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Cover legend: In each panel, the radial distance from the center represents contrast along a particular color direction. The saturation of the color at a point in the plane indicates the strength of the response of neurons tuned to that color direction. Different panels show the population response either in the control state (center panel) or after prolonged exposure to color modulation along four different directions in the isoluminant plane. Left and right panels of the center row show the effects of habituation that stimulates only L and M cones (left–right); top and bottom panels of the middle column show the effects of habituation that stimulates only S cones (top–bottom); panels at top left and bottom right, and bottom left and top right, show effects of habituation to intermediate directions that stimulate all three classes of cones. For more information, see the article by Tailby et al. in this issue (pages 1131–1139).

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1213 The Expression of MicroRNA miR-107 Decreases Early in Alzheimer's Disease and May Accelerate Disease Progression through Regulation of β -Site Amyloid Precursor Protein-Cleaving Enzyme 1

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1236 Hyperoxia Causes Maturation-Dependent Cell Death in the Developing White Matter

Bettina Gerstner, Tara M. DeSilva, Kerstin Genz, Amy Armstrong, Felix Brehmer, Rachael L. Neve, Ursula Felderhoff-Mueser, Joseph J. Volpe, and Paul A. Rosenberg

Corrections: In Figure 1 of the article “Accelerated Accumulation of Misfolded Prion Protein and Spongiform Degeneration in a *Drosophila* Model of Gerstmann–Straussler–Scheinker Syndrome,” by Brendan A. Gavin, Maria J. Dolph, Nathan R. Deleault, James C. Geoghegan, Vikram Khurana, Mel B. Feany, Patrick J. Dolph, and Surachai Supattapone, which appeared on pages 12408–12414 of the November 29, 2006 issue, the anti-PrP monoclonal antibody D13 (InPro Biotechnology, South San Francisco, CA) was used to compare PrP expression levels between different transgenic *Drosophila* lines. In retrospect, the authors have realized that the D13 epitope encompasses residue 101 and that consequently, this antibody has poor affinity for P101L PrP. As a result, the Western Blot shown in Figure 1 does not accurately reflect the relative PrP expression levels between the PRNP/GAL4-Cha2 and GAL4-Cha2;P101L^D lines assayed. Using two alternative anti-PrP antibodies, whose epitopes do not include residue 101, the authors have found that GAL4-Cha2;P101L^D flies actually express >20-fold more PrP than PRNP/GAL4-Cha2 flies. Thus, with the transgenic lines currently available, the authors cannot exclude the possibility that expression of WT PrP at higher levels might also cause spongiform degeneration in *Drosophila*.

For the article “The Neural Coding of Stimulus Intensity: Linking the Population Response of Mechanoreceptive Afferents with Psychophysical Behavior,” by Michael A. Muniak, Supratim Ray, Steven S. Hsiao, J. Frank Dammann, and Sliman J. Bensmaia, which appeared on pages 11687–11699 of the October 24, 2007 issue, the formula for r_{max} (Eq. 6) (page 11695, in Results, Population firing rate), is incorrect. The fraction within the parentheses should be flipped. The correct formula, used in all subsequent calculations and derivations, is as follows:

$$r_{max} = r_0 \left(\frac{A}{10^\beta} \right)^{\frac{1}{2}}.$$

In the article “The Structure of Multi-Neuron Firing Patterns in Primate Retina,” by Jonathon Shlens, Greg D. Field, Jeffrey L. Gauthier, Matthew I. Grivich, Dumitru Petrusca, Alexander Sher, Alan M. Litke, and E. J. Chichilnisky, which appeared on pages 8254–8266 of the August 9, 2006 issue, there were two errors. On page 8255 (in Materials and Methods, Spike sorting, third sentence), “spikes were identified using a threshold of three times the voltage SD” should read “spikes were identified using a threshold of four times the voltage SD.” On page 8256 (in Materials and Methods, Stimulation and receptive field analysis, first paragraph), “intensity 9200 (8700, 7100)” should read “intensity 4300 (4200, 2400).”

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