against calretinin (SWant) or β-actin (Sigma, St. Louis, MO) were used at 1:10,000 or 1:5,000, respectively. Detection of bound antibody was done using an ECL detection system (Amersham, Piscataway, NJ).

**Preparation of subcellular fractions.** Subcellular fractions of chick brainstem were obtained by differential centrifugation. Chick brainstem pieces containing the nucleus magnocellularis were harvested in 5 mM HEPES, pH 7.4, containing 0.32 M sucrose, 0.2 mM calcium chloride, and a mixture of protease inhibitors (Sigma). Tissue was homogenized on ice with a Teflon pestle (20 strokes) followed by a glass pestle (40 strokes). The nuclear fraction was removed by a low-speed spin at 1000 × g for 10 min. The resultant supernatant was centrifuged at 9800 × g for 20 min to obtain a crude membrane pellet and a cytosolic supernatant. The pellet was washed by resuspension in 5 mM HEPES, pH 7.4, containing 1.5 M sucrose to lyse any unbroken cells, followed by centrifugation to remove any remaining cytosolic components. Aliquots of membrane and cytosolic fractions were subjected to SDS-PAGE and then immunoblotted (as above). To characterize the solubilization properties of calretinin, the membrane pellet was incubated in 2% Triton X-100 for 10 min at 4°C and recentrifuged. The resultant supernatant and pellet were subjected to SDS-PAGE and immunoblotted for calretinin.

**RESULTS**

We have shown previously that both calretinin transcript and immunoreactivity increase in chick NM neurons during development (Parks et al., 1997). The previous study provided us with important temporal information regarding calretinin expression in the auditory system, but the immunohistochemical techniques used to detect calretinin were not optimal for providing quantitative information. Here, we used a biochemical approach to obtain a measure of the relative concentration of calretinin in NM at the same developmental stages used previously. Western blot analysis of NM proteins isolated from chicks at embryonic days 10, 13, 15, and 18 clearly showed an increase in calretinin concentration with age (Fig. 1). Calretinin expression was barely detectable at E10 but increased dramatically by E13 and continued to increase until E18. To perform single-cell analysis and determine the subcellular location of calretinin, we used immunofluorescence labeling of calretinin in NM slices, together with confocal microscopy. To our surprise, the subcellular location of calretinin within NM neurons changed as a function of developmental age. At E10, calretinin was expressed at low levels, in agreement with our biochemical analysis, and diffusely throughout the NM neuron, in agreement with other reports on calretinin immunohistochemistry (Rogers, 1989). By E15, however, calretinin was less homogeneous in its subcellular distribution, and an accumulation of immunofluorescence was observed underneath the plasma membrane in 86% of the cells examined (Fig. 2A). This submembrane distribution was even more pronounced at E18 when the majority of the cells (91%) contained high levels of immunofluorescence in the form of a distinctive shell under the plasmamembrane. At posthatching day 2, the animals showed the same distribution of calretinin (results not shown), showing that the effect persists into postnatal life. Using the confocal microscope to obtain thin optical sections throughout the entire thickness of each NM neuron, we saw the characteristic ring-like distribution of calretinin in all optical sections, with the exception of the top and bottom sections that contain areas directly underneath the plasma membrane. To rule out staining artifacts and to determine whether the observed localization of calretinin was...
concentrated at the plasma membrane at E18. The distribution of calretinin was performed as described in Materials and Methods. These values were expressed as a ratio, $I_{\text{perimeter}}/I_{\text{center}}$, and the ratio values were plotted as a cumulative frequency histogram for the four developmental ages (Fig. 2C). The frequency histogram clearly shows a difference in calretinin distribution between the cell populations of E10–E13 chicks and E15–E18 chicks. As predicted for a uniform homogeneous distribution of calretinin, 54% of the E13 cells displayed a ratio of 1, whereas >90% of the E18 cells had $I_{\text{perimeter}}/I_{\text{center}}$ of >1; these ratios were reliably different ($p < 0.01$).

During development, NM neurons increase in volume together with their subcellular organelles (Fig. 2; Parks, 1979). To exclude the possibility that the change in localization may be attributable to a physical restriction of cytoplasm and cytosolic proteins to the periphery of the cell, we stained NM neurons with an antibody against the cytosolic protein calmodulin. In contrast to calretinin, confocal analysis showed that calmodulin immunoreactivity was homogeneously distributed throughout the neuron (Fig. 3a,b) with the exception of the nucleus. To examine whether calretinin was distributed similarly in other neurons, we examined two avian brainstem auditory nuclei, nucleus angularis (NA) and nucleus laminaris (NL), which are known to express calretinin (Parks et al., 1997). Neurons in both nuclei indeed stained for calretinin, but in contrast to NM neurons, calretinin was homogeneously distributed throughout NA and NL cells (Fig. 3c,d).

To confirm the observations obtained with the confocal microscope, we prepared subcellular fractions of chick brainstem by differential centrifugation (see Materials and Methods). Western blot analysis demonstrated that calretinin immunoreactivity was associated with both cytosolic and particulate fractions (Fig. 4A). Calretinin immunoreactivity in the particulate fraction resisted several washes, suggesting that the particulate localization did not reflect contamination from the cytosolic fraction. Densitometric measurements of band intensities using the NIH Image analysis program determined that 60 ± 7% ($n = 5$ independent experiments) of calretinin immunoreactivity was associated with the particulate fraction. This estimation is likely to be an underestimate because, due to the small size of NM, our fractionation procedure required the use of whole brainstem pieces containing not only NM but also NL and NA. Confocal analysis of NL and NA neurons revealed homogeneous staining of calretinin, which implies that it would contribute only to our cytosolic fraction.

Previous studies on rat cerebellum estimated that no more than 10% of calretinin was membrane-associated (Winsky and Kuznicki, 1995). Therefore, we made cytosolic and membrane fractions from E18 chick cerebellum and ran the fractions under the same conditions as the brainstem and, in agreement with Winsky and Kuznicki (1995), we found that cerebellum calretinin is predominantly cytosolic (Fig. 4A). To examine the nature of the membrane association of brainstem calretinin, we manipu-
DISCUSSION

We have shown that during development of NM neurons, calretinin changes from being uniformly distributed and at low concentrations to being highly concentrated under the plasma membrane. Previous biochemical estimates indicated that in cerebellum (Winsky and Kuznicki, 1995) and whole brain (Hubbard and McHugh, 1995), <10 and 27%, respectively, of calretinin is associated with the particulate fraction. Our biochemical data show that 60% of total brainstem calretinin is localized to the particulate fraction (and significantly >60% of NM calretinin). The functional consequences of a CBP being localized to the plasma membrane, as opposed to freely mobile within the cytosol, are considerable. The action of calcium ions in the cytosol is constrained by many effectors of calcium buffering and removal, including CBPs (Miller, 1991). The spatial localization of these effectors will, in turn, affect the behavior, both temporally and spatially, of a calcium signal within a neuron (see below).

Other CBPs have been found to be localized under the plasma membrane (calcineurin; Lukyanetz, 1997) or directly associated with the plasma membrane [S100 (Donato et al., 1989), recoverin (Dizhoor et al., 1993), and visinin-like protein (VILIP) (Lenz et al., 1996)]. Unlike calretinin, in some cases this interaction is dependent on the cytoskeleton or on intracellular calcium levels [recoverin (Dizhoor et al., 1993), VILIP (Lenz et al., 1996), caldendrin (Seidenbecher et al., 1998), and neuronal calcium sensor-1 (Martone et al., 1999)]. Although functional significance of these associations for calcium signaling has not been directly tested, the theoretical implications are important. During normal physiology NM neurons experience unusually high calcium influxes directly at the cell soma (Trussell, 1999). Because the estimated $K_d$ of calretinin for calcium is 380 nM (Stevens and Rogers, 1997), most of the calretinin should be in an unbound state at resting calcium levels, enabling it to function during periods of elevated calcium. In addition, localizing calretinin to underneath the site of high calcium influx would presumably attenuate global rises in cytosolic calcium because of its ability to bind calcium. Because our biochemical analysis suggests that calretinin is associated with the membrane and therefore likely immobile, one would predict that calretinin would act to lower the diffusion of calcium ions through the cytosol (Gabso et al., 1997). Although calretinin is clearly not the only effector of calcium buffering and removal in NM neurons, its position under the membrane places it spatially to interact with membrane-associated calcium extrusion pumps. Conversely, calcium ions released from intracellular sites would be relatively unaffected by the high concentration of calretinin at the membrane.

Although calretinin expression levels are not influenced by removal of afferent input or the onset of synaptogenesis (Parks et al., 1997; Kubke et al., 1999) the developmental period over which calretinin becomes localized correlates conspicuously with the formation of the mature calycine synapse and the onset of mature...
activity patterns (Rubel and Parks, 1988; Lippe, 1994). It is intriguing to speculate that the developmentally regulated localization of calretinin is an activity-dependent phenomenon not dissimilar to the activity regulated positioning of other proteins (Tongiorgi et al., 1997; Craig, 1998; Steward et al., 1998). Alternatively, localization of calretinin may equally be influenced by the formation of the presynaptic calyx or expression of postsynaptic AMPA receptors.

The message of the present investigation is that calretinin can be, at a given time in development, highly localized. We hypothesize that localization of calretinin beneath the plasma membrane is an adaptation of NM neurons to spatially restrict calcium influxes. Given the tools at hand and the potentially large functional consequences, it may be important to reevaluate the spatial disposition of calcium-binding proteins in other regions of the nervous system during development.

REFERENCES