

# Changes in Expression and Function of Extrasynaptic GABA<sub>A</sub> Receptors in the Rat Hippocampus during Pregnancy and after Delivery

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Pregnancy is associated with changes in mood and anxiety level as well as with marked hormonal fluctuations. Increases in the brain concentrations of neuroactive steroids during pregnancy in rats are accompanied by changes in expression of subunits of the GABA type A receptor (GABA<sub>A</sub>-R) in the brain. Granule cells of the dentate gyrus (DGGCs) exhibit two components of inhibitory GABAergic transmission: a phasic component mediated by synaptic GABA<sub>A</sub>-Rs, and a tonic component mediated by extrasynaptic GABA<sub>A</sub>-Rs. Recordings of GABAergic currents were obtained from hippocampal slices prepared from rats in estrus, at pregnancy day 15 (P15) or P19, or at 2 d after delivery. Exogenous GABA or 3 $\alpha$ ,5 $\alpha$ -THP induced an increase in tonic current in DGGCs that was significantly greater at P19 than in estrus. Neither tonic nor phasic currents were affected by pregnancy in CA1 pyramidal cells. Immunohistochemical analysis revealed a marked increase in the abundance of the  $\delta$  subunit of the GABA<sub>A</sub>-R and a concomitant decrease in that of the  $\gamma_2$  subunit in the hippocampus at P19. Expression of the  $\alpha_4$  subunit did not change during pregnancy but was increased 2 d after delivery. Treatment of rats from P12 to P18 with the 5 $\alpha$ -reductase inhibitor finasteride prevented the changes in tonic current and in  $\delta$  and  $\gamma_2$  subunit expression normally apparent at P19. These data suggest that the number of extrasynaptic GABA<sub>A</sub>-Rs is increased in DGGCs during late pregnancy as a consequence of the associated marked fluctuations in the brain levels of neuroactive steroids.

**Key words:** GABA<sub>A</sub> receptor  $\delta$  subunit; pregnancy; delivery; 3 $\alpha$ -hydroxy-5 $\alpha$ -pregnan-20-one; dentate gyrus granule cells; tonic current

## Introduction

Fast inhibitory synaptic transmission in the CNS is mediated predominantly by GABA type A receptors (GABA<sub>A</sub>-Rs). These ligand-gated Cl<sup>−</sup> channels are characterized by a high level of structural (subunit composition) and functional (agonist affinity, channel kinetics) heterogeneity (Barnard et al., 1998; Whiting et al., 1999; Sieghart and Sperk, 2002; Jacob et al., 2008) and are also the principal target of endogenous neuroactive steroids (Belelli and Lambert, 2005; Mitchell et al., 2008) as well as of anxiolytic, hypnotic, myorelaxant, anticonvulsant, and anesthetic drugs (Sieghart, 1995; Barnard et al., 1998).

Whereas synaptic GABA<sub>A</sub>-Rs mediate a phasic inhibitory current, extrasynaptic GABA<sub>A</sub>-Rs with a high affinity for GABA and nondesensitizing properties are activated by the low concentrations of GABA normally present in the extracellular space and mediate a tonic inhibitory current (Brickley et al., 1996; Semyanov et al., 2004; Farrant and Nusser, 2005). Synaptic GABA<sub>A</sub>-Rs are pentamers consisting of combinations of  $\alpha_n$ ,  $\beta_n$ , and  $\gamma_2$  sub-

units, whereas extrasynaptic GABA<sub>A</sub>-Rs are pentamers characterized by the presence of the  $\delta$  subunit together either with the  $\alpha_4$  subunit in granule cells of the dentate gyrus (Pirker et al., 2000; Nusser and Mody, 2002; Wei et al., 2003) and in thalamic neurons (Sur et al., 1999), with the  $\alpha_6$  subunit in cerebellar granule cells (Jones et al., 1997; Nusser et al., 1998; Pirker et al., 2000), or with the  $\alpha_1$  subunit in a small population of dentate gyrus interneurons (Glykys et al., 2007). Additionally, extrasynaptic  $\alpha_5$ -containing GABA<sub>A</sub>-Rs mediate tonic current in CA1/CA3 pyramidal cells (Pirker et al., 2000; Caraiscos et al., 2004), and ~30% of the total tonic current in the dentate gyrus (Glykys et al., 2008).

Neurosteroids are steroid derivatives that are synthesized both locally in neurons and glia as well as peripherally in the gonads and adrenals (Hu et al., 1987; Mathur et al., 1993; Mellon and Griffin, 2002; Sanna et al., 2004). Among these compounds, the progesterone metabolite 3 $\alpha$ -hydroxy-5 $\alpha$ -pregnan-20-one (3 $\alpha$ ,5 $\alpha$ -THP or allopregnanolone) exerts a selective and potent modulatory action at GABA<sub>A</sub>-Rs and exhibits an especially high affinity and efficacy at extrasynaptic GABA<sub>A</sub>-Rs (Majewska et al., 1986; Belelli and Lambert, 2005). Prolonged physiological or pharmacologically induced fluctuations in brain and plasma concentrations of 3 $\alpha$ ,5 $\alpha$ -THP play an important role in regulation of GABA<sub>A</sub>-R plasticity and function (Brussaard et al., 1997; Smith et al., 1998a,b; Biggio et al., 2006; Mostallino et al., 2006) as well as in development of the GABAergic system in the prefrontal cortex

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(Grobin et al., 2003). Fluctuations in neurosteroid levels during pregnancy (Brussaard et al., 1997; Concas et al., 1998) and the ovarian cycle (Maguire et al., 2005; Maguire and Mody, 2007) thus result in selective changes in the expression and function of GABA<sub>A</sub>-Rs in various regions of the rat and mouse brain, effects that are mediated by a direct regulatory action of neurosteroids at GABA<sub>A</sub>-Rs. We have now shown that the expression and function of extrasynaptic GABA<sub>A</sub>-Rs in the rat hippocampus are markedly modified during pregnancy and after delivery.

## Materials and Methods

**Animals.** Adult (60–90 d old; body mass, 200–250 g) female Sprague Dawley CD rats (Charles River Laboratories) were studied. They were housed under an artificial 12 h light/dark cycle (lights on from 8:00 A.M. to 8:00 P.M.) and at a constant temperature of  $23 \pm 2^\circ\text{C}$  and relative humidity of 65%. Standard laboratory food and water were freely available at all times.

The specific stage of the estrous cycle (diestrus, proestrus, or estrus) was determined from analysis of daily vaginal smears collected between 9:00 A.M. and 10:00 A.M. for 2–4 weeks. The estrous cycle was monitored for two full cycles before animals were mated by placing them individually in a cage with a proven male on the evening of proestrus. The male was removed from the breeding cage after the appearance of a vaginal plug in the female; the day the plug was detected was designated day 0 of pregnancy (P0). Animals were killed between 9:00 A.M. and 10:00 A.M. when in estrus (control group) or at various stages of pregnancy or after delivery.

Animal care and handling throughout the experimental procedures were in accordance with the European Communities Council Directive of 24 November 1986 (86/609/EEC). The experimental protocols were also approved by the Animal Ethics Committee of the University of Cagliari.

**Pharmacological treatments.** In one series of experiments, pregnant rats were treated with finasteride (25 mg per kilogram of body mass) once a day from P12 to P18 (Concas et al., 1998). Finasteride was extracted and purified from commercial tablets as described previously (Trapani et al., 2002), suspended in a mixture of ethanol (20%) and corn oil (80%), and injected subcutaneously at the nape of the neck in a volume of 3 ml/kg. In another series of experiments, clomiphene citrate (Bruno Farmaceutici) was dissolved in physiological saline and administered intragastrically at a dose of 5 mg/kg once a day from P12 to P18. Control pregnant rats received the respective vehicle according to the same schedule as that for the two drugs.

**Immunohistochemistry.** Rats were administered a lethal dose (0.3 ml/g) of Equithesin (1 g of sodium pentobarbital, 4.251 g of chloral hydrate, 2.125 g of MgSO<sub>4</sub>, 12 ml of ethanol, and 43.6 ml of propylene glycol, adjusted to a total volume of 100 ml with distilled water) and were then perfused through the ascending aorta with 100 ml of PBS followed by 250 ml of 4% paraformaldehyde in PBS. The brain was removed, exposed to the same fixative for 2 h, and then transferred to 20% (w/v) sucrose in PBS. Sagittal sections (thickness, 50  $\mu\text{m}$ ) were cut with a Vibratome 1000 Plus instrument (Vibratome) and stored in antifreeze solution (30% ethylene glycol, 20% glycerol, 50% 50 mM phosphate buffer) at  $-20^\circ\text{C}$ .

Indirect immunohistochemistry for subunits of the GABA<sub>A</sub>-R was performed with free-floating sections. The sections were washed with PBS, incubated for 30 min at room temperature with 0.1% phenylhydrazine to block endogenous peroxidase activity, permeabilized for 1 h with 0.2% Triton X-100 in PBS (PBS-T), and incubated for 1 h with 10% normal donkey serum (Jackson ImmunoResearch) in PBS-T. They were then incubated at  $4^\circ\text{C}$  for 4 d with goat polyclonal antibodies to the  $\alpha_4$  or  $\delta$  subunits (Santa Cruz Biotechnology) or with rabbit polyclonal antibodies to the  $\gamma_2$  subunit (Alomone) at dilutions of 1:500, 1:100, or 1:250, respectively, in PBS-T containing 10% normal donkey serum. After several washes, the sections were incubated at room temperature first for 2 h with appropriate biotinylated donkey antibodies to IgG (Jackson ImmunoResearch) diluted 1:200 in PBS-T and then for 30 min with avidin-peroxidase solution (Vectastain Elite Kit; Vector Laboratories), with the sections being washed three times with PBS-T after each incubation. The

reaction product was visualized by exposure of the sections to 0.4 mM 3,3'-diaminobenzidine (Sigma) and 0.01% H<sub>2</sub>O<sub>2</sub>. After several washes with PBS, the sections were mounted on gelatin-coated slides, air-dried, dehydrated in ethanol, and cleared in xylene, and a coverslip was then applied in the presence of Eukitt mounting medium (O. Kindler). Control sections either incubated with primary antibodies in the presence of the corresponding peptide antigen or not exposed to primary antibodies did not yield positive staining. Sections were examined with a BX-41 microscope (Olympus) and photographed with an F-View charge-coupled device camera. Representative images obtained with a Plan 2 $\times$  objective (numerical aperture, 0.05) are shown.

Semiquantitative analysis of images was performed with AnalySIS 3.2 software (Soft Imaging System). In each image, the different areas of the hippocampus, based on plates 48–50 of the Paxinos and Watson (1986) atlas, were selected by drawing a line surrounding the region of interest (ROI). The intensity of immunostaining in each ROI was determined as an integral from the intensity value for each pixel and the total number of pixels and was corrected for the background obtained in negative control sections. The intensity values, which represent the abundance of the corresponding GABA<sub>A</sub>-R subunit, were represented by a gray scale and expressed as percentage change relative to the control rats in estrus.

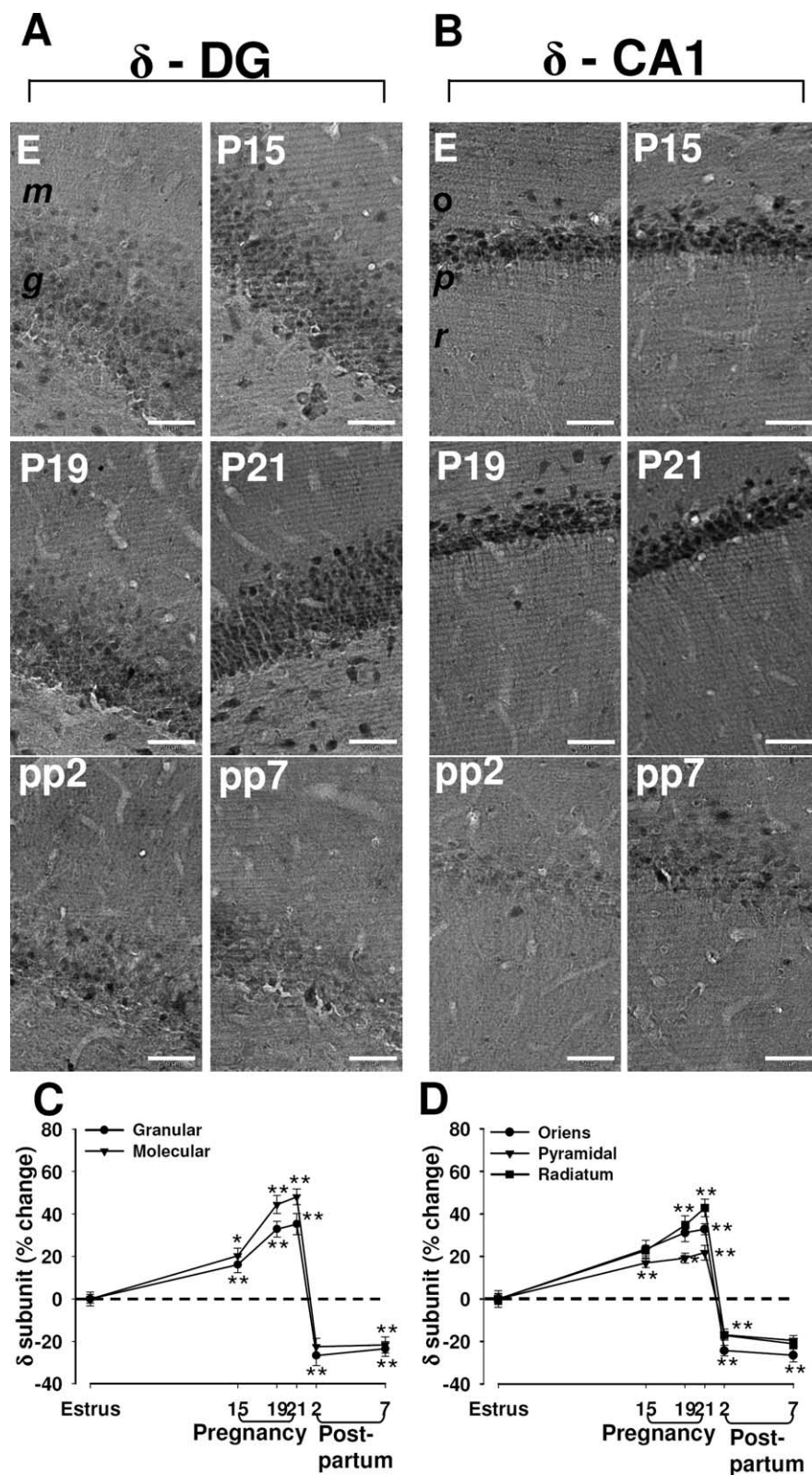
The antibodies to the  $\delta$  subunit were generated in response to a peptide corresponding to the C terminus of the human protein; the amino acid sequence of the peptide differs from that of the corresponding region of the rat protein at two positions. Immunoblot analysis of a crude membrane fraction of the rat hippocampus with these antibodies yielded a single immunoreactive band at a position corresponding to a molecular size of  $\sim 54$  kDa (Follesa et al., 2005). The specificity of the antibody preparation was also confirmed by immunocytochemical analysis (Follesa et al., 2005; Serra et al., 2006). The antibodies to the  $\alpha_4$  subunit were generated in response to a peptide corresponding to the N-terminal region of the rat protein. Immunoblot analysis showed that these antibodies recognized a single protein of  $\sim 70$  kDa in a crude membrane fraction prepared from rat hippocampal neurons (Sanna et al., 2003). The specificity of this antibody preparation was also confirmed by immunocytochemistry (Sanna et al., 2003; Serra et al., 2006). The antibodies to the  $\gamma_2$  subunit were generated in response to a synthetic peptide corresponding to residues 39–53 of the rat protein. Immunoblot analysis of a crude membrane fraction of rat hippocampus with these antibodies yielded a single immunoreactive band of  $\sim 45$ – $47$  kDa; this band was not detected when the blot was incubated with the antibodies in the presence of the peptide antigen, demonstrating antibody specificity (data not shown).

**Electrophysiology.** Rats were anesthetized in a chamber saturated with chloroform and then decapitated. The brain was removed rapidly and placed in an ice-cold solution containing 220 mM sucrose, 2 mM KCl, 1.3 mM NaH<sub>2</sub>PO<sub>4</sub>, 12 mM MgSO<sub>4</sub>, 0.2 mM CaCl<sub>2</sub>, 10 mM glucose, 2.6 mM NaHCO<sub>3</sub> (pH 7.3, equilibrated with 95% O<sub>2</sub> and 5% CO<sub>2</sub>), and 3 mM kynurenic acid. Coronal hippocampal slices (thickness, 200–250  $\mu\text{m}$ ) were prepared with a Vibratome 1000 Plus and then incubated first for 40 min at  $34^\circ\text{C}$  and then for 30 min at room temperature in artificial CSF (ACSF), consisting of 126 mM NaCl, 3 mM KCl, 1.25 mM NaH<sub>2</sub>PO<sub>4</sub>, 1 mM MgSO<sub>4</sub>, 2 mM CaCl<sub>2</sub>, 10 mM glucose, and 26 mM NaHCO<sub>3</sub> (pH 7.3, equilibrated with 95% O<sub>2</sub> and 5% CO<sub>2</sub>).

Slices were transferred to a recording chamber perfused with ACSF at a rate of  $\sim 2$  ml/min and at room temperature. Whole-cell patch-clamp electrophysiological recordings were performed with an Axopatch 200-B amplifier (Axon Instruments) and using an infrared-differential interference contrast microscope. Patch microelectrodes (borosilicate capillaries with a filament and an outer diameter of 1.5  $\mu\text{m}$ ; Sutter Instruments) were prepared with a two-step vertical puller (Sutter Instruments) and had a resistance of 4–6 M $\Omega$ . Spontaneous IPSCs (sIPSCs) as well as tonic GABAergic currents were recorded at a holding potential of  $-65$  mV with an internal solution containing 140 mM CsCl, 2 mM MgCl<sub>2</sub>, 1 mM CaCl<sub>2</sub>, 10 mM EGTA, 10 mM HEPES-CsOH, pH 7.3, 2 mM ATP (disodium salt), and 5 mM QX-314 (lidocaine *N*-ethyl bromide). Access resistance was between 20 and 30 M $\Omega$ ; if it changed by  $>20\%$  during the recording, the recording was discarded.

All GABAergic currents were recorded in the presence of kynurenic





**Figure 1.** Changes in immunoreactivity for the  $\delta$  subunit of the GABA<sub>A</sub>-R in the rat hippocampus during pregnancy and after delivery. **A, B**, Representative immunohistochemical images of the distribution of the  $\delta$  subunit in the dentate gyrus (**A**) and in the CA1 region (**B**) in hippocampal sections from rats in estrus (E), at P15, P19, or P21, or at pp2 or pp7. m, Molecular layer of the dentate gyrus; g, granule layer of the dentate gyrus; o, stratum oriens of the CA1 region; p, pyramidal of the CA1 region; r, radiatum of the CA1 region. Scale bar, 50  $\mu$ m. **C, D**, Images similar to those in **A** and **B** were subjected to semiquantitative measurement of  $\delta$  subunit immunoreactivity in the granular and molecular cell layers of the dentate gyrus (**C**), and in the stratum oriens, pyramidal cell layer, and stratum radiatum of CA1 (**D**). Data are expressed as the percentage change in gray-scale values relative to the corresponding value for rats in estrus (control) and are means  $\pm$  SEM from six to eight animals at each time point. \* $p$  < 0.05, \*\* $p$  < 0.01 versus estrus (one-way ANOVA followed by Scheffé's test).

acid (3 mM) in the external solution. For the recording of miniature IPSCs (mIPSCs), lidocaine (500  $\mu$ M) was added in the external solution. Currents through the patch-clamp amplifier were filtered at 2 kHz and digitized at 5.5 kHz with commercial software (pClamp 8.2, Axon Instruments). Analysis of tonic GABAergic currents was performed with parameters described previously (Carta et al., 2004). In brief, we selected epochs of 3 s every 30 s of recording and visually excluded sIPSCs from the analysis. For the analysis of sIPSCs or mIPSCs, we visually selected all events, which are readily distinguishable from the background noise on the basis of their characteristic fast rise times and slower decay times. All-point histograms were constructed from these epochs and mean values were determined from Gaussian fits. (GABA, bicuculline methiodide, 3 $\alpha$ ,5 $\alpha$ -THP, and Ro15-4513 were from Sigma. Lorazepam was a kind gift from Wyeth).

**Statistical analysis.** Electrophysiological data are presented as means  $\pm$  SEM. Statistical comparisons of pooled data were performed by ANOVA followed by Scheffé's *post hoc* test. Immunohistochemical data are expressed as percentage change in gray-scale values relative to the estrus group and are means  $\pm$  SEM of values from the indicated numbers of animals. Significance of differences was assessed by one-way ANOVA followed by Scheffé's test. For all statistical analysis, a  $p$  value of <0.05 was considered statistically significant.

## Results

### Expression of the $\delta$ subunit during pregnancy and after delivery

The expression of the  $\alpha_4$ ,  $\delta$ , and  $\gamma_2$  subunits of the GABA<sub>A</sub>-R in the hippocampus of rats during pregnancy and after delivery was examined by immunohistochemistry with specific antibodies generated in response to extracellular epitopes of these proteins. In all instances, specific immunostaining was blocked by previous incubation of the antibodies with the respective peptide antigen (data not shown). Staining intensity for the individual subunits depended on the properties of the corresponding antibodies. We therefore used different antibody dilutions to obtain similar intensities of staining. Differences in staining intensity obtained with the subunit-specific antibodies thus do not necessarily reflect real differences in expression level among the respective proteins.

In control (estrus) rats, moderate levels of immunoreactivity for the  $\delta$  subunit of the GABA<sub>A</sub>-R were apparent throughout most regions of the hippocampal formation (Fig. 1; supplemental Fig. S1, available at [www.jneurosci.org](http://www.jneurosci.org) as supplemental material). The abundance of the  $\delta$  subunit appeared greatest in the granule cell layer of the dentate gyrus and in the pyramidal cell layer of regions

CA1 (Fig. 1*A,B*). Expression of the  $\delta$  subunit was also prominent in the molecular layer of the dentate gyrus. Measurement of  $\delta$  subunit immunoreactivity in rats on P15, P19, and P21 revealed that specific staining increased progressively in the dentate gyrus and the CA1 region compared with that in control rats (Fig. 1*A–D*). A significant increase was already evident at P15 in the dentate gyrus and the pyramidal cell layer of CA1, with maximal upregulation in all regions being apparent at P19 or P21. In contrast, the amount of  $\delta$  subunit immunoreactivity had decreased to values significantly less than those for control rats by 2 d after delivery (pp2) (Fig. 1*A–D*). This downregulation of  $\delta$  subunit expression was still apparent 7 d postpartum (pp7) in the granular and molecular layers of the dentate gyrus (Fig. 1*A,C*) and in the stratum oriens of CA1 (Fig. 1*B,D*).

#### Expression of the $\alpha_4$ subunit during pregnancy and after delivery

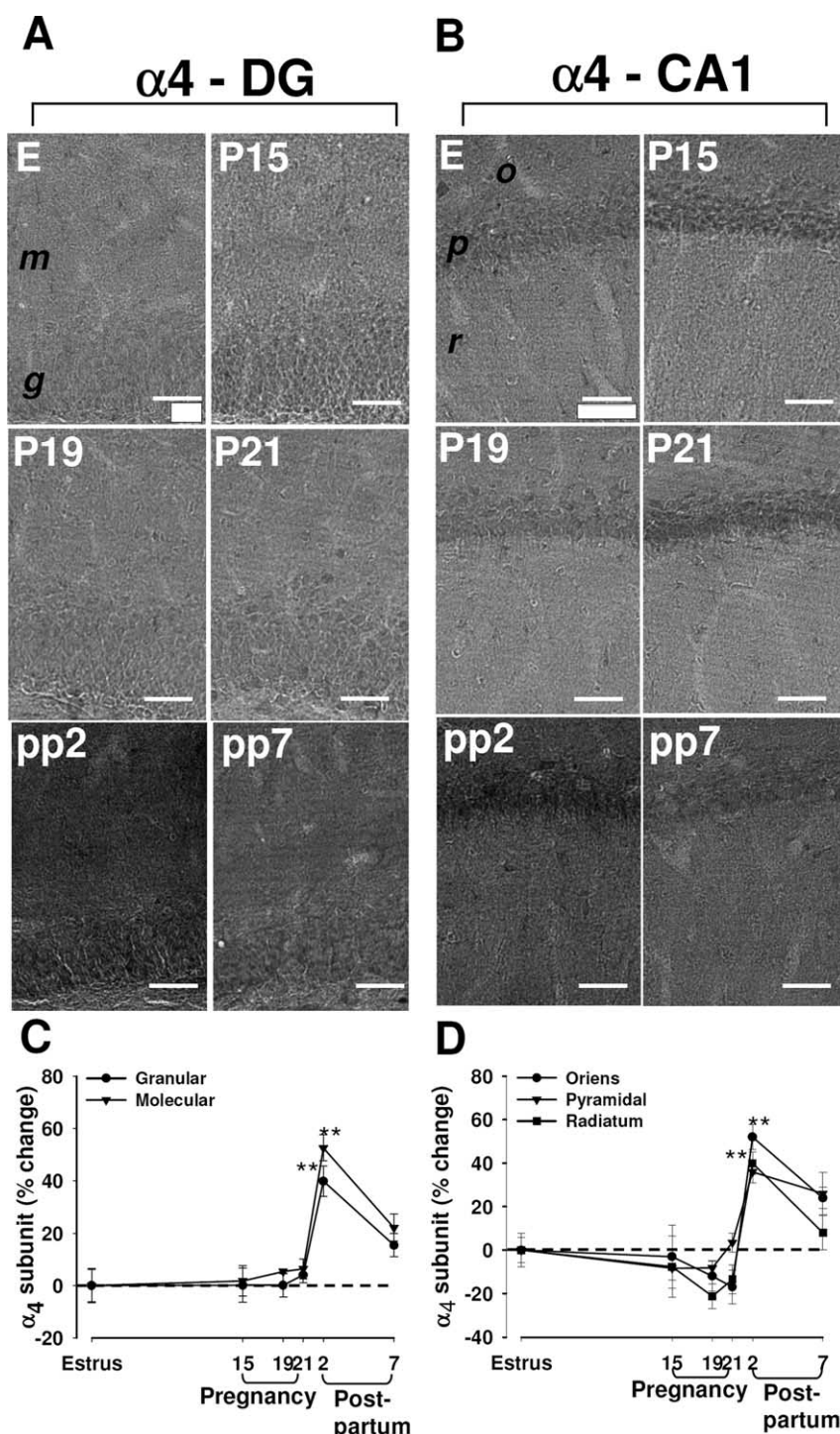
Immunoreactivity for the  $\alpha_4$  subunit of the GABA<sub>A</sub>-R appeared diffuse throughout the hippocampal formation of control rats, being most abundant in the granule cell layer of the dentate gyrus and in the pyramidal cell layer of CA1 (Fig. 2; supplemental Fig. S2, available at [www.jneurosci.org](http://www.jneurosci.org) as supplemental material). Staining was also prominent in the molecular layer of the dentate gyrus. The amount of the  $\alpha_4$  subunit did not change significantly during pregnancy, but it was increased markedly both in the dentate gyrus and the CA1 region at 2 d after delivery (Fig. 2*A–D*). This upregulation of  $\alpha_4$  subunit expression was no longer significant at 7 d after delivery (Fig. 2*A–D*).

#### Expression of the $\gamma_2$ subunit during pregnancy and after delivery

In control rats,  $\gamma_2$  subunit immunoreactivity was concentrated in the strata oriens and radiatum of CA1 and CA3, in the stratum lacunosum-moleculare of CA1, and in granule cells of the dentate gyrus (Fig. 3; supplemental Fig. S3, available at [www.jneurosci.org](http://www.jneurosci.org) as supplemental material). The level of  $\gamma_2$  subunit immunoreactivity was decreased at P15 and P19 in the dentate gyrus and the CA1 region (Fig. 3*A–D*). It then increased immediately before delivery at P21, remained increased at 2 d after delivery, and returned to control values by 7 d after delivery (Fig. 3*A–D*).

#### GABAergic tonic current in granule cells during pregnancy and after delivery

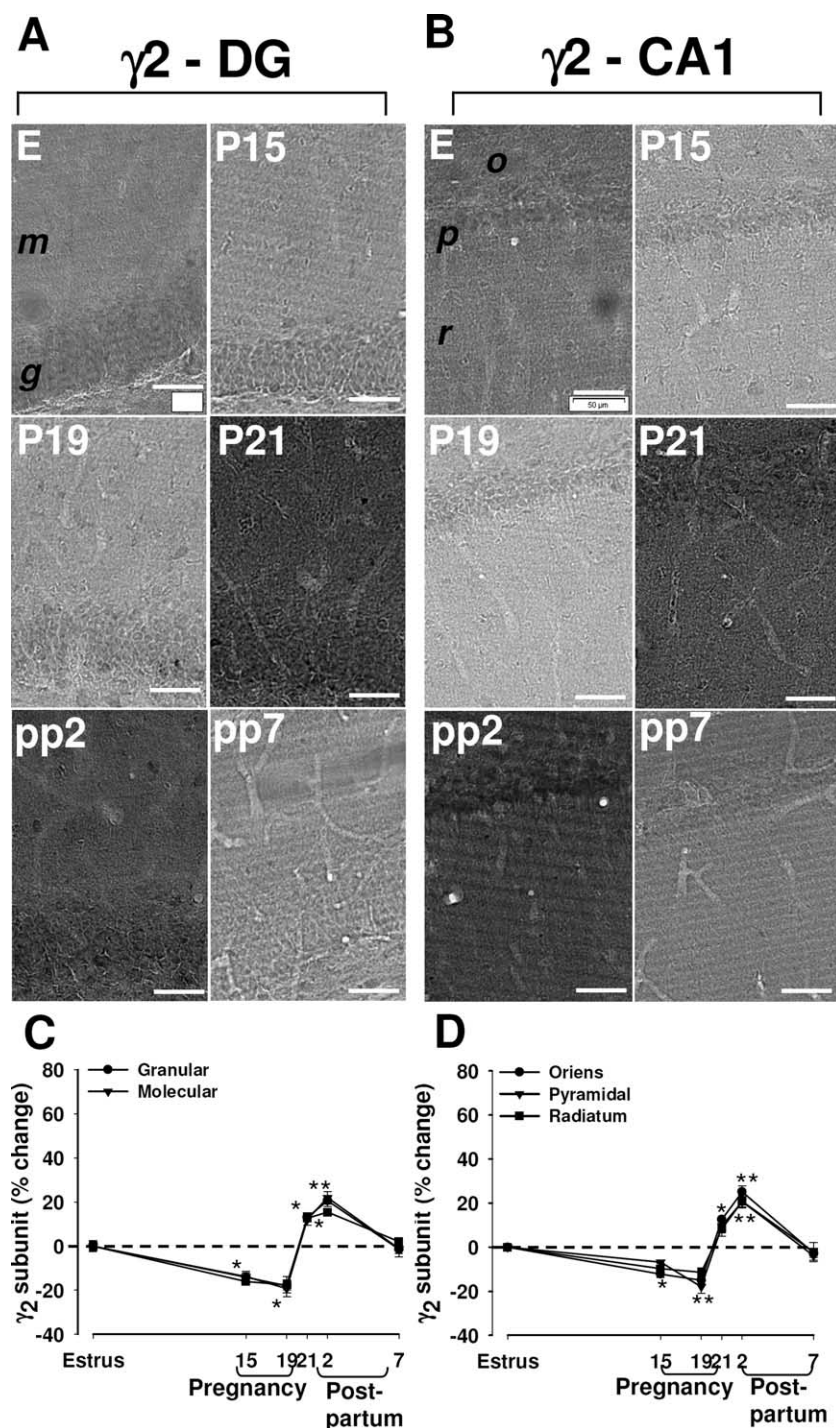
We next examined GABAergic transmission in granule cells of the dentate gyrus using hippocampal slices obtained from rats at



**Figure 2.** Changes in immunoreactivity for the  $\alpha_4$  subunit of the GABA<sub>A</sub>-R in the rat hippocampus during pregnancy and after delivery. *A, B*, Representative immunohistochemical images of the distribution of the  $\alpha_4$  subunit in the dentate gyrus (*A*) and in the CA1 region (*B*) in hippocampal sections from rats in estrus (E), at P15, P19, or P21, or at pp2 or pp7. *m*, Molecular layer of the dentate gyrus; *g*, granule layer of the dentate gyrus; *o*, stratum oriens of the CA1 region; *p*, pyramidal of the CA1 region; *r*, radiatum of the CA1 region. Scale bar, 50  $\mu$ m. *C, D*, Images similar to those in *A* and *B* were subjected to semiquantitative measurement of  $\alpha_4$  subunit immunoreactivity in the granular and molecular cell layers of the dentate gyrus (*C*), and in the stratum oriens, pyramidal cell layer, and stratum radiatum of CA1 (*D*). Data are expressed as the percentage change in gray-scale values relative to the corresponding value for rats in estrus (control) and are means  $\pm$  SEM from six to eight animals at each time point. \*\* $p$  < 0.01 versus estrus (one-way ANOVA followed by Scheffé's test).

P15 and P19 as well as at 2 d after delivery and during estrus (control). Granule cell bodies are densely packed and are readily recognizable on the basis of their rounded shape and small size. In adult rats, granule cells of the dentate gyrus are characterized by a





**Figure 3.** Changes in immunoreactivity for the  $\gamma_2$  subunit of the GABA<sub>A</sub>-R in the rat hippocampus during pregnancy and after delivery. **A, B**, Representative immunohistochemical images of the distribution of the  $\gamma_2$  subunit in the dentate gyrus (**A**) and in the CA1 region (**B**) in hippocampal sections from rats in estrus (E), at P15, P19, or P21, or at pp2 or pp7. m, Molecular layer of the dentate gyrus; g, granule layer of the dentate gyrus; o, stratum oriens of the CA1 region; p, pyramidal of the CA1 region; r, radiatum of the CA1 region. Scale bar, 50  $\mu$ m. **C, D**, Images similar to those in **A** and **B** were subjected to semiquantitative measurement of  $\gamma_2$  subunit immunoreactivity in the granular and molecular cell layers of the dentate gyrus (**C**), and in the stratum oriens, pyramidal cell layer, and stratum radiatum of CA1 (**D**). Data are expressed as the percentage change in gray-scale values relative to the corresponding value for rats in estrus (control) and are means  $\pm$  SEM from six to eight animals at each time point. \* $p$  < 0.05, \*\* $p$  < 0.01 versus estrus (one-way ANOVA followed by Scheffé's test).

prominent GABAergic tonic current mediated by extrasynaptic GABA<sub>A</sub>-Rs containing mainly  $\alpha_4$ ,  $\beta_n$ , and  $\delta$  subunits (Nusser and Mody, 2002) and, in a lower proportion, by extrasynaptic receptors containing the  $\alpha_5$  subunit (Glykys et al., 2008). There is also

a phasic component to inhibition of these neurons that is attributable to the activation of synaptic GABA<sub>A</sub>-Rs with an  $\alpha_n\beta_n\gamma_2$  subunit composition. We recorded from granule cells in the voltage-clamp mode (holding potential,  $-65$  mV) with patch pipettes containing a solution with a high  $\text{Cl}^-$  concentration that gives rise to a  $\text{Cl}^-$  equilibrium potential of  $\sim 0$  mV. Under these recording conditions, activation of GABA<sub>A</sub>-Rs generates inward currents that reflect an outflow of  $\text{Cl}^-$ . GABAergic currents were pharmacologically isolated by the addition to ACSF of kynurenic acid (3 mM), a broad-spectrum antagonist of ionotropic glutamate receptors.

After a baseline period of  $\sim 5$  min, slices were exposed to exogenous GABA (5  $\mu$ M) for 5 min to activate high-affinity extrasynaptic GABA<sub>A</sub>-Rs (Maguire et al., 2005). Exposure of granule cells from control rats to GABA increased current noise variance and shifted the holding current in the negative direction with respect to baseline (Fig. 4), effects attributable to the activation of extrasynaptic GABA<sub>A</sub>-Rs. Bath application of bicuculline (30  $\mu$ M), a competitive antagonist of GABA<sub>A</sub>-Rs, blocked all GABA-induced currents, confirming that they were mediated by GABA<sub>A</sub>-Rs, as well as reduced the noise variance and the holding current compared with those observed during the baseline period (Fig. 4A). The bath application of GABA resulted in marked increases in noise variance and in a shift of the holding current in slices obtained from rats at P15, P19, or 2 d after delivery (Fig. 4A–C). Statistical analysis revealed a significant difference between rats at P19 and control animals for both noise variance and holding current. Tonic current parameters were also increased at P15 relative to control values, but the changes were not significant, and they had returned to control values by 2 d after delivery (Fig. 4B, C).

#### GABAergic phasic inhibition in granule cells during pregnancy and after delivery

The high frequency of sIPSCs observed in the electrophysiological traces in Figure 4A showed that the granule cells were also subjected to pronounced phasic inhibition mediated by synaptic GABA<sub>A</sub> receptors. These currents were readily distinguishable from the tonic current on the basis of their fast rise time and slower decay time.

Analysis of the kinetic properties of these sIPSCs revealed no significant differences among rats of the different experimental groups (Table 1). Because sIPSCs may be affected by alterations in spontaneous action potential firing levels of interneurons, we also recorded mIPSCs in dentate gyrus

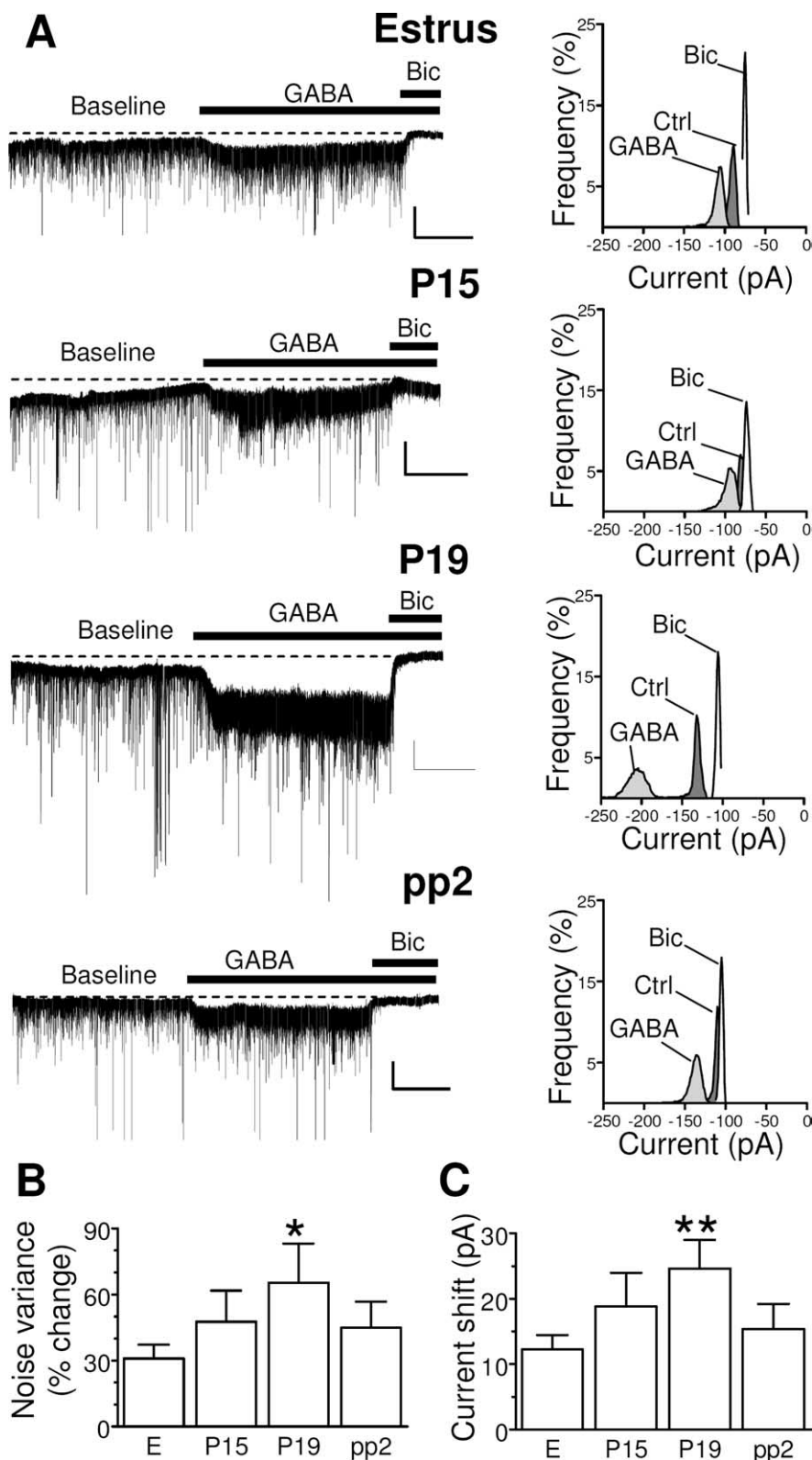
granule cells. Quantitative analysis of basal mIPSC kinetic characteristics did not indicate any statistically significant difference among animals of the various experimental groups, although amplitude, decay time constant, and area of mIPSCs were slightly decreased and increased in P19 and 2 d after delivery rats, respectively, compared with animals in estrus (supplemental Table S1, available at [www.jneurosci.org](http://www.jneurosci.org) as supplemental material).

### Sensitivity of GABA<sub>A</sub>-Rs to 3 $\alpha$ ,5 $\alpha$ -THP during pregnancy

Neurosteroids are among the most potent endogenous positive modulators of GABA<sub>A</sub>-Rs, but their activity is dependent on the receptor subunit composition (Belelli and Lambert, 2005). They thus manifest a higher efficacy at GABA<sub>A</sub>-Rs that contain the  $\delta$  subunit than at those that contain the  $\gamma_2$  subunit (Adkins et al., 2001; Brown et al., 2002; Wohlfarth et al., 2002). Extrasynaptic GABA<sub>A</sub>-Rs are therefore more sensitive (nanomolar affinity) to this class of modulators than are synaptic receptors (micromolar affinity) (Stell et al., 2003). We next tested the effects of the neurosteroid 3 $\alpha$ ,5 $\alpha$ -THP on both tonic and phasic GABAergic transmission in rats at P19 in comparison with those in estrus.

Hippocampal slices were exposed to 3 $\alpha$ ,5 $\alpha$ -THP (1  $\mu$ M) for 7 min after a stable baseline had been established for 4 min. As expected, 3 $\alpha$ ,5 $\alpha$ -THP induced a marked increase in the tonic current (Fig. 5A). However, because this effect of 3 $\alpha$ ,5 $\alpha$ -THP was slower in onset and increased more gradually with time compared with that of GABA (Fig. 5A), we selected only the last 2 min period of 3 $\alpha$ ,5 $\alpha$ -THP application before the addition of bicuculline (corresponding to the maximal increase in tonic current) for our analysis. 3 $\alpha$ ,5 $\alpha$ -THP induced an increase in noise variance and holding current in slices from rats in estrus or at P19 (Fig. 5B,C). The differences in both noise variance and holding current between estrus and P19 were statistically significant. At a lower concentration (0.5  $\mu$ M), 3 $\alpha$ ,5 $\alpha$ -THP induced a small but nonsignificant increase in the tonic current in both groups of animals.

We also examined the effect of 3 $\alpha$ ,5 $\alpha$ -THP on sIPSCs in granule cells. As expected, 3 $\alpha$ ,5 $\alpha$ -THP (1  $\mu$ M) induced marked increases in the decay time constant, amplitude, and area of sIPSCs both in estrus and at P19, but the extent of these increases did not differ between rats of the two groups (supplemental Fig. S4, available at [www.jneurosci.org](http://www.jneurosci.org) as supplemental material). In contrast, 3 $\alpha$ ,5 $\alpha$ -THP sub-



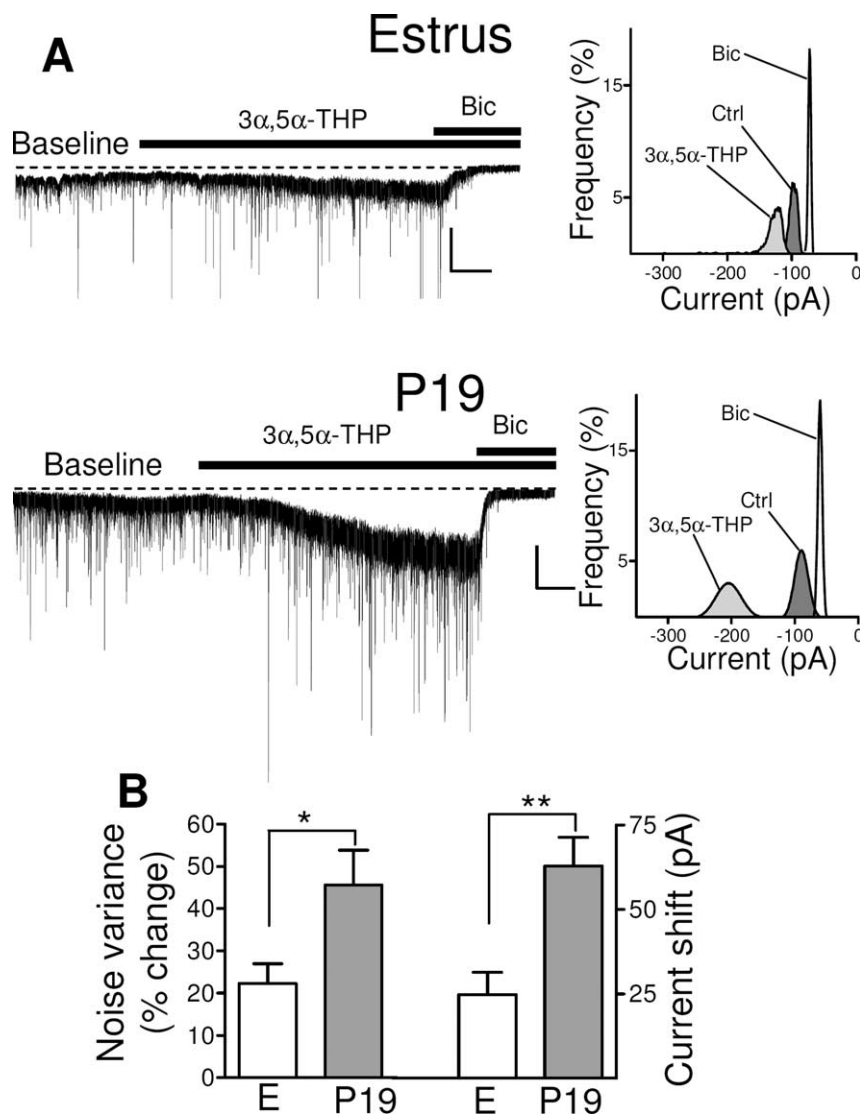
**Figure 4.** Changes in tonic GABAergic current in granule cells of the dentate gyrus during pregnancy and after delivery. **A**, Representative traces of GABAergic currents recorded in the whole-cell mode (holding potential,  $-65$  mV) from granule cells of the dentate gyrus are shown (left panels) for hippocampal slices isolated from female rats in estrus (E), at day P15 or P19, or at pp2. All recordings were performed in the presence of kynurenic acid (3 mM) to block glutamatergic currents. After a baseline period of 5 min, application of GABA (5  $\mu$ M) induced an increase in noise variance and a shift in the holding current. All GABAergic currents were blocked by the application of bicuculline (Bic) at 30  $\mu$ M. Calibration: 50 pA, 2 min. An all-point histogram for 2 min periods before (Ctrl) or during the application of GABA alone or with bicuculline is also shown for each trace (right panels). **B**, **C**, Bar graphs summarizing the changes in noise variance (**B**) and holding current (**C**) induced by the application of exogenous GABA in experiments similar to those shown in **A**. Data are means  $\pm$  SEM from 32 to 67 neurons. \* $p < 0.05$ , \*\* $p < 0.01$  versus estrus (ANOVA followed by Scheffé's test).



**Table 1.** Kinetic characteristics of GABA<sub>A</sub>-R-mediated sIPSCs recorded from granule cells of the dentate gyrus during pregnancy and after delivery

Group (n)	Amplitude (pA)	Decay time (ms)	Area (pA × ms)	Frequency (Hz)
E (68)	50.8 ± 3.5	16.5 ± 0.6	1617 ± 104	1.5 ± 0.1
P15 (38)	47.4 ± 2.2	13.7 ± 0.3	1555 ± 113	2.0 ± 0.2
P19 (55)	46.6 ± 2.1	15.3 ± 0.5	1457 ± 121	1.2 ± 0.1
pp2 (38)	55.1 ± 3.7	15.5 ± 0.6	1726 ± 243	1.6 ± 0.1

Data are means ± SEM for the indicated numbers (n) of neurons. The sIPSCs were recorded in the presence of 3 mM kynurenic acid (external) from voltage-clamped (−65 mV) cells in hippocampal slices isolated from female rats in estrus (E), at P15 or P19, or at pp2.



**Figure 5.** Modulatory action of 3 $\alpha$ ,5 $\alpha$ -THP on tonic GABAergic current in granule cells of the dentate gyrus during pregnancy. **A**, Representative traces of GABAergic currents recorded from granule cells in hippocampal slices from rats in estrus or at P19 (left panels). Recordings were obtained before (Baseline) and during bath application of 1  $\mu$ M 3 $\alpha$ ,5 $\alpha$ -THP either alone or in the presence of 30  $\mu$ M bicuculline. Calibration: 100 pA, 1 min. An all-point histogram for 2 min periods at the baseline (Ctrl) or during the application of 3 $\alpha$ ,5 $\alpha$ -THP alone (final 2 min before the addition of bicuculline) or with bicuculline is shown for each trace (right panels). **B**, Bar graph summarizing the percentage changes in noise variance (left panel) and holding current (right panel) induced by 3 $\alpha$ ,5 $\alpha$ -THP in experiments similar to those shown in **A**. Data are means ± SEM from 13 to 23 neurons. \* $p$  < 0.05, \*\* $p$  < 0.01 versus estrus (ANOVA followed by Scheffé's test).

stantially reduced the frequency of sIPSCs in both groups (supplemental Fig. S4, available at [www.jneurosci.org](http://www.jneurosci.org) as supplemental material), an effect that was likely attributable to the masking of phasic events of smaller amplitude by the marked enhancement of tonic conductance.

### Effect of lorazepam on GABA<sub>A</sub>-R function during pregnancy

To examine further the functional consequences of the pregnancy-induced changes in GABA<sub>A</sub>-R subunit expression, we examined the effect of the benzodiazepine lorazepam on GABAergic currents in granule cells of the dentate gyrus at P19. As expected (Nusser and Mody, 2002), bath application of lorazepam (3  $\mu$ M) for 5 min had no effect on tonic current in rats either in estrus or at P19 (data not shown). In contrast, lorazepam markedly increased the amplitude, area and decay time constant of sIPSCs by similar extents for rats in estrus or at P19 (supplemental Table S2, available at [www.jneurosci.org](http://www.jneurosci.org) as supplemental material). The effect of lorazepam on sIPSC kinetic parameters was however slightly reduced at P19 compared with rats in estrus but this effect did not reach statistical significance.

### Effect of Ro15-4513 on GABA<sub>A</sub>-R-mediated sIPSCs during pregnancy and after delivery

To evaluate the functional relevance of the increased expression of the  $\alpha_4$  and  $\gamma_2$  subunits of the GABA<sub>A</sub>-R in the hippocampus 2 d after delivery, we tested the modulatory effect of Ro15-4513 on GABA<sub>A</sub>-R-mediated sIPSCs in granule cells of the dentate gyrus. Ro15-4513 is an inverse agonist at the benzodiazepine-binding site of GABA<sub>A</sub>-Rs containing either the  $\alpha_1$ ,  $\alpha_2$ ,  $\alpha_3$ , or  $\alpha_5$  subunit together with the  $\gamma_2$  subunit (Barnard et al., 1998), but it is able to bind and positively modulate receptors comprising  $\alpha_4$ ,  $\beta$ , and  $\gamma_2$  subunits (Knoflach et al., 1996; Wafford et al., 1996). In granule cells of the dentate gyrus of rats in estrus and at day 19 of pregnancy, bath application of 3  $\mu$ M Ro15-4513 reduced (24 and 19%, respectively) the time constant of sIPSC decay. In contrast, in granule cells of the dentate gyrus from pp2 rats, Ro15-4513 increased by 28% the time constant of sIPSC decay (Table 2). The frequency and amplitude of sIPSCs were not significantly altered by Ro15-4513.

### GABAergic inhibitory transmission in CA1 pyramidal neurons during pregnancy

Although tonic currents in the hippocampus have been characterized most extensively in granule cells of the dentate gyrus, this type of inhibitory current has also

been detected in CA1 pyramidal neurons (Caraiscos et al., 2004). After a baseline period of 5 min, we exposed hippocampal slices to GABA (5  $\mu$ M) and then blocked all currents with bicuculline (30  $\mu$ M). However, we failed to detect a substantial tonic current in CA1 pyramidal neurons of rats in estrus or at P19 with this

**Table 2. Effect of Ro15–4513 on GABA<sub>A</sub>-R-mediated sIPSCs in granule cells of the dentate gyrus during pregnancy and after delivery**

Group (n)	Amplitude (% change)	Decay time (% change)	Area (% change)	Frequency (% change)
E (5)	−5.5 ± 7.2	−24.1 ± 4.1*	−31.2 ± 9.5*	−11.2 ± 7.3
P19 (8)	−10.3 ± 6.8	−19.5 ± 5.2*	−23.1 ± 10.3*	−3.6 ± 6.9
pp2 (8)	12.2 ± 10.2	28.6 ± 8.3*	25 ± 11.5	−5.8 ± 7.7

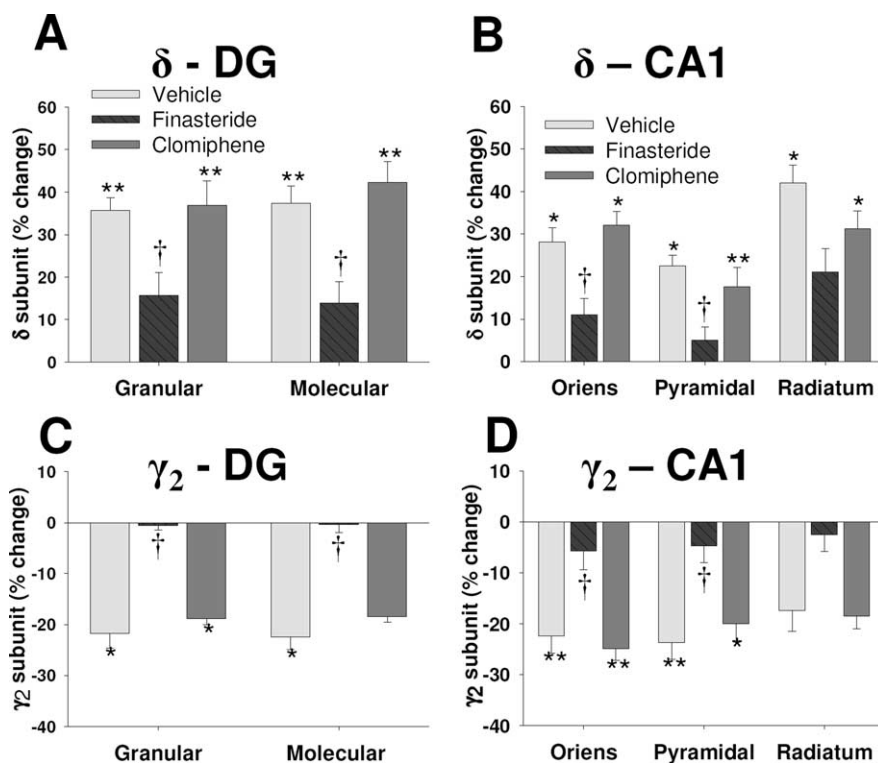
Data are expressed as the percentage of change in current parameters induced by Ro15-4513 (3  $\mu$ M) and are means  $\pm$  SEM for the indicated numbers (n) of neurons in hippocampal slices obtained from rats in estrus, at day 19 of pregnancy, or at 2 d after delivery. \* $p$  < 0.05 versus estrus (one-way ANOVA followed by Scheffé's test).

protocol (supplemental Fig. S5, available at [www.jneurosci.org](http://www.jneurosci.org) as supplemental material). We also characterized the kinetic parameters of sIPSCs (supplemental Table S3, available at [www.jneurosci.org](http://www.jneurosci.org) as supplemental material) as well as the effect of lorazepam on these currents (supplemental Table S4, available at [www.jneurosci.org](http://www.jneurosci.org) as supplemental material) in CA1 pyramidal neurons of rats of the various experimental groups. No significant effects of pregnancy or delivery were apparent.

### Role of neurosteroids in the changes in GABA<sub>A</sub>-R expression and function during pregnancy

Given that the selective and opposite changes in the expression of  $\delta$  and  $\gamma_2$  subunits of the GABA<sub>A</sub>-R as well as the enhancement of GABAergic tonic current in granule cells of the dentate gyrus observed in rats at P19 appeared temporally correlated with the previously demonstrated increases in the brain concentrations of progesterone and 3 $\alpha$ ,5 $\alpha$ -THP (Concas et al., 1998) as well as of estrogen (Taya and Greenwald, 1981), we examined whether these events might be causally related. Finasteride, an inhibitor of the enzyme 5 $\alpha$ -reductase, or clomiphene, an estrogen receptor antagonist, was administered to pregnant rats from P12 to P18 to inhibit the synthesis of 3 $\alpha$ ,5 $\alpha$ -THP or the activation of estrogen receptors, respectively (Azzolina et al., 1997; Homburg, 2005; Finn et al., 2006). Finasteride treatment inhibited the upregulation of  $\delta$  subunit expression (Fig. 6*A,B*) and the downregulation of  $\gamma_2$  subunit expression (Fig. 6*C,D*) observed in the dentate gyrus and the CA1 region at P19. In contrast, clomiphene administration had no significant effect on the expression level of the  $\delta$  subunit (Fig. 6*A,B*) or the  $\gamma_2$  subunit (Fig. 6*C,D*) apparent at P19.

We also examined GABA<sub>A</sub>-R function in granule cells of the dentate gyrus in hippocampal slices prepared from rats in estrus or from those at P19 that had been injected with finasteride or vehicle. The differences in the effects of GABA on tonic current parameters (noise variance and holding current) between rats at P19 and those in estrus were reduced and rendered statistically insignificant by finasteride treatment (Fig. 7*A,B*). In contrast, finasteride treatment had no effect on the kinetic properties of GABAergic sIPSCs in granule cells of rats at P19 (data not shown). Finally, clomiphene treatment had no effect on tonic current parameters in granule cells of rats at P19 (Fig. 7*C,D*).



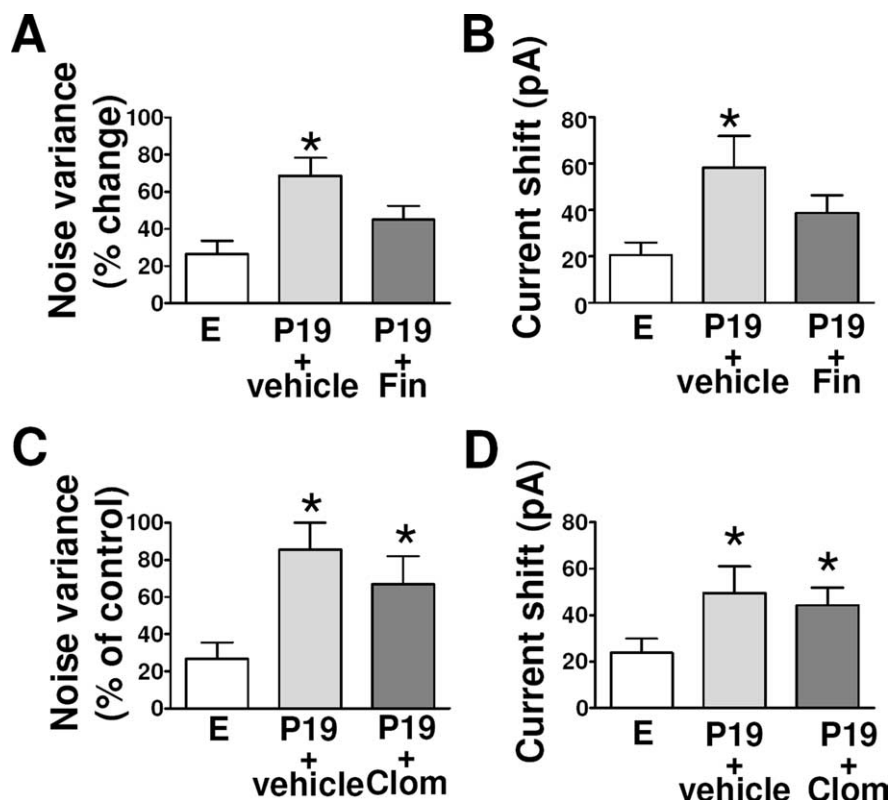
**Figure 6.** Effect of treatment with finasteride or clomiphene on expression of the  $\delta$  and  $\gamma_2$  subunits of the GABA<sub>A</sub>-R during pregnancy. Finasteride (25 mg/kg), clomiphene (5 mg/kg), or vehicle was administered daily from day 12 to day 18 of pregnancy. **A–D**, Rats were killed at day 19 of pregnancy and the hippocampus was subjected to immunohistochemistry for semiquantitative measurement of  $\delta$  subunit (**A, B**) or  $\gamma_2$  subunit (**C, D**) immunoreactivity in the dentate gyrus and the CA1 region, respectively. Data are expressed as percentage of change relative to the corresponding value for rats in estrus and are means  $\pm$  SEM from at least eight animals per group. Results from rats treated with the vehicle for finasteride or with that for clomiphene were pooled. \* $p$  < 0.05, \*\* $p$  < 0.01 versus rats in estrus; † $p$  < 0.05 versus vehicle-treated rats at day 19 of pregnancy (one-way ANOVA followed by Scheffé's test).

Clomiphene treatment also had no significant effect on the kinetic parameters of GABAergic sIPSCs recorded from granule cells at P19 in the absence or presence of 3 $\alpha$ ,5 $\alpha$ -THP (data not shown).

### Discussion

Our present results provide further support to the notion that both the structure and function of GABA<sub>A</sub>-Rs in the brain change during pregnancy and immediately after delivery as a result of the marked associated fluctuations in the brain levels of neuroactive steroids. We have thus shown that the late stage of pregnancy in rats is associated with an increase in the expression of the  $\delta$  subunit of the GABA<sub>A</sub>-R and a decrease in that of the  $\gamma_2$  subunit in various regions of the hippocampus. In contrast, the expression of the  $\alpha_4$  subunit did not change during pregnancy but rather increased markedly after delivery. This selective switch in subunit composition of GABA<sub>A</sub>-Rs during pregnancy and after delivery was associated with a selective enhancement and reduction of tonic current mediated by extrasynaptic GABA<sub>A</sub>-Rs in granule





**Figure 7.** Effect of finasteride or clomiphene treatment on the changes in GABAergic tonic current in granule cells of the dentate gyrus during pregnancy. **A–D**, The percentage of changes in noise variance (**A**, **C**) and holding current (**B**, **D**) induced by GABA were determined in hippocampal slices from rats in estrus as well as from those at day 19 of pregnancy that had been treated with finasteride (**A**, **B**) or clomiphene (**C**, **D**) as in Figure 6. \* $p < 0.05$  versus estrus (ANOVA followed by Scheffé's test).

cells of the dentate gyrus, respectively, with phasic sIPSCs mediated by synaptic receptors being unaffected. These effects of pregnancy on the subunit composition and function of GABA<sub>A</sub>-Rs were inhibited by treatment with finasteride, suggesting that they are mediated by the associated changes in the brain levels of 3 $\alpha$ ,5 $\alpha$ -THP and other neuroactive steroids during late pregnancy (Concas et al., 1998). Such an increase in GABAergic tonic at day 19 of pregnancy may be important to further reduce the excitability and anxiety level usually associated with the end of pregnancy and immediately preceding delivery (Zuluaga et al., 2005; de Brito Faturi et al., 2006; Skouteris et al., 2009).

The brain levels of steroid hormones increase markedly during pregnancy in rats (Concas et al., 1998). Indeed the concentrations of progesterone, 3 $\alpha$ ,5 $\alpha$ -THP, and 3 $\alpha$ ,5 $\alpha$ -tetrahydrodeoxycorticosterone increase severalfold in both plasma and the brain (relative to estrus values) starting from day 15 and peaking at day 19 of pregnancy. However, the brain levels of these hormones fall suddenly to control levels at day 21 of pregnancy, immediately preceding delivery, and they remain at such levels for up to 2 d (Concas et al., 1998).

We have previously shown that the sustained increases in the brain levels of neuroactive steroids during pregnancy are responsible for the associated reduction in the expression of the  $\alpha_5$  and  $\gamma_2$  subunits of the GABA<sub>A</sub>-R in both the cerebral cortex and hippocampus of rats (Concas et al., 1998; Follés et al., 1998). Late pregnancy was also associated with an increase in the abundance of the  $\alpha_1$  subunit mRNA in oxytocin neurons of the supraoptic nucleus of the hypothalamus (Fénelon and Herbison, 1996). Our present immunohistochemical data confirm our previous observation of the downregulation of  $\gamma_2$  subunit expression

in the hippocampus of rats during pregnancy. The amount of  $\gamma_2$  subunit immunoreactivity in the hippocampus was decreased from P15 to P19, an effect that was well correlated temporally with the increase in the brain levels of neuroactive steroids and which was both largely abolished by finasteride treatment (Concas et al., 1998). Furthermore, this reduction was promptly reversed in response to the sudden fall in the brain concentrations of these steroid hormones at day 21 of pregnancy and during the first 2 d after delivery.

The abundance of immunoreactivity for the  $\delta$  subunit showed changes opposite to those for  $\gamma_2$  subunit immunoreactivity during pregnancy and after delivery. In the hippocampus, the  $\delta$  subunit is expressed predominantly in granule cells of the dentate gyrus as well as in CA1 and CA3 pyramidal neurons, where it coassembles mostly with the  $\alpha_4$  subunit (Pirker et al., 2000; Nusser and Mody, 2002; Wei et al., 2003), although it also coassembles with the  $\alpha_1$  subunit in interneurons in the molecular layer of the dentate gyrus (Glykys et al., 2007). Specific immunostaining for the  $\delta$  subunit in the hippocampus increased progressively during pregnancy, reaching a peak at days 19–21, and then fell markedly, being below control levels at 2 and 7 d after delivery. Consistent with the higher

sensitivity of GABA<sub>A</sub>-Rs containing the  $\delta$  subunit to neuroactive steroids (Adkins et al., 2001; Brown et al., 2002; Wohlfarth et al., 2002), fluctuations in the plasma and brain content of these compounds associated with various pathophysiological or pharmacological conditions are accompanied by changes in the expression of the  $\delta$  subunit together, in some instances, with changes in  $\alpha_4$  subunit expression (Smith et al., 1998a,b; Sundstrom-Poromaa et al., 2002; Griffiths and Lovick, 2005; Lovick et al., 2005; Maguire et al., 2005; Shen et al., 2005).

We found that expression of the  $\alpha_4$  subunit in the hippocampus did not change during pregnancy but rather increased markedly after delivery, likely as a consequence of the sudden decrease in the anxiolytic steroid concentrations. This finding under the physiological conditions of pregnancy and delivery is consistent with those of pharmacological studies of prolonged treatment with and subsequent withdrawal of neuroactive steroids (Smith et al., 1998a,b; Sundstrom-Poromaa et al., 2002; Biggio et al., 2006; Mostallino et al., 2006). Immediately after delivery, the marked reduction in the expression of the  $\delta$  subunit is accompanied with an increase in the number of receptors containing the  $\alpha_4$  and  $\gamma_2$  subunit, which, being endowed with a lower sensitivity to GABA, increased desensitization rate, and likely synaptic localization, may contribute to the reversal of the increase in tonic current level and to the development of an arousal state observed in the perinatal period. The idea of an enhanced density of  $\alpha_4\beta_{1-3}\gamma_2$  receptors is further supported by the finding that Ro15-4513 increases (Knoflach et al., 1996; Wafford et al., 1996) the time constant for sIPSC decay in granule cells of the dentate gyrus from rats at 2 d after delivery whereas it reduced this parameter in animals in estrus or at day 19 of pregnancy. This pattern of changes in the expression of these subunits is similar to that

described for the ovarian cycle, during which the abundance of the  $\delta$  and  $\gamma_2$  subunits is regulated in opposite manners (Maguire et al., 2005).

Our results have revealed substantial and selective changes in GABA-mediated transmission in granule cells of the dentate gyrus during pregnancy and after delivery. In particular, we detected a marked increase in tonic current during the late stage of pregnancy relative to that in control rats in estrus, and this increase was no longer apparent 2 d after delivery. Application of exogenous GABA thus induced a larger percentage increase in noise variance and shift in the holding current in rats at P19 than in control animals. Given that tonic transmission in granule cells is mediated by receptors containing the  $\delta$  subunit, our functional data are consistent with an increase in the density of these receptors in the extrasynaptic membrane.

Neuroactive steroids are among the most potent endogenous modulators of GABA<sub>A</sub>-Rs (Majewska et al., 1986; Belelli and Lambert, 2005). These compounds modulate more potently and effectively the tonic transmission mediated by  $\delta$  subunit-containing receptors than the phasic transmission mediated by  $\gamma_2$  subunit-containing receptors (Stell et al., 2003). The finding that  $3\alpha,5\alpha$ -THP potentiated to a greater extent the tonic GABAergic transmission in granule cells of the dentate gyrus at P19 than at 2 d after delivery or in estrus supports the idea that upregulation of the expression of GABA<sub>A</sub>-Rs containing the  $\delta$  subunit plays an important role during late pregnancy. Similarly, changes in the plasma levels of neuroactive steroids, an increase in tonic transmission in granule cells of the dentate gyrus, and upregulation of expression of the  $\delta$  subunit have been shown to be tightly associated during the estrous cycle in mice (Maguire et al., 2005; Maguire and Mody, 2007).

Such changes in tonic current appear to be limited to granule cells of the dentate gyrus, given that neither tonic nor phasic currents recorded from CA1 pyramidal neurons were affected by pregnancy or delivery. The tonic current in CA1 pyramidal neurons has previously been recorded by only a few laboratories and under particular experimental conditions and pharmacological manipulations (Caraiscos et al., 2004).

The kinetic properties of synaptic currents in granule cells of the dentate gyrus were not substantially altered during pregnancy despite a decrease in  $\gamma_2$  subunit expression. Downregulation of the expression of the  $\gamma_2$  subunit during the diestrus phase of the ovarian cycle in mice was also found not to be accompanied by changes in the kinetic parameters of synaptic currents (Maguire et al., 2005). Our results now suggest that the density of GABA<sub>A</sub>-Rs containing the  $\delta$  subunit and the associated tonic current are selectively modified during the late stage of pregnancy and after delivery.

Our results show that the opposite changes in GABAergic tonic inhibition in the dentate gyrus during pregnancy and after delivery correlate with characteristic hormonal changes (Concas et al., 1998). Very recently, a similar study (Maguire and Mody, 2008) evaluated in mice the changes during pregnancy and post partum of GABA<sub>A</sub>  $\delta$  and  $\gamma_2$  subunit expression in the hippocampus and found, at variance with our work, a decreased expression of both subunits at P18 compared with virgin animals. Also in that work, both tonic and phasic inhibitory currents in DGGCs were markedly reduced at P18 compared with control. All of these changes rebounded 48 h after delivery. Apart from the species difference and other differences related to the experimental conditions (i.e., anti- $\delta$  antibodies used, higher temperature used in the patch-clamp recordings, and the continuous application of exogenous GABA to the slices to sustain tonic current in Maguire's study), a more likely and most influential factor that could account for the difference among the two studies may be the animals that were used as controls. In fact, while in our

study we used rats in the estrus phase of the ovary cycle, Maguire and Mody used female mice in the diestrus phase. This may be relevant because at diestrus  $\delta$  expression (and tonic currents) is at its highest level (in association with the peak of neuroactive steroid levels) during the ovary cycle, whereas in the estrus phase both  $\delta$  expression (and tonic current) reaches its lowest level (Maguire et al., 2005). Thus, it is conceivable that the use of such opposite reference points may have contributed to reach a different interpretation of the changes occurring during pregnancy. However, this may not completely account for the different pattern of changes observed during pregnancy and after delivery in these two studies, and at the moment we do not find more convincing reasons that may justify these discrepancies.

In conclusion, the opposite regulation of extrasynaptic GABA<sub>A</sub>-R-mediated tonic current in response to fluctuations of brain concentrations of neuroactive steroids may be crucial for the changes in mood and for the anxiolytic effect associated with pregnancy (de Brito Faturi et al., 2006) as well as for the arousal state typical of period immediately preceding delivery and early postpartum.

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