# Arrest of Afferent Axon Extension by Target Neurons *In Vitro* Is Regulated by the NMDA Receptor

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Cerebellar granule neurons *in vitro* specifically arrest the extension of their appropriate presynaptic axons, mossy fibers. This "stop-growing signal" may be an essential step in the formation and specificity of synapses. Here, we have tested whether ionotropic glutamate receptors are involved in the stop-growing signal. When explants of basilar pontine nuclei, a mossy fiber source, were cultured on granule neurons, most pontine neurites terminated <200  $\mu$ m from their explant of origin, a criterion for the stop-growing signal. In contrast, treatment with the NMDA antagonist D(-)-2-amino-5-phosphonopentanoic acid (D-AP5) greatly increased the number of pontine neurites extending beyond 300  $\mu$ m, whereas treatment with NMDA reduced the number of pontine neurites extending beyond 200  $\mu$ m. A non-NMDA agonist (AMPA) and antagonist (6-cyano-7-

nitroquinoxaline-2,3-dione) did not alter pontine neurite lengths. None of these agents affected neurite outgrowth from pontine explants in the absence of granule neurons, nor did any agent affect the survival of granule neurons. These results indicate that NMDA and D-AP5 specifically perturb an interaction between axons and target cells necessary for the stopgrowing signal, and that NMDA receptors are critical for the development of a major cerebellar afferent system. These findings also suggest that NMDA-sensitive refinement of axon arbors during later development may involve the direct regulation of axon extension by target neurons.

Key words: cerebellum; granule cell; mossy fiber; NMDA receptor; basilar pontine nuclei; axon extension

Axons are highly specific in their innervation of targets, and this specificity manifests itself on several levels. First, axons from a given source innervate specific regions or layers in their target, e.g., in the optic tectum (Yamagata and Sanes, 1995). Second, axons are highly specific in the type of target cell with which they will synapse. For example, mossy fibers synapse with granule neurons of the cerebellar cortex and project into the Purkinje cell or molecular layers only transiently during development (Mason and Gregory, 1984), or when there are ectopic granule neurons in these locations (a rare occurrence) (Palay and Chan-Palay, 1974).

Although the specificity of axonal projections has been well documented, the mechanisms that control axonal growth during termination in specific cell layers and on specific cell types remain obscure. To understand more precisely how target cells regulate the growth of appropriate afferents, we developed an *in vitro* system based on purified cerebellar target neurons (Baird et al., 1992b). Neurite extension from explants of basilar pontine nuclei, a source of mossy fibers, is arrested by granule neurons, their appropriate target cell. This behavior is considered to be medi-

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ated by a "stop-growing signal" presented by target granule neurons to which mossy fibers respond specifically, as granule neurons do not inhibit the extension of axons from explants of retina, or inferior olivary nuclei (Baird et al., 1992a), a source of cerebellar climbing fibers. The stop-growing signal therefore may lead to the arrest of neurite elongation in the internal granule layer and not in other layers of the cerebellar cortex and also may contribute to the conversion of mossy fiber growth cones into mature synaptic terminations.

We have begun to determine which types of molecules comprise the stop-growing signal. Neurotransmitters are a class of molecules that could contribute to target cell selection by growth cones because they are cell- and system-specific and are also known to arrest axon elongation *in vitro* (Kater and Mills, 1991). Further, growth cones can release transmitters before they contact targets (Hume et al., 1983; Young and Poo, 1983), which would allow differences in growth-cone transmitter type and target cell receptor type to contribute to the regulation of axonal growth by specific target cells during their first contacts.

In this study, we have tested whether ionotropic glutamate receptors, which include NMDA receptors, are involved in the stop-growing signal presented to mossy fibers by cerebellar granule neurons. This class of receptors was selected because granule neurons are glutamatergic and express a profile of NMDA receptor subunits that differs from other cells of the cerebellum (Audinat et al., 1994; Moyner et al., 1994). In addition, neurotransmitters can affect the rate of axon extension by altering intracellular calcium levels in the growth cone (Kater and Mills, 1991), and the NMDA receptor is known to gate calcium currents (Mayer et al., 1987).

#### **MATERIALS AND METHODS**

Tissue culture. Granule neurons were purified from dissociated cerebellar cells from postnatal day 5–7 rats, as described (Baird et al., 1992a,b), and plated in Labtek chamber slides (NUNC, Naperville, IL) coated first with 0.5 mg/ml poly-D-lysine (Sigma, St. Louis, MO), then with 20 μg/ml laminin (Gibco, Grand Island, NY). Granule neurons were cultured under conditions known to be effective in arresting the growth of pontine mossy fibers (Baird et al., 1992a): granule neurons were plated at 7500 cells/mm² and cultured in serum-free medium consisting of Eagle's basal medium with Earle's salts (Gibco) supplemented with 20 mM sodium bicarbonate, 2 mM L-glutamine (Gibco), glucose (final concentration 32 mM), penicillin/streptomycin (Gibco, 20U/ml each), 5 mg/l insulin–5 mg/l transferrin–5 μg/ml sodium selenite (Sigma), and 1% bovine serum albumin, fraction V (Sigma). Cultures were incubated at 35.5°C in 5% CO<sub>2</sub>.

After 24 hr, the medium was replaced with serum-free medium or serum-free medium containing 20 or 50  $\mu$ M D(-)-2-amino-5-phosphonopentanoic acid (D-AP5; Tocris Cookson, Bristol, UK), 10  $\mu$ M 6-cyano-7-nitroquinoxaline-2,3-dione (CNQX; Tocris Cookson), 20 or 50  $\mu$ M NMDA (Sigma), or 50  $\mu$ M AMPA (Tocris Cookson). Explants measuring ~200  $\mu$ m in diameter were dissected from the basilar pontine nuclei of newborn mice and plated on subconfluent granule neuron monolayers prepared 24 hr earlier (Baird et al., 1992a,b).

Immunocytochemistry. After an additional 48 hr, the cocultures were fixed in phosphate buffered 4% paraformaldehyde, pH 7.4, and the neurites of explant origin were labeled with a monoclonal antibody specific for mouse neurons, M6, using the indirect immunoperoxidase method (Baird et al., 1992a,b; Lund et al., 1986).

Quantitation of neurite outgrowth. The number of pontine neurites extending beyond 300  $\mu m$  from the center of their pontine explant of origin was determined using the Bioquant System IV for computerized morphometry (R & M Biometrics, Nashville, TN) and a Zeiss microscope with phase-contrast optics (Thornwood, NY) (Baird et al., 1992a,b). In control conditions (explants on laminin in serum-free medium), there is a moderate amount of outgrowth extending beyond 300  $\mu m$ , making this a practical distance for quantitation. Comparable results were obtained for 200 and 400  $\mu m$  (not shown). For measurements of NMDA-treated cultures and controls cultured in serum-free medium without NMDA, the number of pontine neurites extending beyond 200  $\mu m$  was determined. Two hundred micrometers was chosen, as opposed to the 300  $\mu m$  mark used for the experiments with antagonists in the present study, because few pontine neurites extended beyond 300  $\mu m$  on granule neurons when cultured in NMDA-containing medium.

Estimation of granule neuron survival. Companion culture wells from the same chamber slides prepared for the studies described above were used to assess granule neuron survival. As before, purified granule neurons were plated in serum-free medium, and after 1 d in vitro, the medium was replaced with control serum-free medium or medium containing glutamate receptor agonists or antagonists (NMDA, AMPA, D-AP5, and CNQX). Granule neurons were fixed 3 d after plating in phosphate buffered 4% paraformaldehyde, pH 7.4. Estimates of neuron survival were obtained by counting the number of process-bearing neurons in five predetermined areas near the center of each culture well, with each of the five areas measuring 14,400  $\mu$ m². Measurements were made using the Bioquant System IV and a Leitz microscope (Leica, Deerfield, IL) equipped with phase-contrast optics.

### **RESULTS**

# Effect of ionotropic glutamate antagonists on the stop-growing signal

Explants of basilar pontine nuclei, a source of cerebellar mossy fibers (Brodal and Walberg, 1977; Burne et al., 1978), were cultured on granule neurons or on laminin. As in our previous studies (Baird et al., 1992a,b), pontine explants on laminin alone extended 100–300 neurites beyond 300  $\mu$ m from their centers, whereas on laminin with granule neurons, most pontine neurites terminated on granule neurons <100  $\mu$ m from the edge of their explant of origin, resulting in a reduction in long (>300  $\mu$ m) neurites of ~90% on average (Fig. 1A,D; Table 1). Pontine neurites extending on laminin grew in bundles of larger diameter than those extending on granule cells on laminin, where they appeared to extend as single fibers or in much smaller fascicles.

This indicates a greater tendency for pontine neurites to grow in fascicles on simple, defined substrates than on target granule neurons, which is consistent with previous comparisons of pontine neurite extension on polylysine, laminin, and granule neurons (Baird et al., 1992b).

We then tested whether NMDA receptors were involved in the stop-growing signal provided by granule neurons. NMDA or non-NMDA antagonists were added to granule cell cultures immediately before explants of pontine nuclei. Specific antagonists of NMDA or non-NMDA receptors were used: D-AP5 and CNQX, respectively. Among specific NMDA antagonists, D-AP5 was chosen because it is known to block currents gated by NMDA receptors in granule neurons, which have been extensively characterized (Garthwaite and Brodbelt, 1989; Silver et al., 1992; D'Angelo et al., 1993; Farrant et al., 1994). Compared with control cultures grown in serum-free medium, in medium containing 20 µm D-AP5, there was a fourfold increase in the number of long (>300 µm) pontine neurites on granule neurons (Fig. 1D,F; Table 1). In the absence of granule neurons, addition of D-AP5-containing medium to pontine explants cultured on laminin resulted in no statistically significant (p < 0.05) change in outgrowth (Fig. 1A, C; Table 1). Therefore, D-AP5 does not act solely on the pontine neurites to increase their extension, but interferes with an interaction between granule neurons and pontine neurites that is required for the stop-growing signal. As with D-AP5, similar results were obtained with kynurenic acid (1 mm), a general antagonist of both NMDA and non-NMDA ionotropic glutamate receptors (not shown). In contrast, the non-NMDA antagonist CNQX at 10  $\mu$ M had no effect on neurite outgrowth from pontine explants on laminin or on granule neurons on laminin (Table 1). Thus, functional NMDA receptors, but not non-NMDA receptors, are involved in the stop-growing signal.

# Effect of ionotropic glutamate agonists on the stop-growing signal

To further investigate the specific involvement of NMDA receptors, we used the selective agonists of ionotropic glutamate receptors, AMPA and NMDA. AMPA (50 µm), an agonist of non-NMDA receptors, did not affect pontine neurite extension on laminin, or extension on granule neurons on laminin compared with cultures without AMPA (Table 2). In cultures on laminin alone with or without AMPA, large numbers of pontine neurites extended beyond 300  $\mu$ m from the explant center. Similarly, the number of long neurites was dramatically reduced when pontine explants were cultured with or without AMPA on granule neurons on laminin. When the medium contained 20 μM NMDA, however, three- to fourfold fewer long (>200  $\mu$ m) neurites extended from explants cultured on granule neurons, compared with similar cocultures in medium without NMDA, indicating an increase in stop-growing activity by NMDA (Fig. 1D,E; Table 2). In NMDAtreated cultures, very few pontine neurites were observed to extend beyond the edge of the explant, and those that did terminated on granule neurons usually within 50 µm from the edge of the explant (Fig. 1E). The effectiveness of the stop-growing signal was further elevated by a higher concentration of NMDA (50  $\mu$ M), which decreased the number of long neurites by approximately sixfold compared with cocultures in medium without NMDA (Table 2). Both 20 and 50  $\mu$ M NMDA had relatively small, statistically insignificant (p > 0.05) effects on the extension of pontine neurites on laminin without granule neurons (Fig. 1A,B). Therefore, as suggested by the experiments with glutamate antagonists, these results with agonists indicate that NMDA-responsive

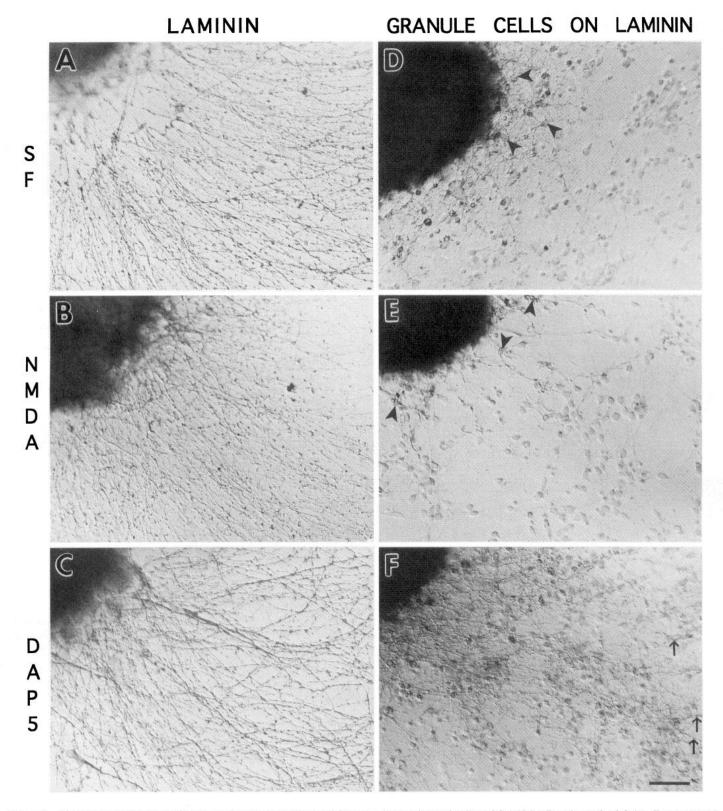


Figure 1. Neurite outgrowth from basilar pontine explants dissected from newborn mice and cultured for 48 hr. Panels on the left (A-C) are controls with pontine explants cultured on laminin in the absence of granule neurons. Numbers of neurites are similar in each case, reaching lengths much >250  $\mu$ m. A, Serum-free (SF) control medium; B, SF medium with 20  $\mu$ m NMDA; C, SF medium with 20  $\mu$ m D-AP5. D, Pontine explants cultured on granule neurons in SF medium. Most pontine neurites do not extend beyond granule cells (arrowheads) within 50  $\mu$ m from the edge of the explant. E, Pontine explants cultured on granule cells in SF medium with 20  $\mu$ m NMDA. Most pontine neurites do not extend beyond granule neurons (arrowheads) within 30  $\mu$ m from the edge of the explant. F, Pontine explants cultured on granule cells in SF medium with 20  $\mu$ m D-AP5. Abundant pontine neurite outgrowth extends beyond 200  $\mu$ m from the edge of the explant (arrows). Scale bar, 30  $\mu$ m (A-F).

Table 1. Effect of ionotropic glutamate receptor antagonists on neurite outgrowth from explants of pontine nuclei

Serum-free medium (SF)	Pontine neurites on laminin ± SEM <sup>a</sup>	Number of explants	Pontine neurites on granule cells $\pm$ SEM <sup>a</sup>	Number of explants
Control	$100 \pm 11$	14	11 ± 6.3	21
D-AP5 (20 μm)	$91 \pm 12$	10	$48 \pm 8.5^b$	16
CNQX (10 μM)	$108 \pm 9$	8	$6.9 \pm 6.6$	9

<sup>&</sup>lt;sup>a</sup>Number of neurites that extended beyond 300 μm on laminin in control medium is defined as 100%.

glutamate receptors are specifically involved in the stop-growing signal and affect an interaction between granule neurons and pontine neurites that mediates the stop-growing signal.

# Effect of glutamate agonists and antagonists on cell survival

NMDA receptor activity is known to affect granule neuron survival under certain culture conditions (Gallo et al., 1987; Didier et al., 1990; Schramm et al., 1990; Bessho et al., 1994). To test whether glutamate receptor activity affects the stop-growing signal by altering granule neuron survival, the number of granule neurons that were able to elaborate processes after 3 d in culture in the absence of pontine explants was determined in control medium and in media containing a glutamate agonist or antagonist (20 and 50 μM NMDA, 50 μM AMPA, 50 μM D-AP5, or 10 μM CNOX). In control cultures in which the medium was replaced with serum-free medium after 1 d of culture in serum-free medium without agonist or antagonist, the mean density (± SD) of process-bearing neurons was found to be  $4000 \pm 630 \text{ cells/mm}^2$ after 3 d of culture. When comparing control granule neurons cultured in serum-free medium with granule neurons cultured in serum-free medium containing agonists or antagonists, no significant difference (p < 0.10) in the number of process-bearing granule neurons was observed on day 3 when control medium had been replaced with control, agonist, or antagonist medium after 1 d (Table 3). A further indication that the agents tested did not affect neuron survival is that none of the agents had a statistically significant effect (p < 0.05) on neurite outgrowth when pontine explants were grown in the absence of target cells (Tables 1, 2). If the survival of pontine neurons were changed, a corresponding change in the number of neurites extending from pontine explants would also be expected.

### DISCUSSION

Our results indicate that NMDA receptor activity affects the arrest of pontine mossy fiber neurites by granule neurons, with NMDA enhancing, and its antagonist D-AP5, disrupting arrest.

None of the glutamate agonists or antagonists tested affected pontine neurite extension in the absence of granule neurons or the survival of the granule neurons themselves. These results have implications for the regulation of neurite extension by NMDA receptors during initial contacts between axons and target neurons, and during the later refinement of axon arbors.

#### Molecular character of stop-growing signals

The molecular mechanism by which target neurons interrupt the extension of afferent axons requires communication between target neuron and growth cone, which might be mediated by receptor–ligand binding. Neurotransmitters and their receptors are candidate molecules for this mechanism because they regulate neuronal motility, including neuronal migration (Komuro and Rakic, 1993) and axonal growth (Kater and Mills, 1991). In addition, neurotransmitters and their receptors are neuron-specific, which might allow target neurons to specifically regulate the growth of appropriate axons.

Neurotransmitters have been found to act in concert with other classes of molecules during the regulation of axonal growth, including adhesion molecules (Glanzman et al., 1990; Landmesser et al., 1990; Bailey et al., 1992; Mayford et al., 1992; Peter et al., 1994). Another stop-signal molecule, s-laminin, has been proposed to arrest the growth of motor axons when they arrive at future synaptic sites of the neuromuscular junction, based on its ability to inhibit the growth of axons *in vitro* (Porter et al., 1995). s-Laminin is also found in the CNS (Hunter et al., 1992) but has not been examined in the cerebellum (Sanes and Hunter, personal communication).

The stop-growing signal also might consist of a modulation of a neurite-promoting activity such as that offered by neurotrophins, including more recently identified factors in the CNS (Cohen et al., 1994; Gao et al., 1995; Segal et al., 1995). Evidence suggesting that neurotrophins could regulate mossy fiber growth includes brain-derived neurotrophic factor (BDNF) and NT-4/5 expression in the developing cerebellum (Maisonpierre et al., 1990;

Table 2. Effect of ionotropic glutamate receptor agonists on neurite outgrowth from explants of pontine nuclei

Serum-free medium (SF)	Pontine neurites on laminin ± SEM <sup>a</sup>	Number of explants	Pontine neurites on granule cells $\pm$ SEM <sup>a</sup>	Number of explants
Control	$100 \pm 8.8$	11	22 ± 4.4	14
AMPA (50 μM	$94 \pm 19$	9	$17 \pm 6.3$	11
NMDA (20 μм)	$136 \pm 25$	10	$5.4 \pm 2.0^{b}$	17
NMDA (50 μm)	94 ± 22	10	$3.7 \pm 1.2^b$	12

<sup>&</sup>quot;The number of neurites that extended beyond 200  $\mu$ m on laminin in control medium is defined as 100%, rather than 300  $\mu$ m as in Table 1 because of the small number of neurites extending beyond 300  $\mu$ m in NMDA-treated cultures of pontine neurites growing on granule neurons.

 $<sup>^</sup>bp < 0.0001$  compared with "Pontine neurites on granule cells" in SF control medium; all others p > 0.05 compared with corresponding control; t test for two independent samples.

 $<sup>^</sup>bp < 0.0001$  compared with "Pontine neurites on granule cells" in SF control medium; all others p > 0.05 compared with corresponding control; t test for two independent samples.

Table 3. Effect of glutamate agonists and antagonists on granule neuron survival

Culture condition	Process-bearing cells after 3 d (% control ± SD)		
20 μm NMDA	$110 \pm 14^a$		
50 μm NMDA	$123 \pm 35^a$		
50 μM AMPA	$101 \pm 19^a$		
50 μM D-AP5	$108 \pm 14^{a}$		
10 μm CNQX	$94 \pm 22^a$		

 $<sup>^{</sup>a}p > 0.10$  compared with control cultures in serum-free medium without agonist or antagonist, t test for two independent samples.

Rocamora et al., 1993; Timmusk et al., 1993), BDNF promotion of neurite outgrowth from pontine explants, and pontine expression of a BDNF and NT-4/5 receptor, trkB (Rabacchi et al., 1995).

Growth-cone collapsing factors might also participate in the stop-growing signal. The first collapsing factor to be isolated was collapsin, a brain-derived membrane glycoprotein, which, like the stop-growing activity of granule neurons, does not act on retinal axons (Luo et al., 1993). In contrast to the action of collapsin, which collapses growth-cone structure or induces turning of growth cones away from bound collapsin (Fan and Raper, 1995), pontine neurite extension is arrested by granule neurons often with the persistence of filopodia and lamellopodia (Baird et al., 1992b).

Neurotransmitters and other molecules involved in the stopgrowing signal would ultimately act on the cytoskeleton of mossy fiber growth cones to arrest their extension (Bentley and O'Connor, 1994). For example, binding of adhesion molecules during neurite extension over cellular substrates activates several second-messenger systems, including the influx of calcium (Williams et al., 1993), which destabilizes the cytoskeleton (Lankford and Letourneau, 1991). Indeed, some of the effects of neurotransmitters on neurite extension have been attributed to changes in calcium levels in growth cones, with decreased growth-cone motility accompanying changes in calcium concentration away from an optimal level (Kater and Mills, 1991). Increases in cytosolic calcium concentration have been shown to mediate growth-cone collapse in some cases (Bandtlow et al., 1993; Igarashi et al., 1993), but an exception has been identified (Ivins et al., 1991). It remains to be determined whether changes in growth-cone calcium levels accompany the stop-growing signal, but if so, such changes would depend on the presence of target granule neurons.

### NMDA receptor regulation of the stop-growing signal

NMDA may regulate the stop-growing signal by binding to receptors on mossy fibers, granule neurons, or both, but the presence of functional NMDA receptors has only been established for granule neurons (Garthwaite and Brodbelt, 1989; Silver et al., 1992; D'Angelo et al., 1993; Farrant et al., 1994). NMDA receptor subunits 1, 2A, and 2B are expressed, however, in the pontine nuclei of newborn rats (Akazawa et al., 1994). NMDA receptors on granule neurons might be directly activated by glutamate released from approaching mossy fiber growth cones, as has been described for acetylcholine, which is known to be released from extending motoneuron growth cones (Hume et al., 1983; Young and Poo, 1983). Subsequently, mossy fibers would then be induced to arrest their extension via a retrograde signal, such as nitric oxide or arachidonic acid, both known to be produced by granule neurons in response to NMDA receptor activation (Lazarewicz et al., 1990; Garthwaite, 1991). Preliminary results, however, indicate that perturbing nitric oxide using inhibitors of nitric oxide synthase or by adding hemoglobin does not affect the stop-growing signal (not shown).

Alternatively, NMDA may affect the stop-growing signal indirectly by acting on granule or pontine neurons. NMDA receptor activity in pontine neurons also might alter the ability of pontine growth cones to detect stop-growing signal molecules produced by granule neurons. Regulation of stop-growing signal molecules could occur via changes in gene expression after NMDA receptor activation, which is well documented (Didier et al., 1989; Szekely et al., 1990; Ghosh and Greenberg, 1995). Recent results indicate that paraformaldehyde-fixed (Baird and Kruk, 1994) and osmotically killed, as well as living-granule neurons, can provide a stop-growing signal, suggesting that the signal is a passively provided granule neuron surface or matrix molecule(s). These results suggest that the stop-growing signal does not require NMDA receptors, but that the amount or activity of the signal is regulated by NMDA receptor activity. Only in live neurons could NMDA receptor activity modulate the amount of such a molecule or its efficacy, perhaps via calcium activation of other second-messenger systems (Ghosh and Greenberg, 1995).

Although it is possible that NMDA and D-AP5 act on the stop-growing signal by affecting granule neuron survival and, in turn, the amount of granule neuron membrane, our results do not support this alternative. Under some culture conditions, high potassium (10–25 mm) or NMDA affects granule neuron survival (Gallo et al., 1987; Balázs et al., 1988; Didier et al., 1990) or neurite outgrowth (Pearce et al., 1987; Cambray-Deakin and Burgoyne, 1992). Conditions that enhanced the stop-growing signal (5 mm potassium and 20  $\mu$ m NMDA over a 2 d period), however, had no effect on granule neuron survival in our studies or in those of Balázs et al. (1988).

### NMDA receptors and the remodeling of axon arbors

In addition to their effects on the stop-growing signal, NMDA receptors have been shown to be involved in the refinement of axon arbors. In the cerebellum, active NMDA receptors are required during the elimination of multiple-climbing fiber innervation of Purkinje cells (Rabacchi et al., 1992). In the tectum, lateral geniculate nuclei, visual cortex, and trigeminal nuclei, active NMDA receptors are required during the remodeling of exuberant branches and the focusing of arbors to specific territories, stages that are essential for the proper establishment of synaptic circuitry (Goodman and Shatz, 1993; Li et al., 1994). Recent work by O'Rourke et al. (1994) in which retinal growth cones were imaged in real time during arborization in tectum has shown that reduction in synaptic activity increased the net rate of extension and withdrawal of new branches. Our results indicate that when cerebellar cultures are treated for 48 hr with NMDAantagonists, also presumably reducing synaptic activity, axons extend for longer distances over target cells. Perhaps a similar increase in growth-cone motility occurs in our cultures during axon extension in the presence of NMDA antagonists and contributes to the extension of longer axons. In addition, chronically decreasing the effectiveness of NMDA receptors in the frog tectum reduces the number of synaptic contacts formed by retinal axons (Yen et al., 1995). Whether the longer pontine axons we observed in D-AP5-treated cultures also form fewer synapses with granule neurons as predicted from the in vivo results with retinal axons remains to be determined.

Our results suggest that the NMDA receptor-dependent stages of axonal development ultimately involve the regulation of growth-cone advance by target cells either during initial contacts or later arborization, and thereby allow synaptic activity to shape both of these stages of development. NMDA receptors would not directly participate in the stop-growing signal but would serve to modulate its effector molecules, which are not yet identified. NMDA receptor activity is known to affect the form of axon arbors in several regions of the CNS, and in these cases, the stop-growing signal might be a link between changes in NMDA receptor activation and changes in the rate of neurite extension.

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