Table 1. Reaction conditions used for the quantitative RT-PCR assay

<table>
<thead>
<tr>
<th>Gene</th>
<th>PCR cycle parameters</th>
<th>Number of cycles</th>
<th>Sense primer</th>
<th>Antisense primer</th>
<th>Product Size(bp)</th>
<th>GenBank Accession number</th>
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</thead>
<tbody>
<tr>
<td>NGF</td>
<td>94°C, 15 sec, 55°C, 2 min, 72°C, 2 min</td>
<td>26</td>
<td>247-266</td>
<td>630-649</td>
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<td>BDNF</td>
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<td>2124-2149</td>
<td>2845-2870</td>
<td>747</td>
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<td>Ret</td>
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</table>

Each cDNA product from the reverse transcription procedure was used as the template for PCR amplification in a reaction mixture containing PCR buffer (in mM: 10 Tris-HCl, pH 8.3, 50 KCl, 1.5 MgCl₂), 0.2 mM dNTPs, a 0.2 mM each set of oligonucleotide primers, and 2.5 U of Taq DNA polymerase in 100 µl of final volume. Primers for G3PDH mRNA were added to the PCR mixture after some reactions in order to make its final PCR cycle number to be 18.