

# Propagating Activation during Oscillations and Evoked Responses in Neocortical Slices

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Population activity in the cortex is poorly understood. In this report we use voltage-sensitive dye imaging to examine the spatiotemporal patterns of a 7–10 Hz oscillation in neocortical slices from rat somatosensory areas. This oscillation appeared as a component of spontaneous epochs when the preparation was bathed in low [Mg] artificial CSF (ACSF) (Silva et al., 1991). Each epoch started with a synchronized spike, and 3–200 cycles of oscillation emerged afterward. Voltage-sensitive dye imaging revealed that the oscillations in the local field potential recordings were actually caused by a propagating population activation. This activation propagated in a relatively uniform size (not expanding). We call this confined, propagating activation a “dynamic ensemble.” During each oscillation cycle, one (occasionally two) dynamic ensemble(s) appeared in the slice and was sustained for 60–200 msec. Dynamic ensembles propagated at ~30 mm/sec; the activity could propagate in both

directions in cortical slices. The propagation consisted in part of “jumps,” the locations of which were not fixed. Dynamic ensembles were distinguishable from the epileptiform spikes that occurred in low [Mg] ACSF. Population events similar to dynamic ensembles were also evoked under conditions of unaltered excitability (slice in normal ACSF) by electrical stimulation that activated a low density of neurons in a large area. Our data suggest that self-sustained, spatially confined, and propagating dynamic ensembles might be related to the epoch oscillations in somatosensory cortex seen *in vivo* (Nicolelis et al., 1995) and thus resemble one form of population activation in the neocortex.

*Key words: oscillation; somatosensory cortex; dynamic ensemble; optical recordings; voltage-sensitive dyes; population activity; synchronous events; cortical slices*

Oscillations are common population events in the CNS (Gray, 1994). Synchrony during oscillations on different spatial scales has been extensively observed (Engel et al., 1991; Llinas and Ribary, 1993; Steriade et al., 1993; Nicolelis et al., 1995; Murthy and Fetz, 1996). It has been reported that over short distances of 1 mm or less, neighboring neurons dynamically form synchronized assemblies (Gerstein and Aertsen, 1985; Gerstein et al., 1989). Such dynamically organized neuronal assemblies have been a subject of interest (Singer, 1985; Palm, 1990; Churchland and Sejnowski, 1992; Amit, 1995; Freeman, 1995; Crick, 1996; Nicolelis et al., 1997). Imaging the spatiotemporal patterns of population oscillations may help to understand the correlation between long distance synchrony and localized neuronal assemblies.

Optically imaging an oscillation in the *in vivo* cortex is technically difficult. Intrinsic optical signals (Ts'o et al., 1990; Maloney and Grinvald, 1996; Wang et al., 1996) are too slow for imaging most oscillations. Voltage-sensitive dye imaging is fast, but the signals (without averaging) are usually smaller than the artifacts of brain pulsation. In measurements from rodent somatosensory cortex, the signal-to-noise ratio only allowed a resolution of epileptiform events in single trials. The evoked response to whis-

ker movement was ~1/10th the size of the signals of epileptiform events (London et al., 1989). Even with improvements in the recording techniques, averaging was required to resolve a sensory-evoked response in the cortex (Kleinfeld and Delaney, 1996; however, see Arieli et al., 1996, on cat visual cortex). Furthermore, voltage-sensitive dye recordings from turtle visual cortex (Prechtl et al., 1997) showed that the oscillation signals were approximately five times smaller than were the sensory-evoked response. Thus, examining the oscillation patterns from mammalian cortex *in vivo* would require substantial technical improvement [e.g., new dyes (Glaser et al., 1998)].

Imaging in brain slices has a large signal-to-noise ratio, allowing the resolution of oscillation patterns in single trials. Voltage-sensitive dye imaging has been used in studying population activation in brain slices (e.g., Grinvald et al., 1982; Albowitz and Kuhnt, 1991; Colom and Saggau, 1994; Nelson and Katz, 1995; Demir et al., 1998; Tsau et al., 1998); however, the spatiotemporal pattern of oscillations in adult neocortical slices has not been reported, probably because in adult slices oscillations have not been clearly distinguished from epileptiform activity.

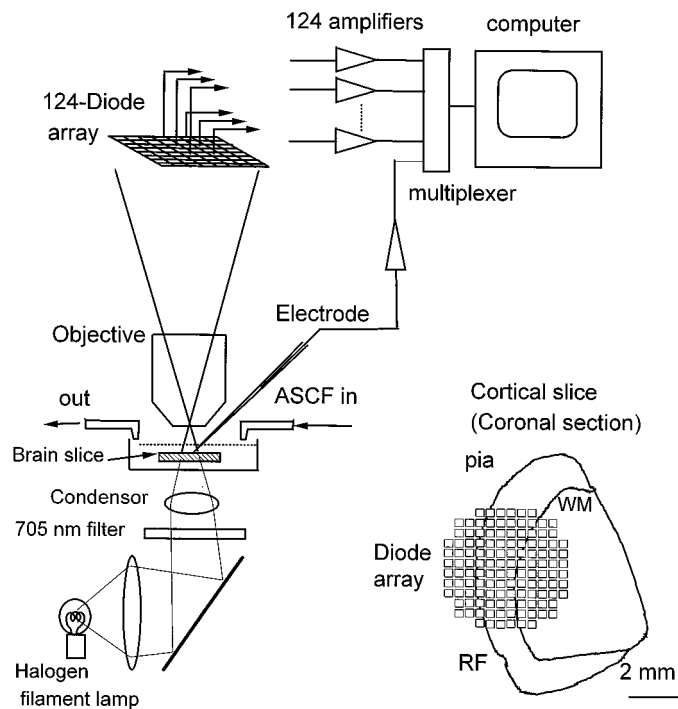
A cortical slice preserves a complex local network with a large number of intrinsic bursting neurons (Connors and Gutnick, 1990) that may serve as pacemakers for the oscillations (Silva et al., 1991). The 7–10 Hz oscillation in slices from rat somatosensory cortex (Silva et al., 1991) is similar to the oscillations occurring *in vivo* (Nicolelis et al., 1995). In this report we use this preparation to address two questions. (1) With many intrinsic bursters, does this oscillation start in one spot (a local pacemaker) and propagate to other areas or start from many spots, distributed in an interactive pattern? (2) Do active neurons form localized clusters during this oscillation?

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**Figure 1.** Apparatus for optical imaging. The slice preparation was continuously perfused with low [Mg] ACSF in a submerge chamber on the stage of a modified Leitz Orthoplan microscope (Wetzlar, Germany). The chamber and the perfusing solution were heated at 26°C. To reduce the vibration noise for optical recordings, we mounted the microscope on an air table and paused the perfusion during optical recording trials. We made absorption measurements using transmission illumination. Light from a halogen tungsten filament bulb (12 V; 100 W) was collected and illuminated the preparation through a  $705 \pm 60$  nm filter. The image was formed by a  $2.8\times$  (NA, 0.05) objective. With this optical magnification, the six laminae of the cortex were projected to five rows of photodetectors (*lower right diagram*), and a 4-mm-wide cortical section was monitored by  $\sim 50$  neighboring detectors. RF, Rhinal fissure; WM, white matter.

## MATERIALS AND METHODS

Cortical slices were harvested from postnatal day 21 to adult Sprague Dawley rats of both sexes. Following the National Institutes of Health guidelines, the animals were deeply anesthetized with halothane and quickly decapitated using a Stoelting small animal decapitator. The whole brain was moved into cold (0–4°C) artificial CSF (ACSF) containing (in mM): NaCl, 132; KCl, 3; CaCl<sub>2</sub>, 2; MgSO<sub>4</sub>, 2; NaH<sub>2</sub>PO<sub>4</sub>, 1.25; NaHCO<sub>3</sub>, 26; and dextrose 10, saturated with 95% O<sub>2</sub>/5% CO<sub>2</sub>, pH = 7.4. The brain was chilled in the cold ACSF for 90 sec before blocking and slicing.

The Paxinos and Watson (1986) stereotaxic atlas for rat brain was used to locate the cortical areas. The slicing angles were determined by the angles with which the brain was blocked. The tissue block was glued on a vibratome stage (752M Vibroslice; Campden Instruments, Sarasota, FL). Slices of 400  $\mu$ m thickness from somatosensory areas were obtained. The slices were then transferred into a holding chamber, immersed in ACSF, and oxygenated with 95% O<sub>2</sub>/5% CO<sub>2</sub> at room temperature for at least 2 hr. The slices were then perfused in ACSF containing the oxonol voltage-sensitive dye RH 479 [first synthesized by R. Hildesheim and A. Grinvald and kindly provided by Dr. L. Loew (University of Connecticut, Farmington, CT) as JPW 1131] at 0.02 mg/ml for 1 hr. The stained preparation was then perfused with dye-free ACSF at 29°C for 30 min before the measurements. Low [Mg] ACSF was used in some experiments. This ACSF was made with the same components and concentrations as the ACSF described above, but the concentration of MgSO<sub>4</sub> was reduced from 2 to 0.1 mM.

Optical imaging was performed with a 124-element photodiode array (Centronics, Newbury Park, CA) at a frame rate of 1000 frames/sec (Fig. 1). A  $2.8\times$  [numerical aperture (NA), 0.05] objective was used to project the image of the preparation to the array. On the array each detector

received light from a  $0.3 \times 0.3$  mm<sup>2</sup> area of the slice. The photocurrent of dye-related absorption signals (705 nm) from each photodetector was individually amplified through a two-stage amplifier system. The first stage of amplification performed a current-to-voltage conversion using a feedback resistor of 1 M $\Omega$ . The signals were then amplified and filtered by a second stage amplifier before digitizing. This second stage provides a voltage gain of 200 or 2000, high-pass filtering with a 100 msec time constant, and a four-pole Bessel analog low-pass filter with a 300 Hz corner frequency. [The amplifiers are commercially available as NeuroPlex systems (OptImaging, Fairfield, CT).] This parallel amplifier arrangement (Wu and Cohen, 1993) allows a low dark noise ( $10^{-6}$  of the illumination intensity), a large dynamic range (17–21 bits), and a fast sampling rate. The photocurrent on each diode was  $\sim 100$  nA. The optical signal size ( $dI/I$ ) for the oscillations was  $\sim 0.05$ – $0.2\%$  ( $5$ – $20 \times 10^{-4}$ ). The signal-to-noise ratio at our light level was  $\sim 5$  for oscillation signals, so that averaging was not necessary. Optical recording trials were 8–16 sec long; on each slice we record 4–10 trials (with a total exposure of <150 sec). Phototoxicity was not a concern with this level of exposure time and intensity. [For additional details about the apparatus, see Wu et al. (1999).]

Glass micropipettes or tungsten lacquer-coated microelectrodes were used to sample local field potentials (LFPs) and the spiking of individual neurons simultaneously. Glass electrodes were made from borosilicate glass containing a capillary fiber (0.75 mm inner diameter; 1.0 mm outer diameter), were filled with 1 M NaCl, and had resistance ranging from 2 to 5 M $\Omega$ . In most experiments, the recording electrodes were placed in layer II of the cortex. Tips of electrodes were inserted  $\sim 100$   $\mu$ m into the slice. LFP recordings were made with the use of two intracellular amplifiers (IE201; Warner Instrument Corporation, Hamden, CT). In some experiments, one electrode was used as a stimulus electrode when the amplifier probe was set to electrode input mode. Stimuli consisted of rectangular current pulses 10–30 V in amplitude and 50–200  $\mu$ sec in duration. Electrical recordings were digitized and stored concurrently with the optical images. For long-term electrical records, data were also stored on videotapes through a digital recorder (CRC VR-100) and then transferred to a personal computer (PC) for further analysis.

Signals from the photodiode array and electrodes were multiplexed and digitized with a DAP3200e/214 12-bit data acquisition board installed in a pentium PC and two multiplexer boards (Microstar Laboratories, Bellevue, WA) controlled by a program written in BASIC. Data were acquired and directly transferred into a hard drive.

The optical data were analyzed using the program NeuroPlex (A. Cohen and C. Falk; OptImaging). NeuroPlex displays the data in the form of traces for numerical analysis or pseudocolor images for visualizing the spatiotemporal pattern. The digitized data were further filtered by NeuroPlex, low-pass, 300 Hz (Butterworth) and high-pass, 1.2 Hz (RC filter) before displaying. The pseudocolor display was made using variable scaling and contour display. In the variable-scaling mode, the data from each diode are first normalized to their own maximum, and then each normalized data point is assigned a color according to a linear color scale (0 = deep blue and 1 = red). This kind of scaling best preserves the timing information in the signals. The contour mode spatially low-pass filters the raw data, so that the areas of equal signal amplitude are assigned the same color, and the amplitude difference between neighboring diodes is interpolated; the pseudocolor maps are not displayed in discrete pixels.

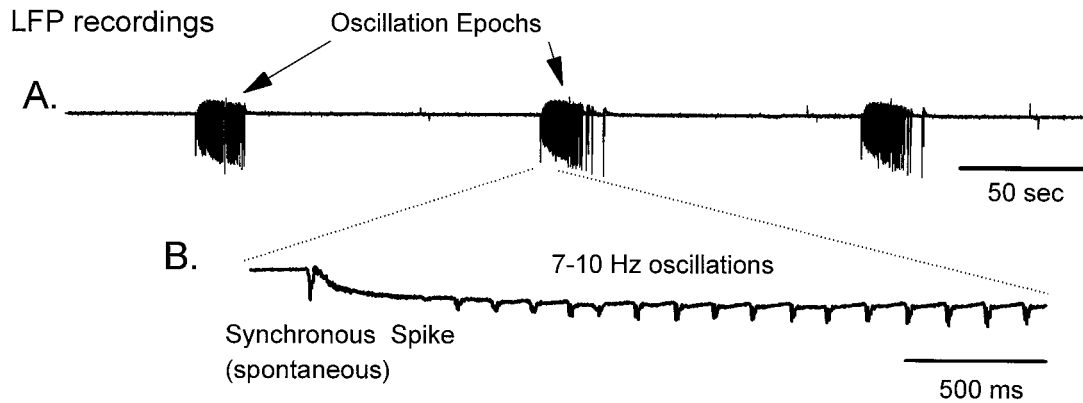
Numerical analysis, fast Fourier transformation (FFT), and auto- and cross-correlation were performed using MatLab (Mathworks, Natick, MA).

## RESULTS

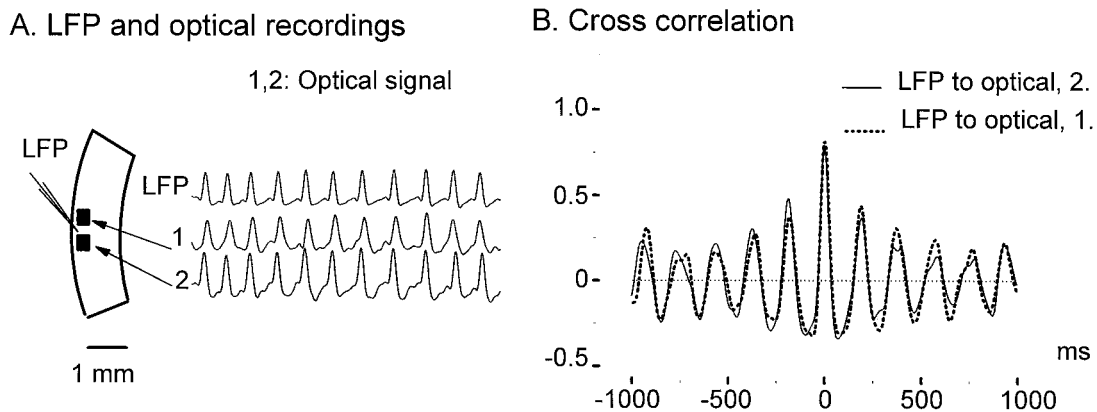
In this report we examine dynamically organized, multineuronal ensemble activity during oscillations. We are interested in whether this kind of activity exists during a spontaneous oscillation and can also be evoked as an all-or-none event. Here we first examine spatiotemporal patterns of 7–10 Hz oscillations in rat somatosensory cortical slices and then explore whether similar activation can be evoked by specific methods of stimulation.

### Optical signals of population oscillations

The oscillations appear in cortical slices from the somatosensory areas as part of the spontaneous epochs that occur when the preparation is bathed in low [Mg] ACSF (Silva et al., 1991; Flint and Connors, 1996) (Fig. 2A). Each epoch started with a synchro-



**Figure 2.** Epoch oscillations measured with an LFP electrode. *A*, When a cortical slice is perfused in low [Mg] ACSF, epochs of oscillations spontaneously occur. This recording is made by an electrode placed in cortical layer II, after 2 hr of perfusion in low [Mg] ACSF. *B*, An expanded display shows that an epoch started with a spontaneous synchronous spike and 7–10 Hz oscillations gradually emerged a few seconds later. In different preparations the epochs consisted of 3–200 cycles of oscillation. In all preparations tested ( $n > 50$ ), all oscillations appeared in spontaneous epochs, and each epoch started with a synchronous spike. However, in seven preparations, there were only one or two oscillation cycles in each epoch. The number of oscillation cycles in each epoch appeared to increase the longer the interepoch intervals were.



**Figure 3.** Comparison of electrical (LFP) and voltage-sensitive dye signals. *A*, During oscillations, the LFP signals and optical signals from neighboring sites were highly correlated. The recording sites were  $\sim 0.5$  mm apart, and there was a small phase difference between the different locations. *B*, The cross-correlation between LFP signals and the two optical detectors was high during the oscillations.

nous spike, and oscillations emerged afterward (Fig. 2*B*). In each epoch there was a variable number (usually 3–200) of oscillation cycles. The low-amplitude peaks of oscillations appeared to emerge from low-amplitude, asynchronized activities. As the epoch progressed, the oscillation frequency gradually decreased while the amplitude increased (Fig. 2*B*). At 26°C, the frequency was 5–8 Hz.

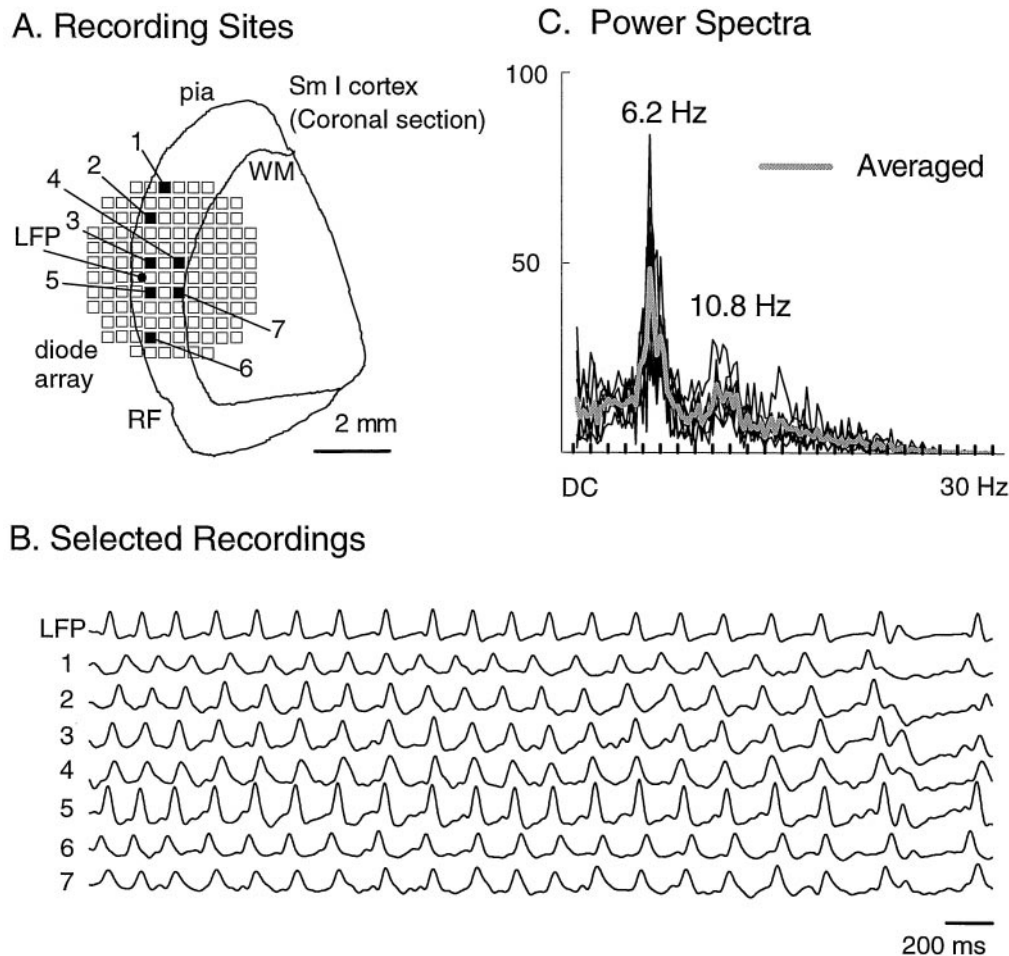
We found that these LFP signals were highly correlated with the voltage-sensitive dye signals. When LFP and voltage-sensitive dye signals were simultaneously recorded, the waveform of LFP and optical detectors at nearby areas was similar and highly correlated (Fig. 3). The optical signals were wavelength-dependent, with a maximum signal ( $dI/I$ ) at 705 nm and no signal at 670 nm. These are major features of the signal from RH 479, the oxonol voltage-sensitive dye. The amplitude of an optical signal is proportional to the depolarization of all the stained membranes under each detector (for review, see Wu and Cohen, 1993). These signals had a rapid time course and thus could not have come from other optical sources, such as activity-related intrinsic optical signals (Hochman et al., 1995; Malonek and Grinvald, 1996; Yuste et al., 1997; Peterson et al., 1998). These results indicate that the optical signals faithfully represent the

population neuronal activity, which allows us to examine this oscillation from many neighboring locations optically.

### Spatial distribution of the oscillations

In the measurement shown in Figure 4, a 4-mm-wide cortical slice was imaged (Fig. 4*A*), in which  $\sim 50$  neighboring locations were recorded simultaneously (Fig. 4*A,B*). In these measurements we began an optical recording trial when an epoch occurred. In most of the preparations examined ( $n > 20$ ), all locations oscillated at the same overall frequency (Fig. 4*C*). To our surprise, the correlations between two locations did not decrease significantly with distance, although there were large phase shifts at a large distance (Fig. 5*A*). The correlation was better between two points within one vertical “beam” than between two points separated by the same distance but in a tangential direction (Fig. 5*B*), suggesting that cortical columns might serve as synchronized units during oscillations.

Autocorrelations of the signals from five locations (Fig. 5*B*) suggested that the entire preparation was dominated by one frequency. However, the autocorrelation peaks were no longer superimposed after two cycles in either direction from 0 time, indicating that there were slow variations in frequency at different



**Figure 4.** Multiple site recording of the oscillations. *A*, A section of cortical slice (*Sm I* area) imaged by  $\sim 50$  neighboring photodetectors and one LFP electrode (in layer II). The imaging field was  $\sim 4.5$  mm in diameter. *RF*, Rhinal fissure; *WM*, white matter. *B*, Selected recording traces from the sites (solid squares in *A*). Both electrical and optical signals were low-pass filtered at 300 Hz, and the amplitude of the traces has been normalized to emphasize the phase relationships. *C*, The power spectra of the data in *B*. The gray trace is the average of all individual FFTs, in which the major frequency peak was 6.2 Hz (at 26°C). A minor frequency peak ( $\sim 10.8$  Hz) does not have a systematic representation in the raw data shown in *B*. This minor peak was also seen in *in vivo* LFP recordings (Nicoletis et al., 1995). This recording trial contained 16,000 frames of voltage-sensitive dye images. Subsequent figures (see Figs. 5–8) were made from selected sections of these same data.

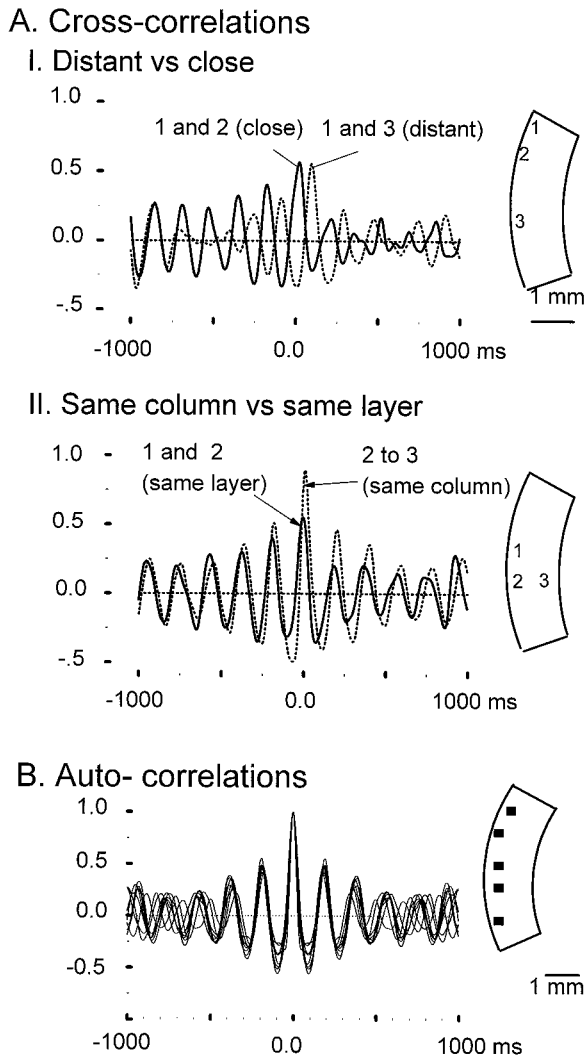
locations (Fig. 5*B*). A plot of the time interval between sequential cycles at several locations (Fig. 6) indicated that for most cycles (cycles 1–6, 11–17) the frequency variations were highly correlated at different locations. However, in addition to a general tendency to slow down at all locations, in 72 out of 105 of the epochs we found sudden changes in the cycle-to-cycle period that occurred simultaneously at different locations (as illustrated in Fig. 6, cycles 7–10). The correlations between different locations during this sudden change in frequency followed fixed patterns. The change appeared to occur first at one side of the preparation and moved to the other side in approximately two cycles. Although it is difficult to understand what is happening from the plot in Figure 6, the pseudocolor images from the same data set (Fig. 7*C*, *II*; see next section) offer a simple and elegant explanation for the sudden variation in frequency.

#### Propagating activations during the oscillations

The images derived from all the optical detectors revealed that the neurons activated during oscillations were not uniformly distributed over the slice. Instead, each oscillation peak appeared as a localized hot spot propagating across the cortex. In Figure 7 we made pseudocolor images based on the amplitude of the

voltage-sensitive dye signals, from three time sections (*I–III*) within one epoch. When the LFP recordings in Figure 7*B* are compared with the pseudocolor images in Figure 7*C*, it is obvious that each oscillation peak correlated to a propagating hot spot. In each oscillation cycle, when a hot spot passed by an LFP electrode (or an optical detector), a depolarization peak was detected. When many hot spots repeatedly passed a detector, a periodic depolarization was recorded that appeared as oscillations (Fig. 7*B*). Thus at each individual recording point, the activity could be defined as an oscillation by its periodicity in FFT and autocorrelations (Figs. 4*B*, 5*B*). However, the images of the same data set revealed propagating hot spots (Fig. 7*C*), suggesting that the periodicity is in part determined by the propagation velocity.

The shape of the hot spots varied somewhat from cycle to cycle of the oscillations, and no particular pattern was correlated with each individual preparation. It is difficult to determine accurately the physical size of a hot spot from the amplitude pseudocolor image. However, using the 50% isopotential lines of the contour map, we determined that the size of the spots was  $\sim 0.5$  mm wide in a tangential direction and encompassed all cortical layers. The spots were relatively similar in size but somewhat irregular in

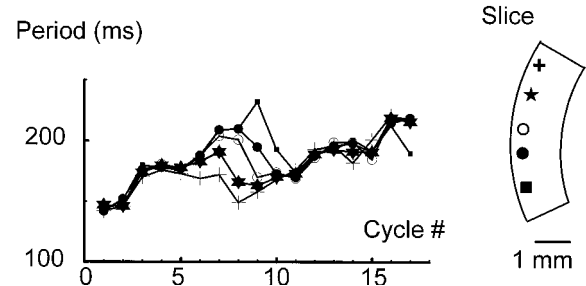


**Figure 5.** Auto- and cross-correlations. *A*, Cross-correlations. *I*, The amplitude of the correlations between the signals did not decrease significantly with distance. The *solid* and *dotted* traces are correlations between two detectors 800 and 3200  $\mu\text{m}$  apart, respectively. *II*, Signals from the same column (detectors 2, 3) have a somewhat larger correlation (*dotted* trace) than do the signals from the same layer but different columns (detectors 1, 2; *solid* trace). *B*, Autocorrelations of oscillation signals at five different locations (*filled rectangles*). At different locations, phase variations appear in outside peaks. The overall frequency was 6.2 Hz.

shape (Fig. 7). The cycle-to-cycle variability of the shape and size suggested that these propagating hot spots were most likely dynamically organized. The hot spots were seen in the slices with different slicing angles from the barrel cortex. The isopotential contour did not appear to be correlated to the barrel patterns, suggesting that the activity is likely to be organized by distributed intracortical connections. Because the hot spots are likely to be dynamically organized and they are not a classical stationary oscillation, we give them the name “dynamic ensembles” to describe this propagating activation and to distinguish it from other kinds of population activity.

#### Propagation, collisions, and jumps of dynamic ensembles

For each cycle of an oscillation, one and occasionally two dynamic ensembles emerged and propagated away. The overall propaga-



**Figure 6.** Variations in the cycle period at different locations. The cycle-to-cycle periodicity has sudden changes in some cycles (at cycles 7–10 in this epoch). At different locations the changes are in different directions. These kinds of sudden changes were very common. We examined 105 epochs from four preparations, and the sudden changes were found in 72 epochs.

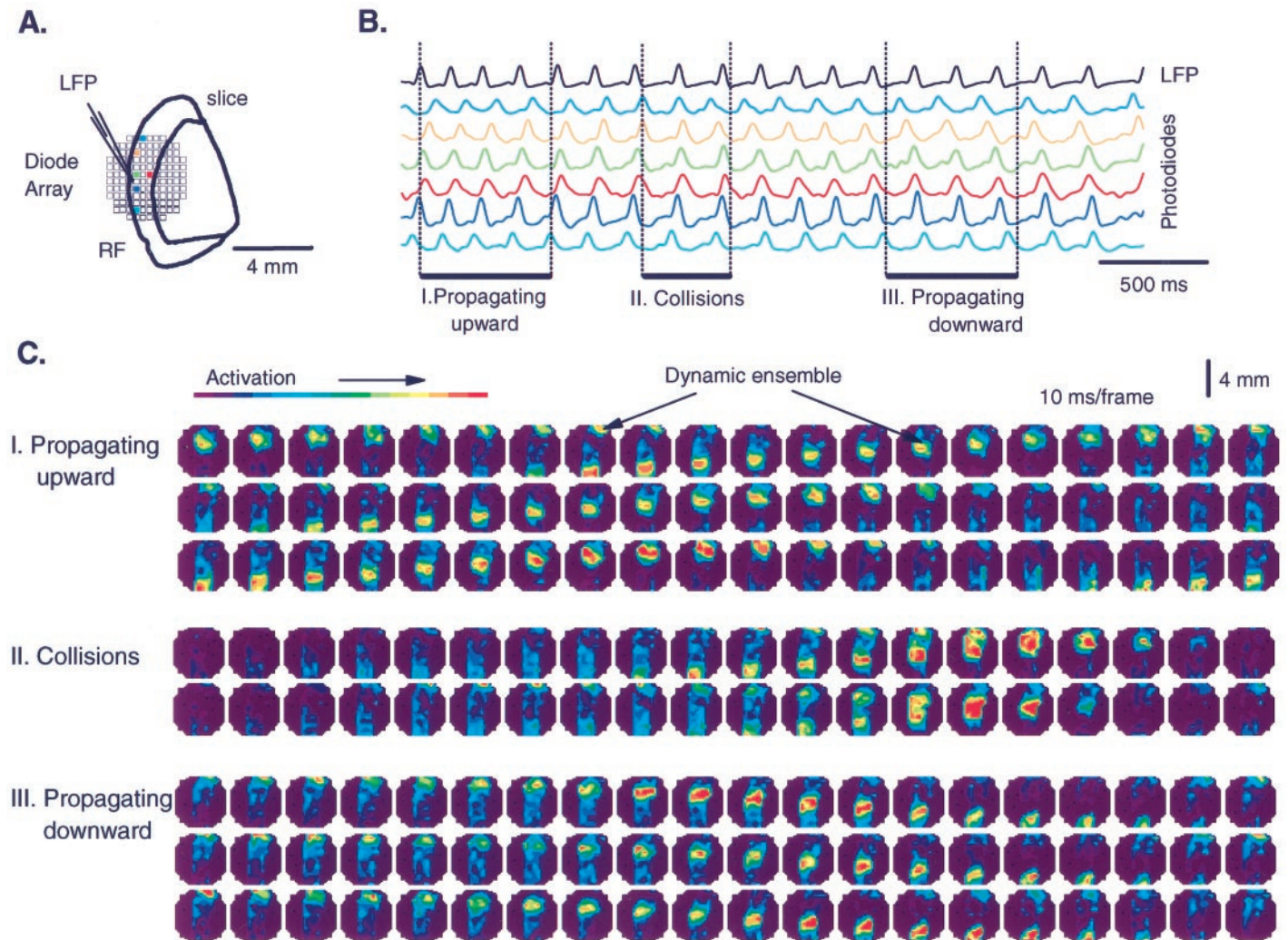
tion velocity was  $30 \pm 10$  mm/sec (mean  $\pm$  SD;  $n = 15$ ; at  $26^\circ\text{C}$ ), and the distance traveled was usually larger than the field that was imaged (i.e.,  $>5$  mm). In a brain slice only two propagation directions are possible. A consecutive series of dynamic ensembles usually propagated in the same direction; 97% ( $n = 200$ ) kept the same direction as their predecessor. However, the propagating direction was not fixed. The direction could reverse in the middle of an oscillation epoch (Fig. 7*B,C, I, III*). When two dynamic ensembles emerged simultaneously at different locations, they sometimes propagated toward each other, resulting in collision and annihilation of the activity (Fig. 7*B,C, II*). These collisions and changes in propagation direction explain why the oscillation frequencies varied drastically in the middle of some epochs (Fig. 6, cycles 7–10).

The propagation of excitation in cortical tissue often contains abrupt changes or “jumps” (Chervin et al., 1988; Wadman and Gutnick, 1993). We used the isopotential contour lines of the dynamic ensembles to examine variations in the propagation velocity. We found that the traveling course of each dynamic ensemble often had several jumps (Fig. 8*B*, arrows) that appeared as a rapid progression of the front edge of the isopotential contour line within a few milliseconds (Fig. 8*A*), followed by rapid withdrawal of the back edge (data not shown). Although the pattern of horizontal intracortical connections may be important in the propagation velocity of an excitation wave (Chervin et al., 1988; Wadman and Gutnick, 1993; Golomb and Amitai, 1997), in our data the locations of the jumps appeared to vary for each dynamic ensemble (Fig. 8*A, I–III*). It is thus more likely that the jumps in our image are dynamically distributed and not determined by the intracortical connectivity patterns.

#### Neuronal activity within a dynamic ensemble

The density of active neurons in a dynamic ensemble can be estimated by the amplitude of voltage-sensitive dye signals. A maximum response evoked by an electrical shock around the tip of a stimulating electrode was used as a standard of “maximum activation” of a neuron population. We assume that the majority of neurons near the electrode were simultaneously (within 1 msec) activated, orthodromically or antidromically, by the shock. The amplitude of the optical signal of a dynamic ensemble was measured from the same field of view and was 5–7% of the amplitude of maximum activation. This suggested a low-firing probability within a dynamic ensemble.

The low-firing probability in a dynamic ensemble is further confirmed by extracellular, multiunit recordings that directly



**Figure 7.** Propagating activation during the oscillations. *A*, The recording arrangement for the data in *B* and *C*. *RF*, Rhinal fissure. *B*, Raw data from the LFP electrode and six optical detectors. Three sections from these data (*I*, *II*, *III*) are displayed in the pseudocolor images in *C*. *C*, Pseudocolor images from the three selected sections *I*, *II*, and *III* in *B* displayed at 10 msec/frame. In the images warm colors represent a larger depolarization. *I*, During each cycle of oscillation, one dynamic ensemble emerged and traveled across the field of view. Three dynamic ensembles appeared in this section and moved upward across the field of view. When they moved past the LFP electrode, a cycle of oscillation was detected on the electrode (black trace in *B*). *II*, Later (0.5 sec), two dynamic ensembles appeared simultaneously (at the top and bottom of the field of view) and collided near the center. Two sequential collision events occurred during period *II*. These collisions generated the dramatic change in oscillation frequency seen in Figure 6. Activity was quickly annihilated after the collisions. *III*, After the collisions in period *II*, the propagation direction of the ensembles was reversed. The dynamic ensembles emerged from the top of the field of view and traveled downward. In all eight preparations imaged, the oscillations appeared as propagating activation—dynamic ensembles. The physical size of dynamic ensembles was estimated by the isopotential contour lines in the pseudocolor display and was ~0.5 mm wide in the tangential direction.

measure the spiking density within a dynamic ensemble. One electrode can usually distinguish spikes from two to three neurons near its tip by the amplitude of the spikes. When a dynamic ensemble was passing by the region of the electrode, the neurons sampled by the electrode only fired one to three spikes [from two preparations, 50+ dynamic ensembles (J.-Y. Wu and L. Guan, unpublished observation)]. The same electrode could record many more spikes during the synchronous spike occurring at the beginning of an oscillation epoch. This showed that the electrode was able to detect the spikes from many neurons but only a few fired during each dynamic ensemble.

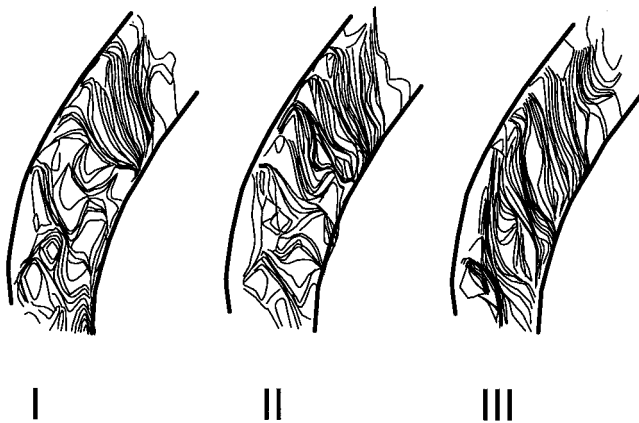
From limited multiunit recordings, we did not find spikes correlated in a millisecond time domain even though the two neurons were physically close to each other. This is consistent with the optical images because correlated firing of a large number of

neurons in the millisecond domain would have generated a sharp spike in the optical signal.

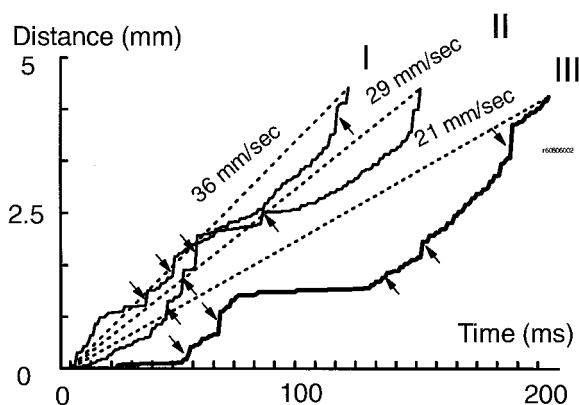
#### Distinguishing dynamic ensembles from epileptiform events

We attempted to use voltage-sensitive dye imaging to distinguish dynamic ensembles from epileptiform events. The synchronous spike at the beginning of each oscillation epoch (Fig. 2*B*) had characteristics of epileptiform events in brain slices bathed in low [Mg] (Anderson et al., 1986; Zhang et al., 1996). This allowed us a direct comparison of the two different events in the same preparation. The synchronous spikes and dynamic ensembles appeared different in several ways. First, the synchronous spike appeared in the imaging as a spreading of an excitation wave, starting from a small initiation site and expanding to the entire

## A. Front edge contour for three ensembles



## B. Propagating velocity



**Figure 8.** Details of the propagation of dynamic ensembles. *A*, The front edge contours of three dynamic ensembles are shown every 2 msec. The wide gaps between the contour lines suggest a sudden increase in propagating velocity. *I*, *II*, Two consecutive dynamic ensembles jumped at similar (but not identical) locations. *III*, A dynamic ensemble that emerged 1 sec later had quite different jump locations. *B*, The propagation velocity of the three dynamic ensembles (measured at cortical layer II) is shown. The overall velocities are 36, 29, and 21 mm/sec, composed of jumps (arrows) and slow propagation. We have examined the front edge contours from 10 dynamic ensembles from three preparations; the dynamically distributed jumps were seen in all 10.

preparation (Fig. 9*B*, top four images). This pattern is similar to the imaging of epileptiform events described previously (Sutor et al., 1994; Albowitz and Kuhnt, 1995; Jackson and Scharfman, 1996; Tsau et al., 1998, 1999). In contrast, the activation of a dynamic ensemble was cohesive (not expanding), its physical size remaining relatively constant during propagation (Figs. 7*C*, 9*B*, bottom row). Second, the amplitude of the dye signal of the synchronous spike was 5–20 times larger than that of dynamic ensembles (Fig. 9*A*), suggesting that the former had a higher density of active neurons. This is consistent with the extracellular electrode recordings, which showed that a synchronous spike had many more spikes than did a dynamic ensemble (data not shown). Third, epileptiform events generated a large peak in the intrinsic optical signal that was slow and wavelength independent (Fig. 9*A*, right) (see also Grinvald et al., 1986; Hochman et al., 1995; Yuste et al., 1997; Pazdalski et al., 1998). With dynamic ensembles this

intrinsic signal was much smaller. Finally, epileptiform events and dynamic ensembles had different refractory periods. The former could not be evoked by an electrical stimulus applied at a rate higher than four times per minute, and the latter occurred at 5–8 Hz at 26°C.

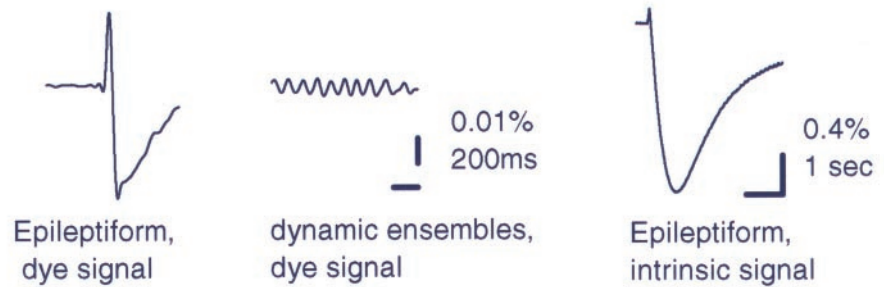
## Evoking dynamic ensembles in normal ACSF

We attempted to show that a dynamic ensemble is an activity related to the intrinsic cortical circuits rather than a phenomenon only appearing during low [Mg] oscillations. We attempted to evoke dynamic ensembles when the cortical slice was bathed in normal ACSF, in which the neuronal network is not in a hyperexcitable condition. It is well known that in normal ACSF, the duration of a “passive” evoked response in the cortical slice is short [5–20 msec (Chagnac-Amitai and Connors, 1989)] and not self-sustaining. We propose, however, that with different ways of activating the cortical network, the excitation and inhibition balance may be altered, which would sustain the long-lasting activations of dynamic ensembles.

We used two kinds of electrical stimulation to evoke population activation in slices bathed in normal ACSF: (1) a conventional thin coaxial electrode, which evoked a high density of coactivation in a small area and which almost always elicited passive short-lasting evoked responses, and (2) a large tip single-pole electrode placed 100–300  $\mu\text{m}$  above the tissue surface, which produced an electrical field over a large area, including all cortical layers (Fig. 10*A*). With this single-pole electrode, the area of activation and the density of activated neurons can be independently adjusted by changing the distance between the electrode and the tissue surface and by using different stimulus intensities. We can thus obtain a low level of activation (0.3–3 times the LFP response threshold) in a large area (1–2 mm in diameter). With this electrode and proper stimulus intensities, the evoked activity developed into long-lasting, propagating activations. In four preparations we delivered weak electrical stimuli at an interstimulus interval of 30 sec, and an LFP recording was used to detect the long-lasting population events (Fig. 10*B*, traces *II*, *III*). The optimum stimulus intensity was  $\sim 0.5$ – $0.7$  of the passive LFP response threshold. With an optimum stimulation intensity,  $>90\%$  of the stimuli evoked these long-lasting events. Increasing the stimulus intensity dramatically reduced the probability of inducing dynamic ensembles. When the intensity was increased by 20%, only  $\sim 40\%$  of evoked activations became long-lasting; other responses were brief (Fig. 10*B*, trace *I*). Although the duration of the events varied from trial to trial, it was obvious that the activity was an all-or-none event. These data suggested that the long-lasting events are self-sustained population activations, which can be evoked in a cortical network with unaltered excitability.

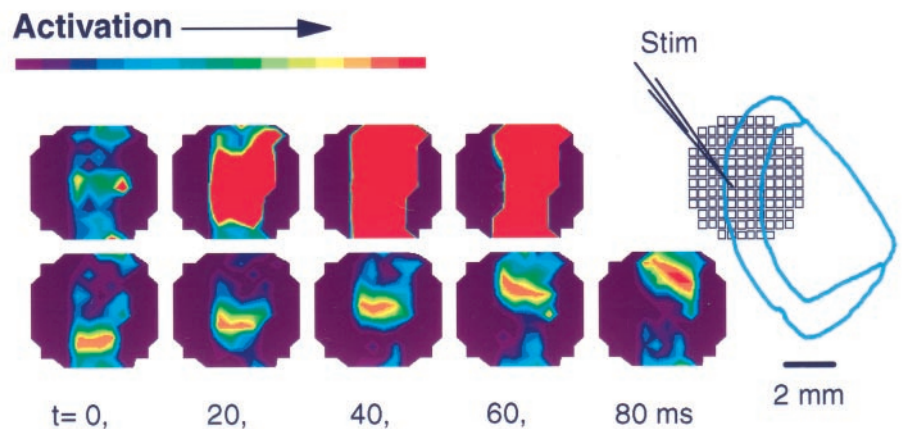
Optical imaging confirmed that these long-lasting events were similar to the dynamic ensembles in their size, shape, and active neuron density (Fig. 10*C*). Optical imaging also showed that the long-lasting activity always emerged in a location 1–2 mm away from the center of the stimulation point and in most of the cases 20–50 msec after the stimulus. This further suggested that the long-lasting events were dynamically organized by the cortical network and not a passive activation resulting from the electrical stimulus. On the basis of these observations, we extend our definition of dynamic ensembles to include both spontaneous and evoked population events sustained by the intrinsic cortical circuits that are dynamically organized and self-sustaining and have a low density of active neurons.

## A. Difference in optical signals



**Figure 9.** Dynamic ensembles are different from epileptiform events. *A, Left, Middle.* The epileptiform event was 10 times larger in dye signal amplitude (705 nm absorption) than were the dynamic ensembles (which are seen as an oscillation in one detector). Intrinsic optical signal also appears at 705 nm, but with a much slower temporal course. We used high-pass filters (both analog and digital, with the RC time constant of 0.1 sec) to emphasize the voltage-sensitive dye signals in these two panels. *Right.* The intrinsic optical signal of an epoch is shown. The signal is much larger and slower than are those on the left and in the middle. These were the same data shown on the left but displayed with a minimum high-pass filter and different time and gain scales. We examined three preparations in white light in which voltage-sensitive dye signals were undetectable. In these conditions the intrinsic optical signals were the same as that shown on the right. *B, Top row.* An epileptiform event was evoked by an electrical stimulus (*Stim*) at the center of the field of view (see diagram on the right). This event expanded to the entire field of view in ~30 msec. *Bottom Row.* A dynamic ensemble emerged from the same cortical area (same field of view, 7 sec later) and remained cohesive for at least 80 msec. The two rows are displayed with different pseudocolor scales to show the propagation of both events.

## B. Difference in propagation pattern



## DISCUSSION

In this report we have characterized the population activation during 7–10 Hz oscillations. We also found that similar activity could be elicited in conditions of normal excitability (slice in normal ACSF). We name this dynamically organized, self-sustaining, and propagating activation dynamic ensembles.

### Dynamic ensembles and population oscillations

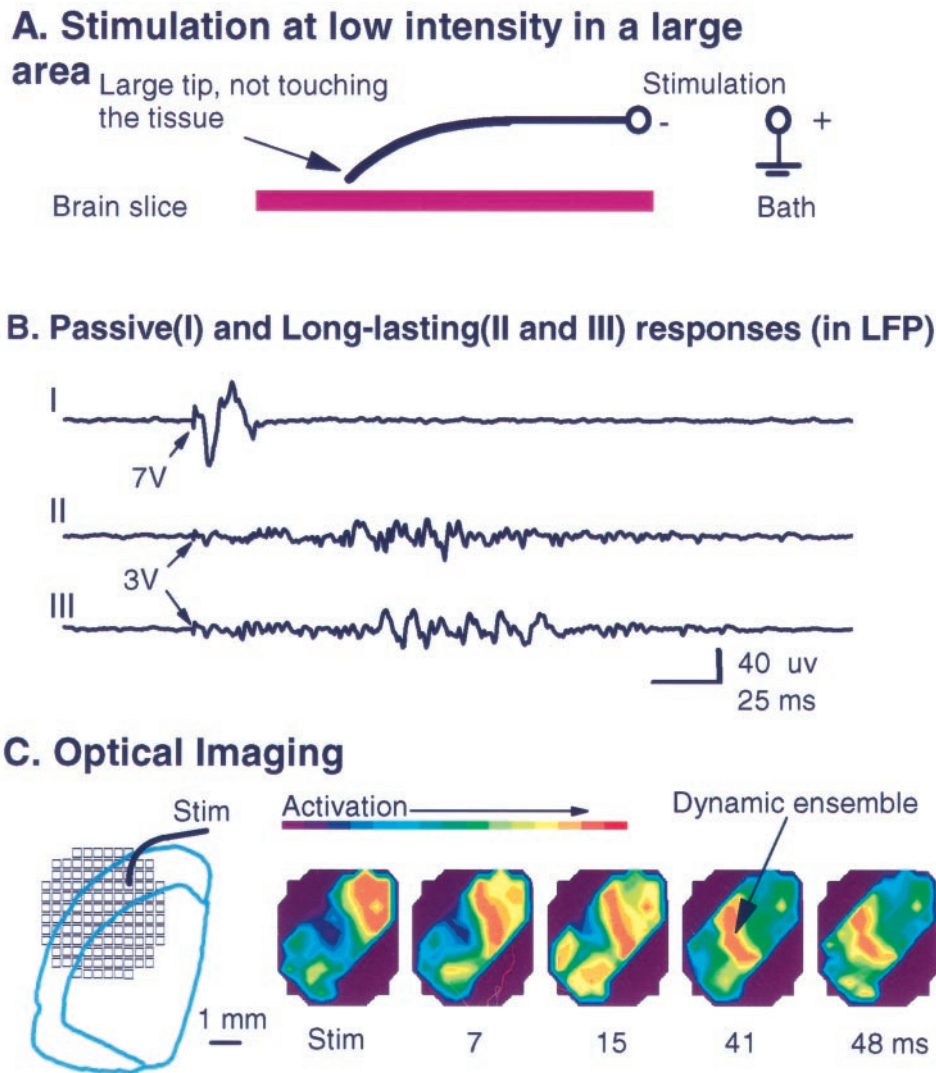
We have demonstrated that the same population events can be described either as oscillations at an individual location (measured optically or with electrodes) or as propagating dynamic ensembles (measured by imaging) (Figs. 7, 10*B*). The ubiquitous existence of oscillations (Tank et al., 1994; Singer and Gray, 1995) and the link between population oscillations and dynamic ensembles raise the possibility that dynamic ensembles might be a common form of population event in the neocortical neuronal network.

Slice preparations from different parts of the brain often have an oscillation frequency that is similar to those found *in vivo*. These oscillations include 6–10 Hz spindles in the thalamus (Bal et al., 1995; Kim et al., 1995), 7–10 Hz in somatosensory cortex (Silva et al., 1991) (this paper), 20–80 Hz in auditory cortex (Metherate and Cruikshank, 1999), 10–50 Hz in other parts of the neocortex (Llinas et al., 1991), and 40 Hz in the hippocampus (Whittington et al., 1995). In a few slice preparations, the spatio-temporal patterns of oscillations have been examined. Field potential measurements from ferret thalamus slices indicated that the 5–10 Hz spindle oscillations also appear as propagating waves

(Kim et al., 1995). These thalamic spindle waves had a similar frequency but propagated 10 times slower than did the dynamic ensembles we have seen in the cortex. Because propagation velocity determines the wavelength or distance between dynamic ensembles, it would be interesting to compare the images of oscillation waves generated by different mechanisms, e.g., “lurching” thalamic spindle versus “continuous” cortical waves (Golomb et al., 1996; Rinzel et al., 1998). This would allow us to examine whether dynamic ensembles are of more general occurrence in different local circuits.

Imaging of *in vivo* oscillations has shown that they also appeared as propagating excitation waves (Delaney et al., 1994; Kleinfeld et al., 1994; Lam, 1997; Precht et al., 1997). The rostral oscillation in turtle olfactory bulb has a simple propagating pattern (Y.-W. Lam, L. B. Cohen, M. Wachowiak, and M. R. Zochowski, unpublished observations), which is strikingly similar to our *in vitro* observations. At each cycle of the rostral oscillation, a “blob” of activation emerged from one location and propagated with a certain velocity and direction in the bulb. The physical size, active neuron density, and propagation velocity of the blob were very similar to those of the dynamic ensembles that we observed in cortical slices. Visually induced oscillations in turtle cortex have multiple frequency domains, with propagating blobs representing each frequency domain (Precht et al., 1997). Oscillations in the somatosensory cortex of behaving rats (7–10 Hz) are also organized in epochs during whisking, and the oscillations also appear as traveling waves (Nicolelis et al., 1995). Unfortunately, descriptions of the spatial pattern of *in vivo* oscil-





**Figure 10.** Dynamic ensembles evoked in normal ACSF. *A*, Schematic diagram of a single-pole electrode placed above the slice is shown. This stimulation method can evoke a low density of activated neurons distributed in a large area including several cortical layers. The intensity and the size of the area can be independently controlled by adjusting stimulation voltage and the distance between the electrode and the slice. *B*, Long-lasting response in local field potential evoked by weak electrical shocks is shown. In this preparation an electrode was placed on top of layer IV,  $\sim 80 \mu\text{m}$  above the preparation. The interstimulus interval was 30 sec. *Trace I*, With stimulus intensity of 7 V (duration = 0.05 msec, for all stimuli), just above the threshold of the passive LFP response, a passive response (short and more synchronized) was evoked. *Traces II, III*, With intensity reduced to 3 V, long-lasting activity was evoked. The long-lasting responses were all-or-none events. In this preparation, 98% ( $n = 120$ ) of 3 V stimuli evoked a long-lasting response. When the stimulus was increased to 5.5 V,  $\sim 60\%$  of the stimuli failed to evoke a long-lasting response. At 7 V, all the stimuli ( $n = 20$ ) evoked the passive response (*trace I*). *Traces II and III* are consecutive trials, illustrating the variation of the long-lasting response. *C*, The long-lasting response was spatially confined (not expanding) and propagating, similar to dynamic ensembles (Fig. 7). A weak electrical shock ( $\sim 10 \text{ V} \times 0.05 \text{ msec}$ ) from an electrode placed  $100 \mu\text{m}$  above the preparation [at the upper right corner of the image field (see diagram on the left)] induced activity over a large area including all layers (*Stim image on the left*). Seven milliseconds later, a long-lasting, propagating activation emerged next to the area activated by the stimulus (*second image from the left*). This activity had an amplitude that was similar to that of the dynamic ensembles observed during 7–10 Hz oscillations (e.g., Fig. 7). The activity remained cohesive for  $\sim 40$

msec and propagated downward in the image field. The slice was perfused in normal ACSF, and the optical signal had a small amplitude compared with that of an epileptiform spike. Note the propagation velocity was  $\sim 6$ – $10 \text{ mm/sec}$  ( $n = 10$ , measured from 2 preparations), significantly slower than the propagation during oscillations. *Stim*, Stimulus.

lations have been limited because no imaging data from mammalian cortex have been available.

Going from an intact cortex to a cortical slice is a significant reduction from a large three-dimensional cortical network to a relatively small and virtually two-dimensional network. This reduces the number of possible initiators of oscillations and the connections between them. Thus a propagating wave would encounter fewer interactions with activation initiated by other initiators. The propagation of spindle waves in thalamic slices (Kim et al., 1995) might have much fewer collisions than in the intact thalamus (Contreras et al., 1997). Thus our images of dynamic ensembles might only present a simplified situation of oscillatory events *in vivo*.

Population oscillations have been proposed to serve as a timing signal (von der Malsburg, 1985; Singer, 1993), as reference-carrying waves for temporal coding (Hopfield, 1995) or for arousal or attention (Steriade et al., 1991; MacKay and Mendonca, 1995; Munk et al., 1996). Our data showed that during oscillations different locations were activated sequentially as dynamic ensembles propagated. This presents questions about the

definition of synchrony in the hundred millisecond time domain and how such synchrony is used for cortical processing.

### Dynamic ensembles and cortical neuronal organization

The interaction between cortical cellular organization (e.g., columns) and propagating dynamic ensembles remains to be explored. Our data suggested that the correlation within a cortical column is better than that between neighboring columns (Fig. 5). It might be speculated that a dynamic ensemble consists of activity in one or several cortical columns and that this activity jumps from one column to another. However, we noticed that the propagation patterns of the dynamic ensembles (Fig. 8) were not consistent with the speculation that dynamic ensembles jump from barrel to barrel. Also, in tangential slices from the somatosensory areas, we could not find a correlation between the activity of dynamic ensembles and the whisker barrels (Wu and Guan, unpublished observation). [Chervin et al. (1988) also found no correlation between barrel patterns and propagation jumps.] These observations suggested that dynamic ensembles

were more likely to be organized by intrinsic (local) circuits rather than by afferent-related cellular organizations.

The organization of intrinsic circuits in the neocortex is relatively similar in different areas (Shepherd, 1988; Douglas and Martin, 1990). This similarity [the “canonical” circuit (Douglas and Martin, 1990)] suggests universal rules for organizing local population activation in different cortices. As a continuation of this work, it would be interesting to compare oscillations and evoked population events in different cortical areas, to test whether dynamic ensembles can be evoked in all cortical tissues.

### The nature of activation in a dynamic ensemble

We estimate that a dynamic ensemble has only ~5–7% of the maximum activation of the cortical network. The voltage-sensitive dye signals come from a combination of spikes and synaptic potentials. If we assume that our signals were a linear summation of spikes and synaptic potentials in the population (Wu and Cohen, 1993), then one spike (100 mV) of 1 neuron would contribute equally with 10 neurons that have a 10 mV depolarization. At one extreme, we could assume that all signals were contributed by spikes, so that 5–7% of the total population was firing simultaneously in each sampling period. Thus during the 200 msec duration of a dynamic ensemble, each activated neuron would fire ~10 times. However, extracellular unit recordings revealed that each neuron only fired one or two spikes. Therefore it seems likely that subthreshold depolarizations of many cells are an important contributor to the dynamic ensemble signals. We thus speculate that a dynamic ensemble is organized as a domain of moderately depolarized neurons in which some neurons will increase their firing probability to reach one to two spikes per event. This kind of organization is similar to the spontaneous population coactivation observed in developing cortex (Yuste et al., 1992) and retina (Wong et al., 1995; Feller et al., 1997).

### Self-sustainability: functional significance

The activation of dynamic ensembles lasted much longer than an ordinary evoked response. We considered this activation to be self-sustaining, i.e., sustaining a certain level of activity in the cortex without afferent input. We hypothesize that self-sustainability is a population property of a reciprocally activating neuronal network. The development of a dynamic ensemble might follow certain population rules, e.g., depend on the size of the activation, the density of active neurons in the area, and the temporal pattern of the input (cf. Golomb and Amitai, 1997). In this report we evoked long-lasting activity by activating a low density of neurons in a large area involving all cortical layers (Fig. 10). The all-or-none nature of a population response supports our speculation that the activity is self-sustained. This kind of long-lasting evoked response can also be evoked by stimulating thalamocortical afferents (Metherate and Cruikshank, 1999).

How dynamic ensembles interact with evoked events remains to be explored. In intact brain, dynamic ensembles may be evoked by sensory input or emerge during evoked oscillations. This would allow spatial interactions between different ensembles and temporal interactions between concurrent input and dynamic ensembles evoked by previous input. Self-sustained activity in dynamic ensembles may provide an uneven local excitability, in which the same input may elicit a higher (or lower) population firing rate within an ensemble than in the regions outside of it. This suggests a mechanism for cortical reverberation (Hebb, 1949; Amit, 1995), a process in which a local network can amplify a weak input with a compatible spatiotemporal pattern and reject a strong input with incompatible patterns.

The existence of dynamically organized population modules has been postulated as a “cell assembly” (Hebb, 1949; Amit, 1995; Nicolelis et al., 1997) that reverberates with afferent input and may play a role in sensory discrimination (Zhang, 1996), temporary information storage, attentional modulation of sensory processing, and transition from short- to long-term plasticity (Rauschecker, 1995). Our study of dynamic ensembles suggests that cortical intrinsic circuits can serve as a host for self-sustained and dynamically organized population activity. Such a form of activity could be used to “bind” afferent input distributed in time and space.

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