

The C Domain of Netrin UNC-6 Silences Calcium/Calmodulin-Dependent Protein Kinase- and Diacylglycerol-Dependent Axon Branching in *Caenorhabditis elegans*

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Second messenger systems mediate neuronal responses to extracellular factors that elicit axon branching, turning, and guidance. We found that mutations in *Caenorhabditis elegans* that affect components of second messenger systems, a G-protein subunit, phospholipase C β , diacylglycerol (DAG) kinase, and calcium/calmodulin-dependent protein kinase (CaMKII), have no obvious effect on axon responses to UNC-6 except in animals in which the N-terminal fragment, UNC-6 Δ C, is expressed. In these animals, the mutations enhance or suppress ectopic branching of certain axons. Netrin UNC-6 is an extracellular protein that guides circumferential migrations, and

UNC-6 Δ C has UNC-6 guidance activity. We propose that the guidance response elicited by the UNC-6 N-terminal domains involves mechanisms that can induce branching that is sensitive to CaMKII- and DAG-dependent signaling, and that the UNC-6 C domain is required in *cis* to the N-terminal domains to silence the branching and to maintain proper axon morphology.

Key words: *netrin*; *UNC-6*; *Caenorhabditis elegans*; *guidance*; *axon branching*; *genetics*; *G-protein subunit*; *Gq α* ; *phospholipase C β* ; *PLC β* ; *diacylglycerol kinase*; *DAK*; *calcium/calmodulin-dependent protein kinase*; *CaMKII*; *neuropeptide Y receptor*

The regulation of axon guidance and branching is critical for the proper development of the nervous system. Recent studies suggest that guidance and branching share common mechanisms (Brose et al., 1999; Lim et al., 1999; Kalil et al., 2000). For example, fragments of proteins known to mediate axon guidance can promote axon branching. In *Caenorhabditis elegans*, netrin UNC-6 guides circumferential migrations, and the expression of an N-terminal fragment has been shown to cause additional axon branches from ventral cord motor neurons (Lim et al., 1999). In vertebrates, Slit2 has been implicated in axon guidance and the N-terminal fragment has been shown to promote axon elongation and branching in an *in vitro* collagen assay system (Wang et al., 1999). Aside from such protein fragments, there are a number of other secreted factors, such as neurotrophins, that have been shown to influence axon guidance and promote branching (for review, see Acebes and Ferrus, 2000; Kalil et al., 2000). Although such factors have been identified, little is known about the underlying mechanisms by which such molecules dictate axon morphology.

The netrins are a family of extracellular guidance proteins that can function *in vivo* to attract and repel axons from sources that secrete the molecule (Hedgecock et al., 1990; Ishii et al., 1992; Serafini et al., 1994; Colamarino and Tessier-Lavigne, 1995). In *C. elegans*, netrin UNC-6 guides circumferential cell and axon

migrations (Hedgecock et al., 1990; Ishii et al., 1992; Wadsworth et al., 1996). Axons that express the UNC-5 and UNC-40 netrin receptors migrate away from UNC-6 sources; axons that express UNC-40 migrate toward UNC-6 sources. UNC-6 is expressed in changing patterns by 12 types of cells and is predicted to create a stable global cue peaking near the ventral midline and to create local cues on cell surfaces (Wadsworth and Hedgecock, 1996; Wadsworth et al., 1996).

UNC-6 was the first characterized member of the netrin family, and residues 1–437 were designated domains VI, V-1, V-2, and V-3 based on the similarity of the domains to the N-terminal domains of laminin subunits (Ishii et al., 1992). Residues 438–591 were designated domain C, and it was observed that the same motif is found in C3, C4, and C5 complement proteins but not in a paralogous protein, α_2 macroglobulin (Ishii et al., 1992). Recently, a modified domain C (C') was found in a functionally divergent form of vertebrate netrin designated netrin-G₁ (Nakashiba et al., 2000). These observations indicate that UNC-6 C is a conserved structural module; they suggest that UNC-6 C has a biological function.

We have shown previously that in *unc-6 Δ C* transgenic animals, which express UNC-6 without UNC-6 C, the ventral nerve cord motor neurons extend additional processes circumferentially (see Fig. 3B) (Lim et al., 1999). This activity requires the netrin receptors UNC-5 and UNC-40 (Lim et al., 1999). The expression of UNC-6 Δ C provides a means to explore the mechanisms by which axons respond to secreted factors. We have examined the morphology of neurons in *unc-6 Δ C* animals and have uncovered mutations that modulate these morphologies. From these results, we are able to make predictions regarding the mechanisms by which morphological changes to axons might be controlled.

MATERIALS AND METHODS

Strains. The following mutations and strains were used for mapping and double-mutant constructions: N2; RW7000; IM145 *url577*[IM#183

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IM#175 pRF4] II; IM222 *npr-1(ur89)*; CX4056 *npr-1(ad609)lon-2(e678)* X; CX3048 *npr-1(ky13)* X; DA658 *npr-1(n1353)lon-2(e678)* X; and MT1434 *egl-30(n686)* I. The following strains were generated in this study: IM342 *lon-2(e678)unc-6(ev400)* X; IM290 *npr-1(ur89)lon-2(e678)* X; IM337 *npr-1(ky13)lon-2(e678)* X; IM234 *lon-2(e678);unc-6ΔC(urIs77)*; IM289 *npr-1(ur89)lon-2(e678);unc-6ΔC(urIs77)*; IM288 *npr-1(ad609)lon-2(e678);unc-6ΔC(urIs77)*; IM341 *npr-1(ky13)lon-2(e678);unc-6ΔC(urIs77)*; IM235 *npr-1(n1353)lon-2(e678);unc-6ΔC(urIs77)*; IM338 *egl-30(n686);npr-1(ur89)lon-2(e678)*; IM339 *egl-30(n686);npr-1(ad609)lon-2(e678)*; IM340 *egl-30(n686);unc-6ΔC(urIs77)*; IM357 *urIs193 [npr-1::npr-1::GFP]*; IM403 *unc-6ΔC(urIs77);npr-1(ky13);urIs231 [unc-129p::npr-1(ur89)::gfp]*; IM405 *unc-129(ev554);npr-1(ur89)lon-2(e678)*; IM406 *unc-129(ev554);npr-1(ad609)lon-2(e678)*; and IM409 *unc-129(ev554);npr-1(ky13)*.

Genetic screen. A second filial generation (F2) screen was performed using ethylmethanesulfonate as a mutagen. The starting strain, IM145, carries a transgene, *urIs77*, which encodes hemagglutinin (HA) epitope-tagged UNC-6 that does not include domain C (Lim et al., 1999). The *urIs77* transgene was made by removing the sequence that encodes for domain C from an *unc-6::HA* clone that had been shown to rescue all mutant phenotypes in the *unc-6* null genetic background (Wadsworth et al., 1996). In *unc-6* null mutants, expression of *urIs77* exhibits rescue of the *unc-6(-)* uncoordinated behavior. In *unc-6(+)* animals, expression causes a slightly uncoordinated phenotype. The transgene also expresses green fluorescent protein (GFP) throughout the nervous system and confers a rolling phenotype to the animals (Lim et al., 1999). F2 animals were screened for wild-type movement; the selected mutants were examined for wild-type axon branching by epifluorescence microscopy. The isolated mutant strains were outcrossed against wild-type N2 six times to remove other possible mutations. Expression of the transgene was confirmed by the rolling phenotype as well as by GFP expression; the presence of the UNC-6 protein was further confirmed by Western blot analysis using monoclonal antibody 12CA5 (Boehringer Mannheim, Mannheim, Germany) to detect the HA epitope.

***npr-1* analysis.** *ur89* was mapped to the *npr-1* region. From the cross *ur89 × lon-2(e678)unc-6(ev400)* X, 1/50 Lon non-Unc recombinants were clumping. Noncomplementation between *ur89* and other *npr-1* alleles was tested by scoring for clumping of *trans* heterozygotes (non-Lon progeny) from a cross such as *ur89 × n1353 lon-2*. To characterize *npr-1(ur89)* animals, the allele was linked with *lon-2(e678)* on linkage group X by crossing *ur89 × lon-2(e678)unc-6(ev400)* X, 1/50 Lon non-unc recombinants segregated clumping of *npr-1(ur89) lon-2(e678)*. Transgenic *npr-1* strains were generated by standard methods (Mello and Fire, 1995). *npr-1(ur89)* animals were injected with pM4, a plasmid that contains an insert of *npr-1* genomic DNA derived from the solitary strain N2 (de Bono and Bargmann, 1998). The *npr-1* insert contains a 7.4 kb promoter region and 2.3 kb of the coding region. pM4 was coinjected with the markers pRF4, which expresses *rol-6(su1006)* and causes a dominant rolling phenotype, and pPD118.33, which expresses GFP under the *myo-2* promoter in pharyngeal muscles (Mello and Fire, 1995). Four independently derived strains were found to rescue the *ur89* phenotype in the clumping assay. To test whether the results of the assay were affected by the rolling phenotype conferred by pRF4, the assay was repeated using animals that also carried the *dpy-11(e224)* allele, which suppresses the *rol-6(su1006)* rolling phenotype. *npr-1(ur89);dpy-11(e224)* males were crossed with *npr-1(ur89);urEx162[pM4,pRF4,pPD118.33]* hermaphrodites; *ur89/ur89;dpy-11/+* progeny that carried the *urEx162* transgene were chosen. Dpy non-Rol F2 progeny with pharyngeal-muscle GFP expression were selected and the assay was repeated. There was no significant difference in the assay results. To identify the molecular lesion in *npr-1(ur89)*, PCR fragments including the entire coding sequence and intron region were amplified from the genomic DNA of *npr-1(ur89)* animals. PCR fragments were cloned into a pBluescript SK+ vector (Stratagene, La Jolla, CA), and both PCR fragments and the subclones were sequenced by an automated sequencer. The mutation was confirmed by sequencing two independent PCR fragments.

The *npr-1 lon-2;unc-6ΔC(urIs77)* animals were constructed by crossing *npr-1 lon-2* males with *unc-6ΔC(urIs77)* hermaphrodites and selecting Lon Rol F2 progeny. The *urIs77* transgene maps to LGII and is followed by the Rol phenotype and by the pan-neural expression of *unc-119::GFP*. For the branching assay, the *unc-6ΔC(urIs77)* transgene was crossed into *urEx162* animals by standard methods for unlinked genes.

Transgenic animals expressing an NPR-1::GFP reporter were obtained by microinjection of pIM#200, a plasmid constructed by inserting 7 kb of upstream regulatory sequence and 1 kb of *npr-1* coding sequence from

pM4 in frame with the GFP coding sequence of vector pPD95.79 (supplied by A. Fire, Carnegie Institution of Washington, Baltimore, MD). Transgenes were integrated by γ -ray irradiation and four independent lines were established. To express *npr-1(ur89)* ectopically in DD and VD motor neurons, the *unc-129* motor neuron-specific promoter (Colavita et al., 1998) was placed upstream of the *npr-1* coding sequence, the *npr-1* sequence was altered to encode the C178Y change, and the GFP coding sequence was inserted in frame immediately before the stop codon sequence. The pIM#201 plasmid was used to create integrated transgenic strains. Expression of the transgene was monitored by the motor neuron expression of GFP.

Axon outgrowth and aldicarb sensitivity assays. For the outgrowth assay, animals were scored by epifluorescence microscopy. Living animals were mounted on a slide in a small drop of M9 buffer on a 5% agar pad. L4 larvae or young adults were randomly picked and scored for the presence of ectopic processes at the ventral sublateral-lateral boundary on the right side of the body wall between the vulva and the retrovesicular ganglion (Lim et al., 1999). The *unc-6ΔC(urIs77)* animals were picked at random from plates in this study, whereas in the study by Lim et al. (1999), animals with the strongest roller phenotype were selected. Animals with a strong roller phenotype have a slightly lower percentage of ectopic axons.

For scoring the DD and VD motor neurons, immunofluorescence histochemistry was used to stain animals for GABA as described by McIntire et al. (1992) using rabbit anti-GABA antiserum (Sigma, St. Louis, MO) and Cy^{tm3}-conjugated AffiniPure goat anti-rabbit antiserum (Jackson ImmunoResearch, West Grove, PA).

Acute sensitivity to aldicarb (Chem Services Inc., West Chester, PA) was determined as described previously (Lackner et al., 1999). Briefly, in each experiment, 20 L4 worms were placed on 1 mm Aldicarb plates and prodded every 10 min over a 2 hr period to determine whether they retained the ability to move; worms that failed to respond to this harsh touch were classified as paralyzed. Each experiment was repeated a minimum of three times.

RESULTS

Netrin UNC-6 and UNC-6ΔC affect axon guidance and branching of the DD and VD motor neurons

Netrin UNC-6 is required to guide circumferential cell and axon migrations (Hedgecock et al., 1990; Ishii et al., 1992). To investigate the relationship between axon guidance and axon branching that is influenced by UNC-6, we examined the DD and VD motor neurons (Fig. 1). The guidance of the circumferential axons of these particular neurons was shown by McIntire et al. (1992) to require UNC-6. We reasoned that the branching of these axons might be especially susceptible to the effects of UNC-6, because each neuron extends circumferentially an UNC-6 responsive axon from an UNC-6 nonresponsive axon that runs longitudinally in the ventral nerve cord. The DD axons develop in the embryo and the VD axons develop in the larva. Together these neurons circumferentially extend 17 axons along the right body wall and 2 axons along the left body wall in wild-type animals (White et al., 1986). The axons are GABAergic and can be specifically visualized using anti-GABA antibodies (McIntire et al., 1992).

We scored the total number of DD and VD motor neuron axons leaving from the right side of ventral nerve cord in *unc-6* null mutants. We found that an average of 16.3 ± 0.7 ($n = 23$) axons leave in wild-type animals, whereas an average of 8.0 ± 1.8 ($n = 33$) axons leave in *unc-6* null mutants (Fig. 2B). This raises the possibility that DD/VD neurons may fail to extend circumferential axons in the *unc-6* null mutant. It is also possible that these branches form but simply stay within the ventral nerve cord; however, analyses of the ventral nerve cord of *unc-6* mutants by electron microscopy show that the average number of axons in the cord is equal to that in the wild-type animals (Hedgecock et al., 1990; McIntire et al., 1992). Axons that do dorsally extend often wander laterally and may branch and terminate at lateral posi-

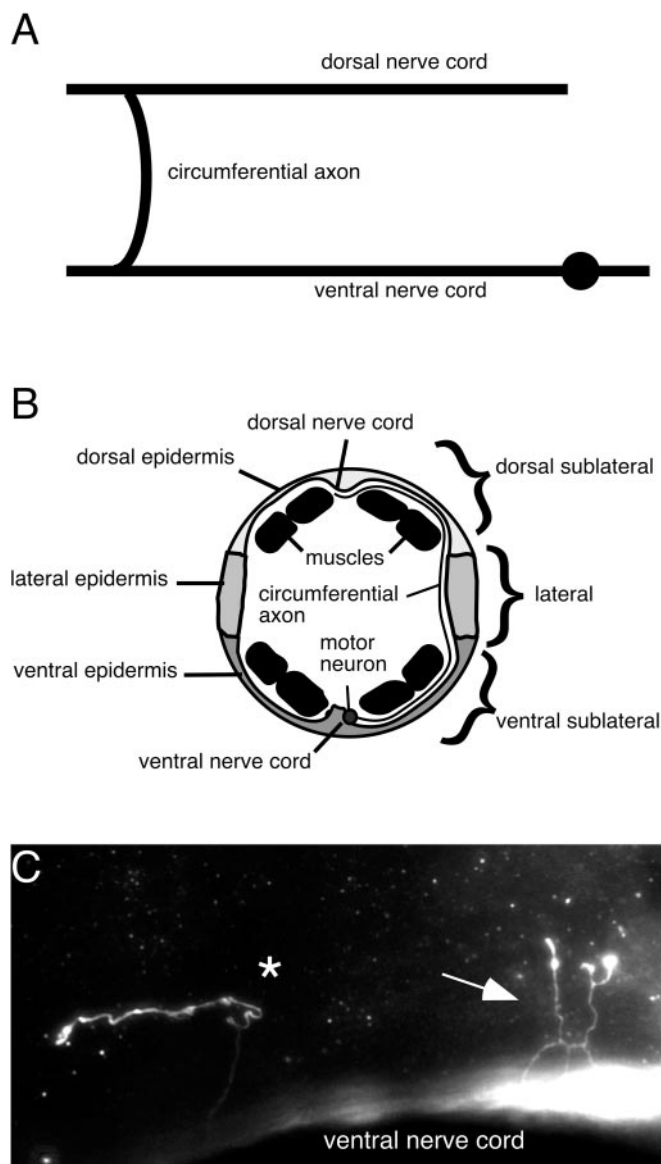


Figure 1. The morphology of DD and VD motor neurons is shown. *A*, The morphology of the DD and VD neurons. The circumferential axons extend from the distal end of the longitudinal axon in the ventral nerve cord and migrate dorsally to the dorsal nerve cord. *B*, Schematic transverse section of the adult hermaphrodite body wall. The DD and VD circumferential axons migrate between the basal surface of the epidermis and the basement membranes. Axon morphology was scored at the different dorsoventral positions indicated. *C*, The DD and VD neurons were visualized using anti-GABA immunocytochemistry. In *unc-6* null mutants, the circumferential axons wander laterally. Some axons terminate without branching (asterisk), whereas others produce ectopic branches before terminating (arrow).

tions (Fig. 1*C*). We analyzed the morphology of these circumferential DD/VD axons by scoring the position at which they terminate and the number of branch points (Fig. 2*B,C*). In *unc-6* null mutants, most axons wander and terminate in the ventral sublateral region after leaving the dorsal nerve cord (Fig. 2*B*); 67% ($n = 166$) have ectopic lateral branches (Fig. 2*A*). Likewise, in *unc-5* and *unc-40* mutants the frequency of ectopic branching is similar, although in *unc-40* mutants the majority of the circumferential axons are guided to the dorsal midline. These results indicate that UNC-6 plays a role in the dorsal extension of the

circumferential axons from the ventral nerve cord and in preventing the ectopic branching of the axons once they leave the cord.

The expression of UNC-6 ΔC was shown to have partial UNC-6 guidance activity and to cause axon branching of ventral nerve cord motor neurons (Lim et al., 1999). Here the morphology of the DD and VD neurons was specifically examined. We find that the number of DD and VD axons that extend from the ventral nerve cord in animals that express *unc-6 ΔC* in the *unc-6* wild-type and *unc-6(-)* background is nearly normal (Fig. 2*B*). Furthermore, the percentage of circumferential axons that reach the dorsal midline is greater in these animals than in *unc-6* null animals (48% in *unc-6(+);unc-6 ΔC* , 42% in *unc-6(-);unc-6 ΔC* , and 1% in *unc-6(-)* animals) (Fig. 2*B*). However, despite the improved guidance of the axons, ectopic branching is not reduced in the *unc-6 ΔC* animals (Fig. 2*C*). These results, together with the branching observed in *unc-40* mutants, indicate that the ectopic branching is not simply the result of the failure of dorsal guidance; they raise the possibility that UNC-6, but not UNC-6 ΔC , mediates another response that prevents inappropriate branching of the circumferential axons.

Alleles of *npr-1* suppress the branching of the circumferential DD and VD axons in *unc-6 ΔC* animals

The above model predicts separate UNC-6 activities that modulate the axon guidance and branching responses. Therefore, we reasoned that it might be possible to isolate mutations that affect the ectopic branching but not the guidance response to UNC-6 ΔC . A genetic screen was performed to isolate mutations that suppress the branching of additional processes from ventral nerve cord motor neurons in *unc-6 ΔC* animals (Fig. 3*A,B*). This screen took advantage of the observation that the additional motor neuron branches caused by the expression of an *unc-6 ΔC* transgene, *urIs77*, in an otherwise wild-type animal result in an uncoordinated phenotype (Lim et al., 1999). We reasoned that mutations that suppress the additional branching might restore wild-type movement. Because in general axon guidance mutants have an uncoordinated phenotype, selecting mutagenized animals with wild-type movement should isolate mutations that affect the ability to induce the extra motor neuron branches but not axon guidance. Thus, in principal, this screen isolates new mutations only if the axon branching response is separable from the axon guidance response. From a screen of 40,000 haploid genomes, we isolated six mutations that partly suppress the *urIs77*-induced motor neuron processes.

We have identified one suppressor, *ur89*, as an allele of *npr-1*. Without *unc-6 ΔC* expression, the *ur89* allele causes a social behavior phenotype; that is, the mutants aggregate together on food to form clumps. The same phenotype was described for *npr-1*, a gene that encodes a predicted seven-transmembrane receptor of the neuropeptide Y receptor family (de Bono and Bargmann, 1998). We examined previously identified alleles of *npr-1* and found that *npr-1(ad609)* also suppresses all additional motor neuron processes in a fraction of *unc-6 ΔC* animals; however, *npr-1(ky13)* and *npr-1(n1353)* do not (Fig. 3*C*). The expression of *unc-6 ΔC* partially suppresses the clumping phenotype conferred by any of the *npr-1* mutations (data not shown). Genetically, the *ur89* allele fails to complement *npr-1(n1353)* or *npr-1(ky13)*, and the expression of an *npr-1(+)* transgene rescues the clumping phenotype of *ur89* animals and the *ur89* suppression of extra motor neuron processes in *unc-6 ΔC ;ur89* animals. By DNA sequence analysis, we determined that the *ur89* mutation changes residue 178 from a cysteine to a tyrosine. Cysteine 178 is con-

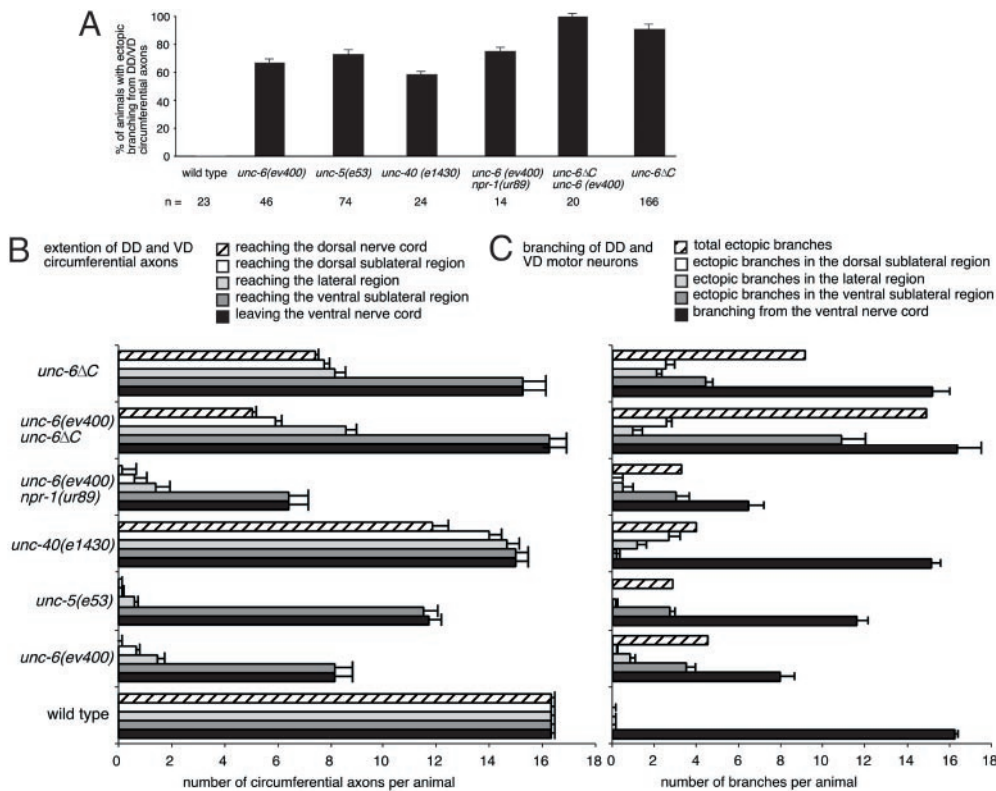


Figure 2. Netrin UNC-6 and UNC-6 Δ C affect the morphology of the DD and VD motor neurons. **A**, For each strain, the percentage of the population that had ectopic axon branching of the circumferential axons was measured by scoring for ectopic branches anywhere along the entire length of an animal. The ectopic branching occurs in *unc-6*, *unc-5*, and *unc-40* loss-of-function mutants as well as in animals expressing *unc-6 Δ C*. **B**, The number of circumferential axons extending from the ventral nerve cord and reaching each of the different dorsoventral positions was measured. For each axon, the trajectory that projected farthest dorsally was scored; the dorsal extension of ectopic branches was not included. A significant fraction of the axons reaches the dorsal nerve cord in *unc-40*, *unc-6(-);unc-6 Δ C*, and *unc-6(+);unc-6 Δ C* animals. **C**, The number of axon branch points occurring at each dorsoventral position was measured. For each axon, only the branch points that occurred along the trajectory that projected farthest dorsally were scored. The ectopic branching is enhanced in *unc-6(+);unc-6 Δ C* animals. The means \pm SEM are reported.

served among neuropeptide Y receptors and is predicted to occur in the second extracellular loop between transmembrane domains 4 and 5 (de Bono and Bargmann, 1998). In comparison, *npr-1(ky13)* introduces a stop codon after the first of seven transmembrane domains, whereas substitutions occur in *ad609* at transmembrane domains 2 and 4 and in *n1353* at transmembrane domain 3 (de Bono and Bargmann, 1998).

npr-1 is expressed in ventral nerve cord motor neurons that produce additional branches in response to UNC-6 Δ C

The DA, DB, DD, and VD motor neurons have been observed to have additional ventral nerve cord motor neuron branches in *unc-6 Δ C* animals (Lim et al., 1999). To determine whether the *npr-1* suppressors might act within any of these ventral nerve cord motor neurons, we made a reporter construct by fusing the sequence coding for GFP after the *npr-1* sequence that encodes to the 10th amino acid of the fourth predicted transmembrane domain. We observed strong expression in DD and VD motor neurons (Fig. 4). In *unc-6 Δ C* animals, ectopic DD and VD branches are induced; these are completely suppressed in a large fraction of the *npr-1(ad609);unc-6 Δ C* and *npr-1(ur89);unc-6 Δ C* animals (Fig. 3D). Expression in DA and DB neurons was weak or absent. Note that rather than scoring the number of processes directly, which is difficult because unambiguously identifying a branch can be difficult (Fig. 3B), we measured the proportion of animals that have ectopic processes. This was done by scoring for the presence of any processes at the sublateral–lateral boundary in a region of the animal in which normally no processes are observed (see Materials and Methods) (Lim et al., 1999) (Fig. 3A,B).

The analysis of *npr-1* provides evidence that signaling pathways within the DD and VD motor neurons mediate the development of the additional processes. For each strain, transcription of the *unc-6 Δ C* transgene was confirmed by the expression of the GFP

comarker; the presence of the UNC-6 Δ C protein, which contains an HA epitope tag, was confirmed by Western blot analysis (data not shown). Our results indicate that different cells express NPR-1 and UNC-6 Δ C. To further examine the cell-autonomous suppression of branching by *npr-1(ur89)*, we ectopically expressed the mutant receptor NPR-1(C178Y) in the cholinergic DA and DB ventral nerve cord motor neurons, which are the other subset of motor neurons affected by *unc-6 Δ C* expression. We observed that when the receptor was expressed in *unc-6 Δ C* animals in the *npr-1* null background the combined ectopic branching of DD, VD, DA, and DB neurons was reduced, whereas the ectopic branching of DD and VD neurons alone was unchanged (Fig. 3C,D). By inference, we conclude that the mutant receptor can function cell autonomously even in the DA and DB motor neurons to suppress ectopic branching.

The *npr-1(ur89)* and *npr-1(ad609)* mutations may specifically affect the activity of components involved in regulation of ectopic branching in *unc-6 Δ C* animals. Aside from suppressing the branching response and causing the *npr-1* phenotype of mutant social behavior, the *ur89* and *ad609* alleles do not have other obvious phenotypes. Interestingly, other alleles of *npr-1*, including the predicted loss-of-function (lf) alleles, affect the social behavior but do not affect the number of motor neuron branches. The *ur89* mutation changes one of the two cysteine residues that are thought to form a disulfide bond that governs the topology of the extracellular loops of G-protein-coupled receptors. This topology is predicted to be critical for receptor activation (Perlman et al., 1995; Le Gouill et al., 1997; Zhang et al., 1999). We speculate that the *ur89* and *ad609* alleles produce an altered NPR-1 protein that inactivates downstream effectors required to mediate the ectopic branching, perhaps by sequestering a factor.

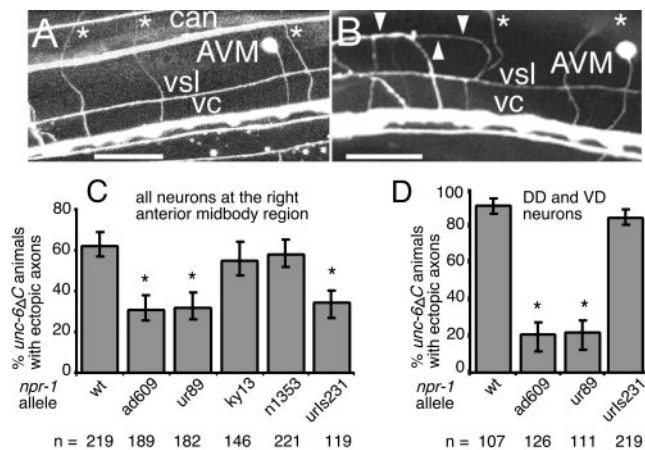


Figure 3. Alleles of *npr-1* suppress the UNC-6 Δ C-elicited outgrowth of ventral nerve cord motor neuron processes. *A–D*, The percentage of animals that had any additional processes induced by *unc-6ΔC* expression was measured by scoring for displaced axons at the anterior midbody right ventral sublateral–lateral boundary using pan-neuronal expression of GFP (Lim et al., 1999) (*A–C*) or for all DD and VD axons using anti-GABA antibodies (*D*). *A*, In the wild type, ventral nerve cord motor neuron processes migrate longitudinally in the ventral nerve cord (*vc*) and circumferentially (*asterisks*) past the ventral sublateral nerve (*vsl*), the lateral canal-associated nerve (*can*), and toward the dorsal midline. The AVM axon migrates to the *vc*. *B*, Expression of *unc-6ΔC* induces the outgrowth of additional processes that migrate from the ventral nerve cord motor neurons, past the *vsl*, and to the boundary (*arrowheads*) between the ventral sublateral and lateral epidermal cells (Lim et al., 1999). No axons are present in wild-type animals at this position. As in wild-type animals, the AVM axon migrates ventrally and circumferential motor neuron axons (*asterisks*) migrate to the dorsal midline (out of the plane of view). Scale bar: *A, B*, 25 μ m. *C*, *npr-1* alleles were examined for the suppression of all additional outgrowth induced by *unc-6ΔC* expression. *npr-1(ky13)* introduces a stop codon after the first of the seven transmembrane domains of the neuropeptide Y receptor homolog and is most likely a loss-of-function allele (de Bono and Bargmann, 1998). *ur1s231* is an integrated transgene that expresses the *npr-1(ur89)* sequence under a promoter that drives expression in the DA and DB neurons. These animals were scored in the *npr-1(ky13)* background. *D*, Expression of *npr-1(ad609)* and *npr-1(ur89)* was tested for the ability to suppress the UNC-6 Δ C-elicited outgrowth of DD and VD motor neuron processes. *Asterisks* indicate values that differ from control *unc-6ΔC* animals at $p < 0.001$; error bars indicate the SE of proportion.

Mutations that affect second messenger systems enhance or suppress ectopic branching in *unc-6ΔC* animals

The evidence that signaling pathways within the motor neurons influence the branching of additional motor neuron processes led us to examine whether known neuromodulatory pathway mutations also have an effect. We first tested whether altering G-protein signaling could affect the induction of ectopic axon branches, because neuropeptide Y receptors are G-protein-coupled and because it was reported that adenosine A2b receptor, also a G-protein-coupled receptor, is a netrin-1 receptor (Corset et al., 2000). We found that *egl-30(n686)*, which affects one of the G-protein α -subunits ($Gq\alpha$) (Brundage et al., 1996), suppresses all induced motor neuron processes in a significant fraction of *egl-30(n686);unc-6ΔC* animals (Fig. 5A). Whereas EGL-30 is required for viability, *egl-30(n686)* is a splice acceptor site mutation that causes reduced copies of full-length EGL-30 and is not lethal (Brundage et al., 1996).

EGL-30 is a component of a signaling network that is present in most, if not all, neurons in *C. elegans*. In the ventral nerve cord motor neurons, activation of the $Gq\alpha$ pathway stimulates neuro-

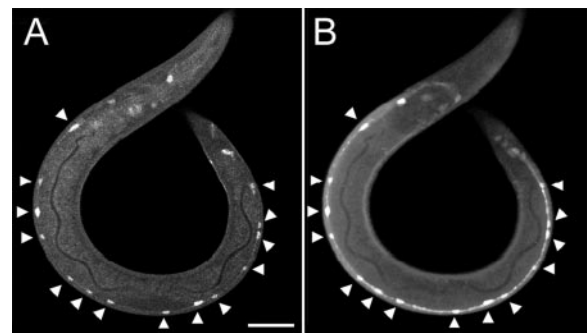


Figure 4. *npr-1* is expressed in DD and VD ventral nerve cord motor neurons. *A, B*, Immunofluorescence micrographs of a larva expressing a *npr-1::gfp* reporter. The larva was stained with anti-GFP antibodies (*A*) and anti-GABA antibodies (*B*). NPR-1::GFP is coincident with the GABAergic motor neurons DD and VD of the ventral nerve cord (*arrowheads*). Scale bar, 25 μ m.

transmitter release (Lackner et al., 1999; Miller et al., 1999). EGL-30, a homolog of phospholipase C β (PLC β), is predicted to be a downstream effector of $Gq\alpha$ EGL-30 (Lackner et al., 1999; Miller et al., 1999); we found that *egl-8(n488)*, a loss-of-function allele, also suppresses all motor neuron processes induced by *unc-6ΔC* in a significant fraction of *egl-8(n488);unc-6ΔC* animals (Fig. 5A). Activated PLC β cleaves phosphatidylinositol 4,5-bisphosphate to produce inositol 1,4,5-trisphosphate and diacylglycerol (DAG). In turn, these two second messengers may modulate intracellular events through their respective regulation of intracellular free Ca^{2+} and protein kinase C isozymes (Singer et al., 1997). In addition, DAG binds the presynaptic protein UNC-13 and recruits it to release sites (Nurrish et al., 1999). DGK-1, a diacylglycerol kinase (DAK), acts antagonistically to EGL-30 and EGL-8, presumably by converting DAG to phosphatidic acid and thereby reducing DAG levels (Miller et al., 1999). Consistent with this antagonistic role, *dgk-1(nu62)*, a loss-of-function allele (Nurrish et al., 1999), increases the proportion of *unc-6ΔC* animals with extra ventral nerve cord motor neuron branches (Fig. 5A).

Calcium/calmodulin-dependent protein kinase (CaMKII) is an enzyme that is thought to be critical for regulating synaptic strength and other neural functions. In *C. elegans*, there is one CaMKII gene, *unc-43*, and mutations affect neuronal gene expression and the density of ventral nerve cord synapses (Reiner et al., 1999; Rongo and Kaplan, 1999; Troemel et al., 1999). CaMKII activity is reduced by the loss-of-function mutation *unc-43(n498n1186)*, whereas constitutive calcium-independent CaMKII activity is caused by the gain-of-function (*gf*) mutation *unc-43(n498)*. Compared with *unc-6ΔC* animals, a greater proportion of animals with additional ventral nerve cord motor neuron branches is observed in the *unc-43(lf);unc-6ΔC* strain and all of the additional motor neuron branches are suppressed in a significant fraction of *unc-43(gf);unc-6ΔC* animals (Fig. 5A). These results suggest that the level of UNC-43 CaMKII activity is important for regulating the branching response to UNC-6 Δ C.

Calcium influx through voltage-gated calcium channels appears to be one means by which UNC-43 CaMKII is activated (Troemel et al., 1995; Rongo and Kaplan, 1999). Moreover, cytoplasmic Ca^{2+} levels affect growth cone extensions and can regulate the turning response of cultured *Xenopus* axons to netrin-1 (Gomez and Spitzer, 1999; Hong et al., 2000; Zheng, 2000). Therefore, we examined whether *unc-2* and *unc-36*, genes that encode

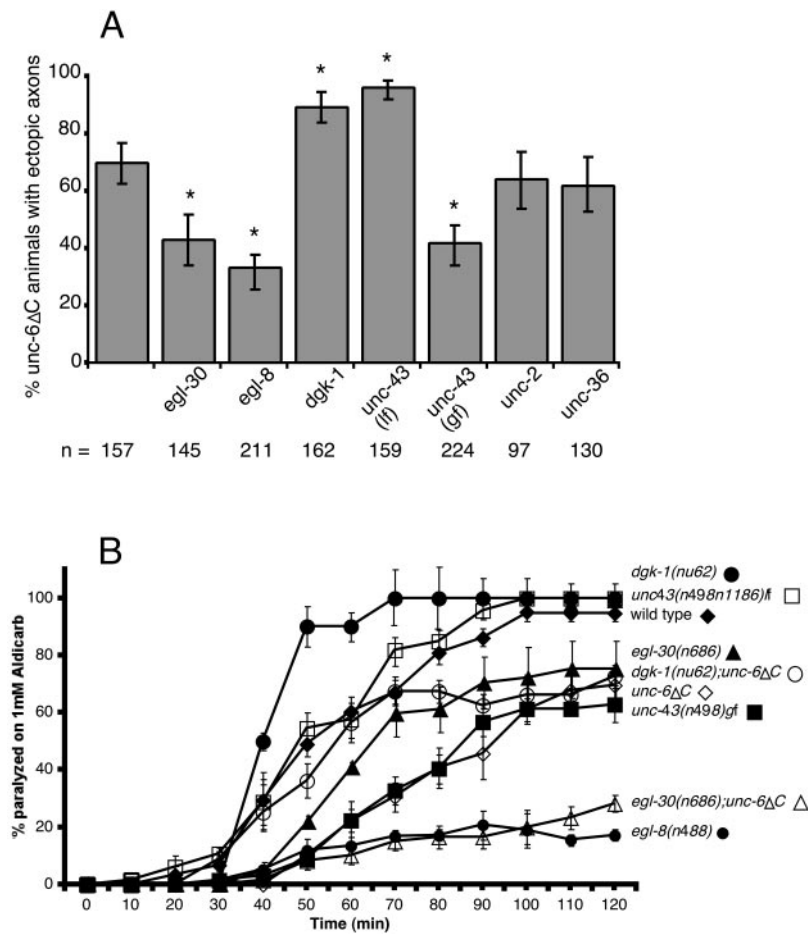


Figure 5. Mutations that affect neuromodulatory pathways enhance or suppress the UNC-6ΔC-elicited outgrowth of ventral nerve cord motor neuron processes. *A*, Seven different mutations affecting Gqα, (*egl-30*), phospholipase Cβ (*egl-8*), DAK (*dgk-1*), CaMKII (*unc-43 lf* and *gf*), the α₁ subunit of voltage-dependent calcium channels (*unc-2*), and the α₂ subunit of voltage-dependent calcium channels (*unc-36*) were examined for the ability to suppress or enhance the UNC-6ΔC-elicited outgrowth of ventral nerve cord motor neuron processes. Asterisks indicate values that differ from control *unc-6ΔC* animals at *p* < 0.001; error bars indicate the SE of proportion. *B*, The synaptic release of acetylcholine was assayed by the degree of paralysis induced by the cholinesterase inhibitor aldicarb. Expression of *unc-6ΔC*, *egl-8(sa47)*, *dgk-1(nu62);unc-6ΔC*, *egl-8(sa47);unc-6ΔC*, *egl-30(n686);unc-6ΔC*, and *unc-43(n498)gf* decreases the release of acetylcholine, making animals more resistant to aldicarb. Expression of *dgk-1(nu62)* enhances the release of acetylcholine and makes the animals less resistant. Data points are the mean ± SEM of at least three trials.

α₁- and α₂-subunits of voltage-dependent calcium channels, respectively (Schafer and Kenyon, 1995; Lee et al., 1997), influence the branching. We found that the loss-of-function alleles *unc-2(e55)* and *unc-36(e251)* do not affect the number of ventral nerve cord motor neuron branches (Fig. 5*A*), suggesting that the branching response to UNC-6ΔC is independent of calcium influx through these channels.

Physiological state of the neurons potentiate branching in *unc-6ΔC* but not *unc-6* wild-type animals

Our genetic analyses indicate that the branching of the ventral nerve cord motor neuron elicited by UNC-6ΔC can be enhanced or suppressed by certain neuromodulatory mutations. However, by themselves these mutations do not induce additional motor neuron branches (0%, *n* = 100 for each). Although altering Ca²⁺ and cAMP levels *in vitro* affects axon responses to netrin (Ming et al., 1997; Song and Poo, 1999; Hong et al., 2000), axon guidance defects are not observed in the mutants we tested. However, in some cases there are defects in the positioning of neuronal cell bodies (Tam et al., 2000) (our unpublished observations). To verify that the physiological state of the neurons is altered by the mutations and to establish the relationship between the branching response and second messenger signaling activity, we determined the sensitivity of animals to the acetylcholinesterase inhibitor aldicarb. In this assay, sensitivity is a measure of acetylcholine release; reduced acetylcholine release confers resistance to aldicarb, whereas increased acetylcholine release causes hypersensitivity (Lackner et al., 1999; Miller et al., 1999). As noted previously, acetylcholine release can be altered by *dgk-1*, *egl-8*, and

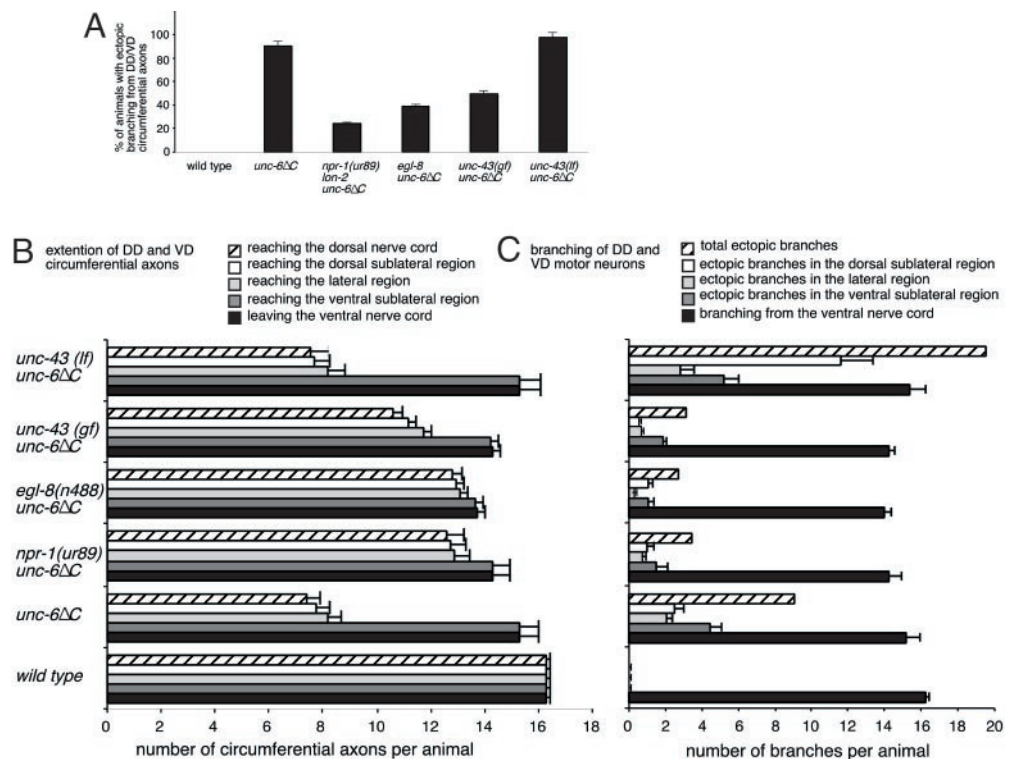
egl-30 mutations (Lackner et al., 1999; Miller et al., 1999), and we now show that alleles of *unc-43* and *unc-6ΔC* also affect release (Fig. 5*B*). However, ectopic circumferential axon branches are observed only in *unc-6ΔC* animals, indicating that an altered physiological state induced by the mutations is not sufficient to modulate branching in *unc-6* wild-type animals. However, the physiological changes caused by these mutations can affect branching in *unc-6ΔC* animals. The *egl-30(n686);unc-6ΔC* animals have less release than *unc-6ΔC* animals and fewer of these mutants have additional branches of the ventral nerve cord motor neuron. In contrast, *dgk-1(nu62);unc-6ΔC* animals have more release than *unc-6ΔC* animals and more of these mutants have additional motor neuron branches. We conclude that the second messenger signaling networks affected by these mutations are altering the physiological state of the neurons, and that these changes can potentiate branching in *unc-6ΔC* animals but not in wild-type animals.

It is interesting that *unc-6ΔC* expression and the mutations that affect the patterning of axon branching in *unc-6ΔC* animals affect acetylcholine release. This establishes a connection between acetylcholine release, axon branching, and the responses to an extracellular protein that guides migrations.

Guidance mediated by UNC-6ΔC is affected by ectopic branching

Several of the mutations inhibit the ectopic branching of the DD and VD circumferential axons. We investigated whether this inhibition affects the dorsal guidance of the axons (Fig. 6). In *unc-6ΔC* animals, which also express endogenous UNC-6, ectopic

Figure 6. Mutations that affect second messenger systems affect axon morphology in *unc-6ΔC* animals. **A**, For each strain, the percentage of the population that had ectopic axon branching of the circumferential axons was measured by scoring for ectopic branches anywhere along the entire length of an animal. The percentage of *unc-6ΔC* animals with ectopic branches is reduced by the presence of the *npr-1*, *egl-8*, or *unc-43(gf)* mutations. **B**, The number of circumferential axons extending from the ventral nerve cord and reaching each of the different dorsoventral positions was measured. For each axon, the trajectory that projected farthest dorsally was scored; the dorsal extension of ectopic branches was not included. In *unc-6ΔC* animals, the presence of the *npr-1*, *egl-8*, or *unc-43(gf)* mutations improves dorsal guidance, whereas *unc-43(lf)* does not. **C**, The number of axon branch points occurring at each dorsoventral position was measured. For each axon, only the branch points that occurred along the trajectory that projected farthest dorsally were scored. In *unc-6ΔC* animals, the presence of the *npr-1*, *egl-8*, or *unc-43(gf)* mutations suppressed ectopic branching, whereas *unc-43(lf)* enhanced branching. The means ± SEM are shown.



axon branches are observed across all lateral regions, with nearly one-half of the axons reaching the dorsal midline. In comparison, *unc-6ΔC* animals with the *npr-1(ur89)*, *egl-8(n488)*, or *unc-43(gf)* mutation have fewer ectopic axon branches across all lateral regions and the axons extend further dorsally, with a majority reaching the dorsal midline (Fig. 6B,C). Thus, inhibiting ectopic branching improves the directed axon extension. However, stimulation of ectopic branching by the *unc-43(lf)* mutation does not alter the ability of the axons to be dorsally guided relative to the expression of *unc-6ΔC* alone (Fig. 6B,C). Taken together, these results suggest that the ability of UNC-6ΔC to mediate dorsal guidance is affected by the mutations, but dorsal guidance mediated by the endogenous UNC-6 is not. This further supports the idea that the ectopic branching is a direct consequence of UNC-6ΔC rather than a consequence of disrupting endogenous UNC-6 functions.

DISCUSSION

Both UNC-6 and UNC-6ΔC can guide circumferential DD and VD axons. Compared with *unc-6(-)* animals, *unc-6(-);unc-6ΔC* or *unc-6(+);unc-6ΔC* animals have better extension of circumferential axons from the nerve cord and better dorsal guidance of the circumferential axons; however, the penetrance of the ectopic branching phenotype is not reduced. In comparison with *unc-6(-);unc-6ΔC* animals, *unc-6(+);unc-6ΔC* animals show slightly improved dorsal guidance and less ectopic branching. These results suggest that UNC-6ΔC competes with the endogenous UNC-6, which can suppress the ectopic branching. This was also suggested from the results of the expression of different *unc-6ΔC* transgenes in *unc-6(-)* and *unc-6(+)* backgrounds (Lim et al., 1999). We also show that mutations that affect CaMKII- and DAG-dependent signaling modulate the ectopic branching phenotype in *unc-6ΔC* animals but do not affect axon morphology in UNC-6 wild-type animals. Our interpretation of these results is

that UNC-6 C is responsible for inhibiting the effects of CaMKII- and DAG-dependent signaling, which, if not silenced, can modulate axon morphology.

The molecular mechanism by which UNC-6ΔC triggers ectopic branching is unknown. Netrin is thought to induce receptor complexes that can trigger different types of axon responses depending on the components they contain (Hong et al., 1999; Stein and Tessier-Lavigne, 2001). UNC-6ΔC may allow the formation of UNC-6 receptor complexes that can promote directed extension of growth cones but cannot inhibit responses that lead to branching. This inhibition requires the UNC-6 C domain working in *cis* to the N-terminal domains within the receptor complex. It is interesting that the UNC-6 C module has been found in a number of proteins, including the complement C345 protein family, frizzled related proteins, type I C-proteinase enhancer proteins (PCOLCEs), and tissue inhibitors of metalloproteinases (TIMPs) (Ishii et al., 1992; Leyns et al., 1997; Banyai and Patthy, 1999). In PCOLCE and TIMP proteins, the UNC-6 C module is involved in the regulation of metalloproteinase activity (Murphy et al., 1991; Hulmes et al., 1997; Langton et al., 1998). This raises the possibility that without UNC-6 C the UNC-6ΔC-containing complexes are more susceptible to regulation by proteases. It has been found that chemical inhibitors of metalloproteinases potentiate netrin-mediated axon outgrowth *in vitro* and that the netrin receptor homolog of UNC-40, deleted in colorectal cancer (DCC), is a substrate for metalloproteinase-dependent ectodomain shedding (Galko and Tessier-Lavigne, 2000).

Models of UNC-6/netrin guidance predict axon responses to gradients of the molecule. Our results indicate that the ectopic branching in *unc-6ΔC* animals is caused by a separate branching mechanism that is sensitive to UNC-6 C function, rather than by guidance errors caused by a novel distribution of UNC-6ΔC.

First, expression of UNC-6 Δ C causes the ectopic branching in only a subset of UNC-6 responsive neurons that extends along the entire body wall (Lim et al., 1999). A novel distribution of UNC-6 Δ C would be expected to affect all of the UNC-6 responsive axons that are present along the body wall. Second, when UNC-6 is ectopically expressed, the branching phenotype is not observed, although the guidance of axons is severely disrupted in such animals (Ren et al., 1999). This indicates that a novel distribution of UNC-6 is not sufficient to cause the ectopic branching phenotype. Third, circumferential axon migrations are partially rescued when *unc-6* Δ C is expressed in *unc-6* null animals, indicating that the proposed gradient and guidance information of UNC-6 Δ C is not significantly different from that of UNC-6 in wild-type animals (Lim et al., 1999). Finally, we have uncovered mutations in genes that enhance or suppress the ectopic branching by acting within the branching neurons themselves. It is more likely that the mutations affect the cellular machinery that mediates an axon branching response than the extracellular distribution of UNC-6 Δ C.

Second messenger signaling pathways, particularly cyclic nucleotides and Ca²⁺, are thought to play an important role in the regulation of axon responses to extracellular guidance molecules (for review, see Song and Poo, 1999; Gomez et al., 2001). For example, *in vitro* culture assays using *Xenopus* spinal neurons have shown that intracellular Ca²⁺ and cAMP levels are involved in dictating growth cone behavior in response to netrin-1 (Ming et al., 1997; Hong et al., 2000). Moreover, CaMKII, acting in a Ca²⁺-dependent manner, can mediate growth cone turning in response to acetylcholine (Zheng et al., 1994), and antagonist blocking of the acetylcholine receptor can inhibit the attractive response to netrin-1 (K. Hong, personal communication). This is interesting because the Gq α EGL-30–PLC β EGL-8 pathway produces DAG in response to acetylcholine in *C. elegans* (Brundage et al., 1996; Lackner et al., 1999). Thus, observations in culture and in *C. elegans* are consistent with the notion that the CaMKII- and DAG-dependent signaling cascades are linked in the control of UNC-6/netrin responses.

CaMKII- and DAG-dependent signaling, which can modulate DD and VD axon morphology and cause ectopic branching, must be silenced during the dorsally directed migrations. In the *unc-6* wild-type background, CaMKII- and DAG-dependent signaling are blocked by the activity mediated by UNC-6 C. In *unc-6* Δ C animals, the ability of CaMKII- and DAG-dependent signaling to alter axon morphology is not inhibited, and mutations such as *dgk-1(lf)* and *unc-43(lf)*, which stimulate the signaling activity (as judged by their ability to elevate acetylcholine release), increase branching activity. Conversely, other mutations that inhibit signaling activity (by decreasing acetylcholine release), such as *egl-8*, *egl-30*, and *unc-43(gf)*, diminish branching activity. The silencing effect of UNC-6 C is physiologically significant, because the suppression is required to prevent inappropriate responses that would cause erroneous morphological changes. Extracellular guidance molecules may have evolved strategies to counteract some process of the guidance machinery that tends to introduce branching. Although our results do not directly address what causes axons to branch at their normal stereotyped positions, they suggest that any mechanism that releases the inhibition mediated by UNC-6 C could trigger branching or turning responses.

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