

# Paracrine Role for Somatostatin Interneurons in the Assembly of Perisomatic Inhibitory Synapses

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GABAergic interneurons represent a heterogeneous group of cell types in neocortex that can be clustered based on developmental origin, morphology, physiology, and connectivity. Two abundant populations of cortical GABAergic interneurons include the low-threshold, somatostatin (SST)-expressing cells and the fast-spiking, parvalbumin (PV)-expressing cells. While SST<sup>+</sup> and PV<sup>+</sup> interneurons are both early born and migrate into the developing neocortex at similar times, SST<sup>+</sup> cells are incorporated into functional circuits prior to PV<sup>+</sup> cells. During this early period of neural development, SST<sup>+</sup> cells play critical roles in the assembly and maturation of other cortical circuits; however, the mechanisms underlying this process remain poorly understood. Here, using both sexes of conditional mutant mice, we discovered that SST<sup>+</sup> interneuron-derived Collagen XIX, a synaptogenic extracellular matrix protein, is required for the formation of GABAergic, perisomatic synapses by PV<sup>+</sup> cells. These results, therefore, identify a paracrine mechanism by which early-born SST<sup>+</sup> cells orchestrate inhibitory circuit formation in the developing neocortex.

**Key words:** collagen; extracellular matrix; inhibitory synapse; neocortex; somatostatin; synaptogenesis

## Significance Statement

Inhibitory interneurons in the cerebral cortex represent a heterogeneous group of cells that generate the inhibitory neurotransmitter GABA. One such interneuron type is the low-threshold, somatostatin (SST)-expressing cell, which is one of the first types of interneurons to migrate into the cerebral cortex and become incorporated into functional circuits. In addition, to contributing important roles in controlling the flow of information in the adult cerebral cortex, SST<sup>+</sup> cells play important roles in the development of other neural circuits in the developing brain. Here, we identified an extracellular matrix protein that is released by these early-born SST<sup>+</sup> neurons to orchestrate inhibitory circuit formation in the developing cerebral cortex.

## Introduction

GABAergic interneurons account for only ~20% of all cortical neurons, but play essential roles in controlling the spatial and temporal spread of neural activity (Huang et al., 2007; Tremblay et al., 2016). In neocortex, GABAergic interneurons represent a

heterogeneous group and can be clustered into distinct subtypes based on morphology, physiology, molecular diversification, developmental origin, spatial distribution, and connectivity (Ascoli et al., 2008; Tremblay et al., 2016; Hu et al., 2017; Wamsley and Fishell, 2017; Lim et al., 2018). Genetic, epigenetic, and activity-dependent mechanisms underlie the diversification of these interneurons, as well as their incorporation into cortical circuits (De Marco García et al., 2011; Kepecs and Fishell, 2014; Sandberg et al., 2016; Hu et al., 2017; Wamsley and Fishell, 2017; Mayer et al., 2018).

One of the most abundant cortical interneuron subtypes is the fast-spiking cell, which expresses the calcium-binding albumin protein parvalbumin, or Pvalb (abbreviated here as PV). Cortical PV<sup>+</sup> interneurons are innervated by glutamatergic thalamocortical inputs and generate feedforward, perisomatic inhibitory inputs onto both excitatory pyramidal neurons and other PV<sup>+</sup> interneurons (Tremblay et al., 2016). These PV<sup>+</sup> perisomatic synapses play important roles in controlling the flow of

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excitatory activity in neocortex. In fact, disruption of these PV<sup>+</sup> circuits leads to seizures and has been associated with a number of neurodevelopmental and psychiatric disorders, including schizophrenia (Benes and Berretta, 2001; Schwaller et al., 2004; Belforte et al., 2010; Gonzalez-Burgos et al., 2010, 2011; Sgadò et al., 2011; Gonzalez-Burgos and Lewis, 2012; Lewis et al., 2012; Wöhr et al., 2015; Ferguson and Gao, 2018).

During embryogenesis, PV<sup>+</sup> interneurons are generated in the medial ganglionic eminence before undergoing long-range, tangential migration into the developing neocortex (Wonders and Anderson, 2006). Despite being early born, PV<sup>+</sup> interneuron incorporation into developing cortical circuits is significantly delayed compared with other interneuron subtypes, even those derived from the same germinal zone (Agmon et al., 1996; Huang et al., 1999; Chattopadhyaya et al., 2004; Miyoshi et al., 2010), such as low-threshold, somatostatin (SST)-expressing interneurons (Wonders and Anderson, 2006; Liguz-Leczna et al., 2016). Based on their important role in neocortical circuit function and neurodevelopmental and psychiatric disorders, understanding the mechanisms that regulate the incorporation of PV<sup>+</sup> interneurons into circuits has received considerable attention. Previously, we identified a role for Collagen XIX, a neuron-derived extracellular matrix (ECM) protein associated with familial schizophrenia, in PV<sup>+</sup> perisomatic synapse assembly (Su et al., 2010, 2016). Specifically, global deletion of *Col19a1*, the gene encoding Collagen XIX, results in a loss of PV<sup>+</sup> perisomatic inhibitory synapses, the development of spontaneous seizures, and the acquisition of schizophrenia-related behaviors (Su et al., 2016). *In vitro* analyses further revealed that an endogenously shed C-terminal fragment of Collagen XIX (termed a matricryptin; Ricard-Blum and Salza, 2014) is sufficient to trigger inhibitory synaptogenesis (Su et al., 2016). However, at present, the cellular mechanisms underlying Collagen XIX-triggered perisomatic synapse formation have remained unclear. To address this issue, we generated a novel conditional allele for cell-specific deletion of this nonfibrillar collagen.

Here, we demonstrate that neuron-derived Collagen XIX is indeed necessary for the assembly of perisomatic inhibitory synapses in mouse neocortex. While our previous studies revealed little, if any, expression of Collagen XIX by excitatory neurons or PV<sup>+</sup> interneurons (the two main cell types on whose somas PV<sup>+</sup> perisomatic synapses form), transcriptomic and single-cell analyses suggest that *Col19a1* (the gene encoding Collagen XIX) is enriched in SST<sup>+</sup> interneurons compared with all other cortical cell types (Hrvatín et al., 2018; Lim et al., 2018; Sugino et al., 2006; Krienen et al., 2019). This is particularly interesting since early-born SST<sup>+</sup> interneurons shape neocortical connectivity and activity during early postnatal development, including shaping the development and maturation of feedforward PV<sup>+</sup> circuits (Liguz-Leczna et al., 2016; Tuncdemir et al., 2016; Wang et al., 2019). Based on these data, we sought to test whether SST<sup>+</sup> interneurons generate Collagen XIX and whether SST<sup>+</sup> interneuron-derived Collagen XIX is necessary for the formation of PV<sup>+</sup> perisomatic inhibitory synapses. Our results reveal that multiple SST<sup>+</sup> interneuron subtypes generate *Col19a1* and that conditional loss of SST<sup>+</sup> interneuron-derived Collagen XIX impairs perisomatic inhibitory synapse formation. Together, these data identify a novel paracrine mechanism through which SST<sup>+</sup> interneurons drive the assembly of feedforward inhibitory circuits in the developing cerebral cortex.

## Materials and Methods

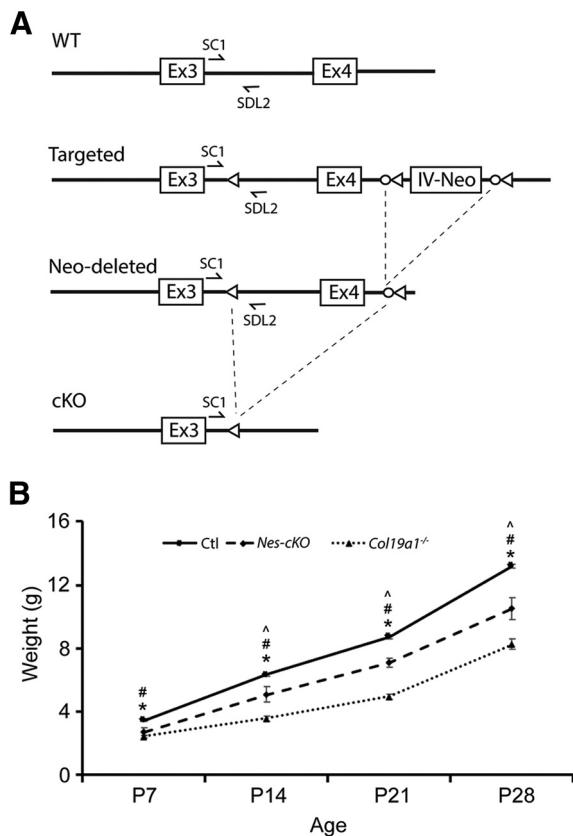
**Mouse lines and husbandry.** C57BL/6 mice were obtained from Charles River Laboratories. The generation of Collagen XIX-null mice

[*Col19a1*<sup>-/-</sup> (previously referred to as N19)] was previously described (Sumiyoshi et al., 2004; Su et al., 2010). The following primers were used for genotyping *Col19a1*<sup>-/-</sup> mice: *Col19a1*, forward (exon 4): 5'-CTT CGC AAA ACG CAT GCC TCA GA-3'; *Col19a1*, reverse: 5'-TTG TTC GTT TGT TTG TTT TTA ATC AAT CAA-3'; *Lacz*, forward: 5'-TTC ACT GCG CGT CGT TTT ACA ACG TCG TGA-3'; and *Lacz*, reverse: 5'-ATG TGA GCG AGT AAC AAC CCG TCG GAT TCT-3' (Sumiyoshi et al., 2004; Su et al., 2010). *PV-Cre*, *Nes-Cre*, *Gad2-Cre*, and *Sst-Cre* mice were obtained from The Jackson Laboratory (stock #008069, #003771, #010802, #013044). *Crh-Cre* mice (stock #030850-UCD) were obtained from the MMRC (Mutant Mouse Resource & Research Centers). The following primers were used for genotyping Cre-expressing mice: *Cre*, forward: 5'-CGT ACT GAC GGT GGG AGA AT-3'; and *Cre*, reverse: 5'-TGC ATG ATC TCC GGT ATT GA-3'. The following two reporter lines were obtained from The Jackson Laboratory: *Sun1-sfGFP-Myc* (stock #021039) and *Rosa-Stop-tdT* (Ai9; stock #007909). The following primers were used for genotyping GFP- and terminal deoxynucleotidyl transferase (tdT)-expressing mice: *GFP*, forward: 5'-AAG TTC ATC TGC ACC ACC G-3'; *GFP*, reverse: 5'-TCC TTG AAG AAG ATG GTG CG-3'; *tdT* forward: 5'-ACC TGG TGG AGT TCA AGA CCA TCT-3'; and *tdT*, reverse: 5'-TTG ATG ACG GCC ATG TTG TTG TCC-3'. *Rosa-Stop-DTA* mice were obtained from The Jackson Laboratory (stock #006331). The following primers were used for genotyping *Rosa-Stop-DTA* mice: *DTA-Common*: 5'-AAA GTC GCT CTG AGT TGT TAT-3'; *DTA-Wt*, reverse: 5'-GGA GCG GGA GAA ATG GAT ATG-3'; and *DTA-Mut*, reverse: 5'-GCG AAG AGT TTG TCC TCA ACC-3'. Mice were housed in a 12 h dark/light cycle and had *ad libitum* access to food and water. All experiments were performed in compliance with National Institutes of Health guidelines and protocols, and were approved by the Virginia Polytechnic Institute and State University Institutional Animal Care and Use Committee.

**Generation of *Col19a1*<sup>fl/fl</sup> mice.** A conditional allele of *Col19a1* (*Col19a1*<sup>fl/fl</sup>) mice was generated by Ingenious Targeting Laboratory. Briefly, a 597 bp region of the genome containing exon 4 of *Col19a1* [Fig. 1A (the same region deleted in *Col19a1*<sup>-/-</sup> mice); Sumiyoshi et al., 2004] was targeted. The 5' LoxP sites were cloned into intron 3, and a Frt-LoxP-Neo-Frt-LoxP cassette was cloned into intron 4. The targeting vector was screened, sequenced, and electroporated into iTL HF4 (129/SvEv × C57BL/6 FLP) hybrid embryonic stem cells, which were subsequently microinjected into C57BL/6 blastocysts. Resulting chimeras were mated with C57BL/6 WT mice to generate Neo Deleted mice (Fig. 1A). The following primers were used for genotyping Neo Deleted and *Col19a1*<sup>fl/fl</sup> mice: SC1: 5'-AGT GGG GAT TCA GGG AGG GTA AAG-3'; and SDL2: 5'-AAT GCG GGC AGA GTA GCA CTA AGG-3'.

**Reagents.** Fluorescein RNA Labeling Mix (catalog #11685619910), digoxigenin (DIG) RNA Labeling Mix (#11277073910), blocking reagent [*in situ* hybridization (ISH); catalog #11096176001], and Yeast RNA (catalog #10109223001) were from Roche. Proteinase K (catalog #E00491), heparin sodium (catalog #BP2425), and Ambion MAXIScript T7 *In Vitro* Transcription Kit (catalog #AM1312) were from Thermo Fisher Scientific. Paraformaldehyde (PFA), EM Grade (catalog #19202) and Tissue Freezing Medium (catalog #72592) were from EMS. Prehybridization Solution (catalog #P1415) was from Sigma. VECTASHIELD mounting medium (catalog #H-1000) was from Vector Laboratories. The Invitrogen Superscript II Reverse Transcriptase First Strand cDNA Synthesis kit (catalog #18064014) was from Thermo Fisher Scientific. The Aurum Total RNA Fatty and Fibrous Tissue Kit (catalog #7326870) was from Bio-Rad. pGEM-T Easy Vector Systems (catalog #A1360) was from Promega. Tyramide Signal Amplification (TSA) Systems (catalog #NEL75300 1KT) were from PerkinElmer. The Invitrogen NeuroTrace500/525 (catalog #N-21 480) was from Thermo Fisher Scientific. All other chemicals and reagents were obtained from Thermo Fisher Scientific or Sigma-Aldrich. All DNA primers were obtained from Integrated DNA Technologies.

**Antibodies.** The following antibodies were purchased: mouse anti-Syt2 [Synaptotagmin 2; znp1; Fox and Sanes, 2007; diluted 1:200 for immunohistochemistry (IHC); Zebrafish International Resource Center]; rabbit anti-vesicular glutamate transporter 1 (VGLUT1; diluted 1:500 for IHC; catalog #135303, Synaptic Systems); rabbit anti-RFT for



**Figure 1.** Generation of *Col19a1<sup>fl/fl</sup>* mice. **A**, Schematic of the strategy to generate *Col19a1<sup>fl/fl</sup>* mice. LoxP sites (triangles) were inserted to flank exon (Ex) 4 of *Col19a1*. A neomycin selection cassette (Neo) was inserted after Ex4 and was flanked with flippase (FLP) recombinase target (FRT) sequences (depicted by circles) so that it could be removed in the presence of FLP. cKO alleles were generated by crossing mice carrying Neo-deleted alleles with various Cre-driver lines. SC1 and SDL2 represent primer sequences for genotyping. **B**, Postnatal growth of *Col19a1<sup>fl/fl</sup>* [control (Ctl)], *Col19a1<sup>fl/fl</sup>::Nes-Cre* (*Nes-cKO*), and *Col19a1<sup>-/-</sup>* mice. Data points represent the mean  $\pm$  SEM. #*Nes-cKO* differs from Ctl by  $p=0.002$  at P7,  $p=0.022$  at P14,  $p=0.0007$  at P21, and  $p=0.002$  at P28 by ANOVA;  $n=7$  for both genotypes. \**Col19a1<sup>-/-</sup>* differs from Ctl by  $p=0.00,001$  at P7,  $p=0.00000008$  at P14,  $p=0.00000000002$  at P21, and  $p=0.0000001$  at P28 by ANOVA;  $n=7$  for Ctl and  $n=13$  for *Col19a1<sup>-/-</sup>*. <sup>a</sup>*Nes-cKO* differs from *Col19a1<sup>-/-</sup>* by  $p=0.003$  at P14,  $p=0.0000003$  at P21, and  $p=0.004$  at P28 by ANOVA;  $n=7$  for *Nes-cKO* and  $n=13$  for *Col19a1<sup>-/-</sup>*.

tdT (diluted 1:500 for IHC; catalog #600–401–379, Rockland); Mouse Anti-Parvalbumin (diluted 1:1000 for IHC; catalog #MAB1572, Chemicon); Invitrogen rabbit anti-GFP (diluted 1:250 for IHC; catalog #A-11122, Thermo Fisher Scientific); mouse anti-GAD67 (diluted 1:1000 for IHC; catalog #MAB5406, EMD Millipore); mouse anti-Cre (diluted 1:500 for IHC; catalog #MAB3120, EMD Millipore); sheep (POD)-conjugated anti-DIG (diluted 1:2000; catalog #11426346910, Roche); and sheep (POD)-conjugated anti-FL (diluted 1:2000; catalog #11207733910, Roche).

**Tissue preparation and immunohistochemistry.** Fluorescent IHC was performed on 20  $\mu$ m cryosectioned PFA-fixed brain tissue (Fox et al., 2007; Su et al., 2010, 2012, 2016). Tissue slides were allowed to air dry for 15 min before being incubated with blocking buffer (2.5% normal goat serum, 2.5% BSA, and 0.1% Triton X-100 in PBS) for 30 min. Primary antibodies were diluted in blocking buffer and incubated on tissue sections overnight at 4°C. On the next day, tissue slides were washed in PBS, and secondary antibodies diluted 1:1000 in blocking buffer were applied to slides for 1 h at room temperature. After thorough washes in PBS, tissue slides were coverslipped with VECTASHIELD (Vector Laboratories). Images of tissue were acquired on a confocal microscope (LSM 700, Zeiss) equipped with a 20 $\times$  air Plan-Apochromat objective [numerical aperture (NA) 0.8; Zeiss] and a 40 $\times$  oil EC Plan-Neofluar objective (NA 1.3; Zeiss). When comparing different ages of tissues or

between genotypes, images were acquired with identical parameters, and similar gamma adjustments were made to age-matched mutant and control images in Adobe Photoshop or ImageJ. A minimum of three animals (per genotype and per age) were compared in all IHC experiments. Mean fluorescent intensities of IHC images were measured in ImageJ (Singh et al., 2012; Su et al., 2016). For quantification of perisomatic synapses, single optical sections of confocal images were analyzed, and the density of synaptic elements (Syt2<sup>+</sup> puncta) was quantified per unit length of the cell surface in ImageJ.

**Riboprobe making and in situ hybridization.** ISH was performed on 20  $\mu$ m sagittal cryosectioned tissues (Su et al., 2010, 2016). Antisense riboprobes were generated from full-length *Col19a1* (EMM1002-97504659), *Spon1* (MMM1013-202701079), and *Col25a1* (MMM1013-202707906) Image Clones (Dharmacon), as described previously (Su et al., 2010, 2016; Monavarfeshani et al., 2017; Carrillo et al., 2018). Antisense riboprobes were generated against a 598 bp fragment of *Sst* (corresponding to nucleotides 362–960) that was PCR cloned into pGEM Easy T Vector (Promega) with the following primers: 5'-AGC GGC TGA AGG AGA CGC TAC-3'; and 5'-CGC CAT AAT CTC ACC ATA ATT TTA-3'. In brief, riboprobes were synthesized using DIG-labeled UTP (Roche) and the MAXIscript In Vitro Transcription Kit (Ambion). Probes were hydrolyzed to 500 nt. Tissue sections were fixed in 4% PFA for 10 min, washed with DEPC-PBS three times, and incubated in proteinase K solution (1  $\mu$ g/ml proteinase K, 50 mM Tris, pH 7.5, and 5 mM EDTA) for 10 min. Subsequently, slides were washed with DEPC-PBS, fixed with 4% PFA for 5 min, washed with DEPC-PBS, and incubated in acetylation buffer (1.33% triethanolamine, 20 mM HCl, and 0.25% acetic anhydride) for 10 min. Slides were then permeabilized in 1% Triton X-100 for 30 min and washed with DEPC-PBS. Endogenous peroxidase was blocked by incubation in 0.3% H<sub>2</sub>O<sub>2</sub> for 30 min. Tissue sections were equilibrated in hybridization buffer (1 $\times$  prehybridization, 0.1 mg/ml yeast tRNA, 0.05 mg/ml heparin, and 50% formamide) for 1 h and incubated with probes at 65°C overnight. After washing in 0.2 $\times$  SSC at 65°C, bound riboprobes were detected by horseradish peroxidase (HRP)-conjugated anti-DIG (diluted 1:2000; Roche) or anti-FL (diluted 1:2000; Roche) antibodies followed by fluorescent staining with TSA Systems (PerkinElmer). After mounting sections in VECTASHIELD, images were obtained on a Zeiss LSM 700 Confocal Microscope equipped with a 20 $\times$  air Plan-Apochromat objective (NA 0.8). A minimum of three animals per genotype and age were compared in ISH experiments.

**Image quantification.** To quantify the number of *Col19a1*-expressing cells with *Sst* and *Spon1* or *Col25a1*, we manually counted >150 *Col19a1<sup>+</sup>* cells and >250 *Sst<sup>+</sup>* cells in supragranular and infragranular layers of different cortical regions or different layers/subregions of the hippocampus. Data were collected from three mice per genotype (WT or *Sst-Cre::Rosa-Stop-tdT*). These data were used to generate the percentage of *Col19a1* cells that express *Sst* and vice versa. We used similar methods to quantify the percentage of *Col19a1* cells that coexpressed SST and *Spon1* or SST and *Col25a1*.

For quantification of perisomatic synapses, single optical sections of confocal images were analyzed, and the density of synaptic elements (Syt2<sup>+</sup> puncta) was quantified per unit length of the cell surface in ImageJ. Student's *t* tests were used to determine any significant difference of the mean values between groups.

The intensity of presynaptic terminals in sections of cortex was measured in ImageJ. Three to four animals (eight more sections per animal) were analyzed per genotype, and the mean values were compared between groups. Student's *t* tests were used to determine any significant difference of the mean values between groups.

**Quantitative real-time PCR.** RNA was isolated using the Bio-Rad Total RNA Extraction from Fibrous and Fatty Tissue kit (Bio-Rad). cDNAs were generated from 250 ng RNA with the Invitrogen Superscript II Reverse Transcription First Strand cDNA Synthesis Kit (Thermo Fisher Scientific). Quantitative real-time PCR (qRT-PCR) was performed on a Chromo4 Four Color Real-Time PCR system (Bio-Rad) using iTaq SYBRGreen Supermix (Bio-Rad; Su et al., 2016). *Col19a1* primers for qRT-PCR were as follows: 5'-ATT GGA CAT AAG GGC GAC AA-3'; and 5'-AGT CTC CTT TGG CTC CTG GT-3'. *Gapdh* primers

for qRT-PCR were as follows: 5'-CGT CCC GTA GAC AAA ATG GT-3'; and 5'-TTG ATG GCA ACA ATC TCC AC-3'. *Spon1* primers for qRT-PCR were as follows: 5'-CGG CCA CAG GAC AGT TAC TC-3'; and 5'-TCA AAG AGA ACC AGG AGG GA-3'. *Col25a1* primers for qRT-PCR were as follows: 5'-GAT TCT CCT CTT GG CCT CT-3'; and 5'-AAA TAA GAA CGG CCA GGG AG-3'. *Sst* primers for qRT-PCR were as follows: 5'-GAA GGA GAC GCT ACC GAA G-3'; and 5'-AAA GCC AGG ACG ATG CAG-3'. qRT-PCR primers were designed over introns. The following cycling conditions were used with 10 ng RNA: 95°C for 30 s, followed by 40 cycles of amplification (95°C for 5 s, 60°C for 30 s, 55°C for 60 s, read plate) and a melting curve analysis. Relative quantities of RNA were determined using the  $\Delta\Delta$ -Ct method. A minimum of double experiments (each in triplicate) was run for each gene. Each individual run included separate *Gapdh* control reactions.

**Mouse weight measurement.** Mice were weighed at postnatal day 7 (P7), P14, P21, and P28. At least 10 mice were weighed per genotype and age. Mean weights were compared between groups, and one-way ANOVAs (using GraphPad Prism 8) were used to determine significant differences between groups.

**Nest-building assay.** Approximately 1 h before the dark phase of the light/dark cycle, mice were transferred into clean cages and housed singly for 12 h. One cotton nestlet was added in the same location of each cage. Each cotton nestlet weighed ~2.8 g at the onset of the experiment. The next morning, we assessed the nests in two ways. First, we visually scored each nest with a rating scale of 1–5 (1, >90% of the cotton nestlet remained intact; 2, 50–90% of the cotton nestlet remained intact; 3, 50–90% of the nestlet was shredded; 4, >90% was shredded but the nest was not complete; 5, >90% was torn and the nest was complete; Su et al., 2016; Deacon, 2006). Second, we weighed the portions of each nestlet that remained unused (which included any fragments weighing >0.1 g). All mice analyzed were 3–5 months old. The data presented were obtained from 6 *Col19a1<sup>fl/fl</sup>::Nes-cre* mutants and 8 littermate controls; 9 *Col19a1<sup>fl/fl</sup>::Gad2-cre* mutant and 13 littermate controls; and 14 *Col19a1<sup>fl/fl</sup>::Sst-cre* mutant and 6 littermate controls. Each mouse was assayed once per week, and this was repeated for 3 consecutive weeks. Student's *t* tests were used to determine significant differences between groups.

**Pentylenetetrazol-induced seizures.** To assess responses to drug-induced seizures, mice were injected with pentylenetetrazol (PTZ; 40  $\mu$ g/kg in 0.2 ml PBS) and then visually monitored for 15 min as described previously (Brooks et al., 2015; Su et al., 2016). Seizures were scored manually every minute after with the following scoring criteria: 0, normal activity; 1, reduced motility and prostate position; 2, partial clonus; 3, generalized clonus; 4, tonic-clonic seizure; and 5, death. All data were from 9 *Col19a1<sup>fl/fl</sup>::Nes-cre* mutant mice and 10 littermate controls; 15 *Col19a1<sup>fl/fl</sup>::Gad2-cre* and 10 littermate controls; and 16 *Col19a1<sup>fl/fl</sup>::Sst-cre* mutant mice and 14 littermate controls. All mice were 3–5 months old. Student's *t* tests were used to determine significant differences between groups at each time point.

**Crawley sociability and social novelty preference assay.** The Crawley sociability and social novelty preference assay was performed in a standard three-chamber box (Stoelting), with an open middle chamber that allows free access to each of the flanking chambers, as described previously (Kaidanovich-Beilin et al., 2011; Su et al., 2016). The test mouse was habituated in the middle chamber for 5 min, and a novel conspecific male was placed inside a wire containment cup in one side of the chamber. The duration of contacts between the subject and either the empty housing or the housing containing the novel conspecific was recorded by three video cameras. A single 10 min session was performed for each subject. Social memory was immediately tested by placing a new, novel conspecific male in the empty chamber and leaving the previous (now familiar) conspecific in place. For 10 min, the duration of contacts between the subject and either the novel conspecific or the familiar conspecific was recorded. StreamPix5 was used to analyze the data. All data were from 16 *Col19a1<sup>fl/fl</sup>::Nes-cre* mutant mice and 17 littermate controls; 22 *Col19a1<sup>fl/fl</sup>::Gad2-cre* and 17 littermate controls; and 14 *Col19a1<sup>fl/fl</sup>::Sst-cre* mutant mice and 15 littermate controls. All mice were 3–5 months old. Student's *t* tests were used to determine significant differences between groups.

**Experimental design and statistical analysis.** The ages, genotypes, experimental conditions, and number of animals used for each experiment are included both in the Materials and Methods above and in the figure legends. For all experiments, both male and female mice were used. For imaging-based experiments, at least three to four animals were analyzed per genotype and group. For behavioral experiments, the number of animals ranged from 6 to 22, depending on the task. Student's *t* test or one-way ANOVAs were used to determine any significant difference of the mean values between groups using GraphPad Prism 8. For each experiment, the statistical test used, the number of animals analyzed, and the specific *p* values are listed in the Results and/or in the figure legends.

## Results

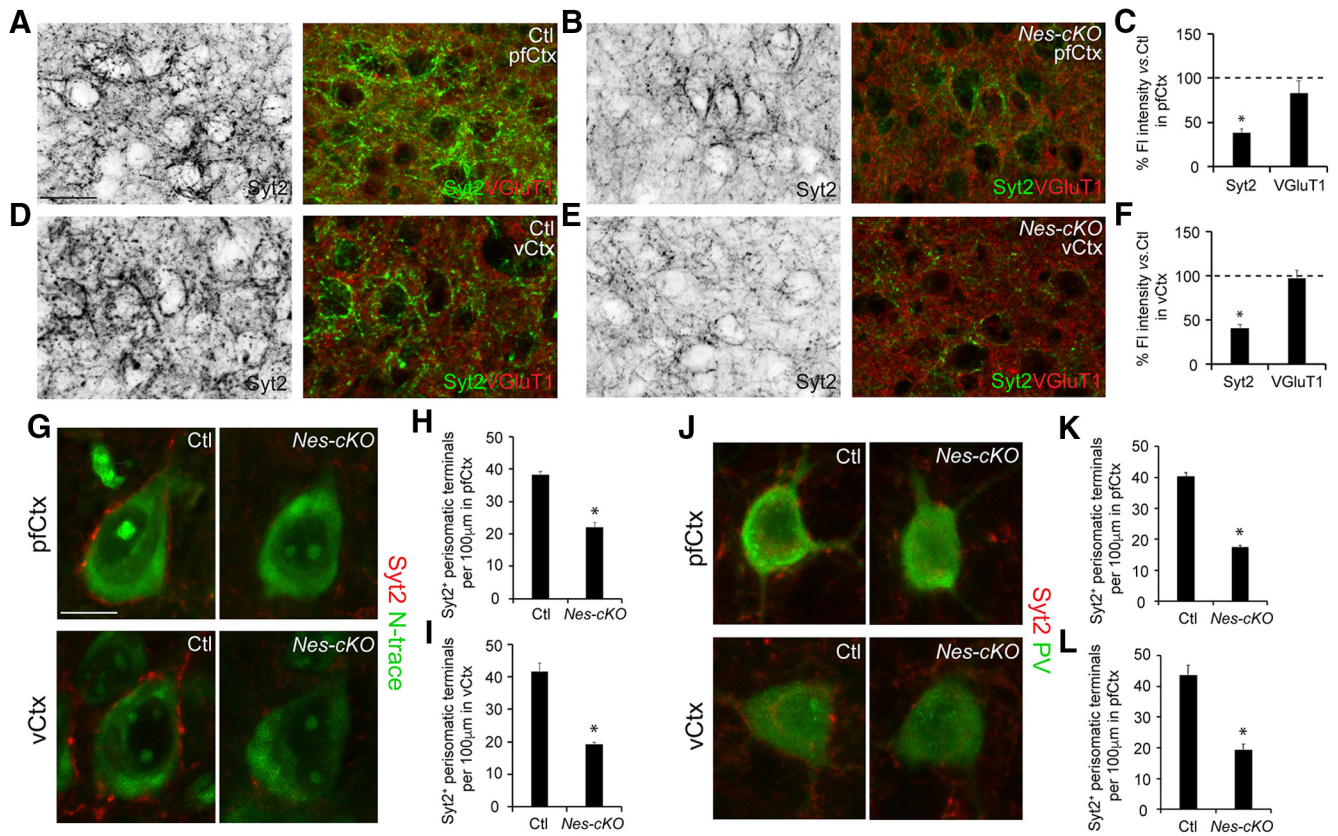
### Brain-derived Collagen XIX is necessary for perisomatic inhibitory synapse formation

Global loss of *Col19a1* impairs the formation of PV<sup>+</sup> inhibitory nerve terminals in the developing cerebral cortex and subiculum (Su et al., 2010, 2016). This could result from the specific loss of Collagen XIX from neurons or, indirectly, from roles of Collagen XIX outside of the brain (Sumiyoshi et al., 1997). To distinguish these possibilities, we generated a conditional allele of *Col19a1* (*Col19a1<sup>fl/fl</sup>*) in which LoxP sites flanked exon 4 of *Col19a1*, the same region deleted in *Col19a1<sup>-/-</sup>* mutants (Fig. 1A; Sumiyoshi et al., 2004). We initially crossed this conditional allele with *Nes-Cre*, a driver line in which Cre Recombinase (Cre) is generated by neuronal and glial progenitors early in the developing brain (Tronche et al., 1999). *Col19a1<sup>fl/fl</sup>::Nes-Cre<sup>+</sup>* mutant mice are born in expected Mendelian ratios, are viable and fertile, and are indistinguishable from littermate controls as adults. However, *Col19a1<sup>fl/fl</sup>::Nes-Cre<sup>+</sup>* mutants are significantly smaller than littermate controls during the first 4 weeks of postnatal development, although they are significantly larger than *Col19a1<sup>-/-</sup>* mutants at these ages (Fig. 1B).

To assess the formation of inhibitory GABAergic synapses in *Col19a1<sup>fl/fl</sup>::Nes-Cre<sup>+</sup>* mutants, GABAergic nerve terminals were labeled with antibodies directed against Syt2, a synaptic vesicle-associated protein whose expression in neocortex is largely restricted to PV<sup>+</sup> GABAergic interneurons (Sommeijer and Levelt, 2012; Su et al., 2016). In layer V of both visual and prefrontal cortex, we detected significantly fewer Syt2<sup>+</sup> nerve terminals in *Col19a1<sup>fl/fl</sup>::Nes-Cre<sup>+</sup>* mutants compared with controls (Fig. 2A–F). While brain-derived Collagen XIX appears to be necessary for GABAergic synapse formation, we detected no significant alteration in the density, distribution, or morphology of excitatory nerve terminals (e.g., those labeled with antibodies against VGluT1 in these regions of *Col19a1<sup>fl/fl</sup>::Nes-Cre<sup>+</sup>* mice (Fig. 2A–F).

Since PV<sup>+</sup> interneurons generate perisomatic inhibitory synapses onto pyramidal neurons and other PV<sup>+</sup> interneurons (Pfeffer et al., 2013; Pi et al., 2013), we next examined the distribution of these nerve terminals in the developing neocortex of *Col19a1<sup>fl/fl</sup>::Nes-Cre<sup>+</sup>* mutants. Neuronal somas in layer V of neocortex were labeled with either NeuroTrace or antibodies against PV, and perisomatic inhibitory nerve terminals were labeled with antibodies against Syt2. Significantly fewer Syt2<sup>+</sup> perisomatic synapses were observed contacting the somas of both types of neurons in the absence of brain-derived Collagen XIX (Fig. 2G–L).

Expression of *Col19a1* is restricted to subsets of GABAergic interneurons in neocortex (Su et al., 2016), so we repeated these analyses in *Col19a1<sup>fl/fl</sup>::Gad2-Cre<sup>+</sup>* mutants. Cre is generated in the majority of neocortical GABAergic cells in *Gad2-Cre<sup>+</sup>* mice, including in *Col19a1*-expressing cells (data not shown).



**Figure 2.** Loss of brain-derived Collagen XIX leads to impaired inhibitory perisomatic synapse formation. **A, B**, Immunostaining for Syt2 and VGLuT1 in layer V of prefrontal cortex (pfCtx) in P14 *Col19a1<sup>fl/fl</sup>::Nes-Cre* (*Nes-cKO*) mutants and littermate controls (Ctl). **C**, Mean fluorescent intensity of Syt2 and VGLuT1 in layer V of pfCtx in P14 *Nes-cKO* and Ctl mice. Dashed line represents Ctl values. Data represent the mean  $\pm$  SD;  $n = 3$ . \*Differs from Ctl by  $p = 0.00006$  by Student's  $t$  test. **D, E**, Immunostaining for Syt2 and VGLuT1 in layer V of visual cortex (vCtx) in P14 *Nes-cKO* and Ctl mice. **F**, Mean fluorescent intensity of Syt2 and VGLuT1 in layer V of vCtx in P14 *Nes-cKO* and Ctl mice. Dashed line represents Ctl values. Data represent the mean  $\pm$  SD;  $n = 3$ . \*Differs from Ctl by  $p = 0.0005$  by Student's  $t$  test. **G**, Immunostained Syt2<sup>+</sup> perisomatic synapses were analyzed on NeuroTrace (N-trace)-labeled neurons in single optical sections from layer V of pfCtx and vCtx in P14 *Nes-cKO* and Ctl mice. **H, I**, Reduced numbers of Syt2<sup>+</sup> perisomatic synapses (per unit length of cell soma) on N-trace-labeled neurons in pfCtx and vCtx of P14 *Nes-cKO* mutants. Data represent the mean  $\pm$  SD;  $n = 3$ . \*Differs from Ctl by  $p = 0.0002$  (pfCtx),  $p = 0.0003$  (vCtx) by Student's  $t$  test. **J**, Immunostained Syt2<sup>+</sup> perisomatic synapses were analyzed on PV-immunolabeled neurons in single optical sections from layer V of pfCtx and vCtx in P14 *Nes-cKO* and Ctl mice. **K, L**, Reduced numbers of Syt2<sup>+</sup> perisomatic synapses (per unit length of cell soma) on PV<sup>+</sup> neurons in pfCtx and vCtx of P14 *Nes-cKO* mutants. Data represent means  $\pm$  SD;  $n = 3$ . \*Differs from Ctl by  $p = 0.00,001$  (pfCtx),  $p = 0.0008$  (vCtx) by Student's  $t$  test. Scale bars: (in **A, B, D, E**, 20  $\mu$ m; (in **G, J**, 10  $\mu$ m).

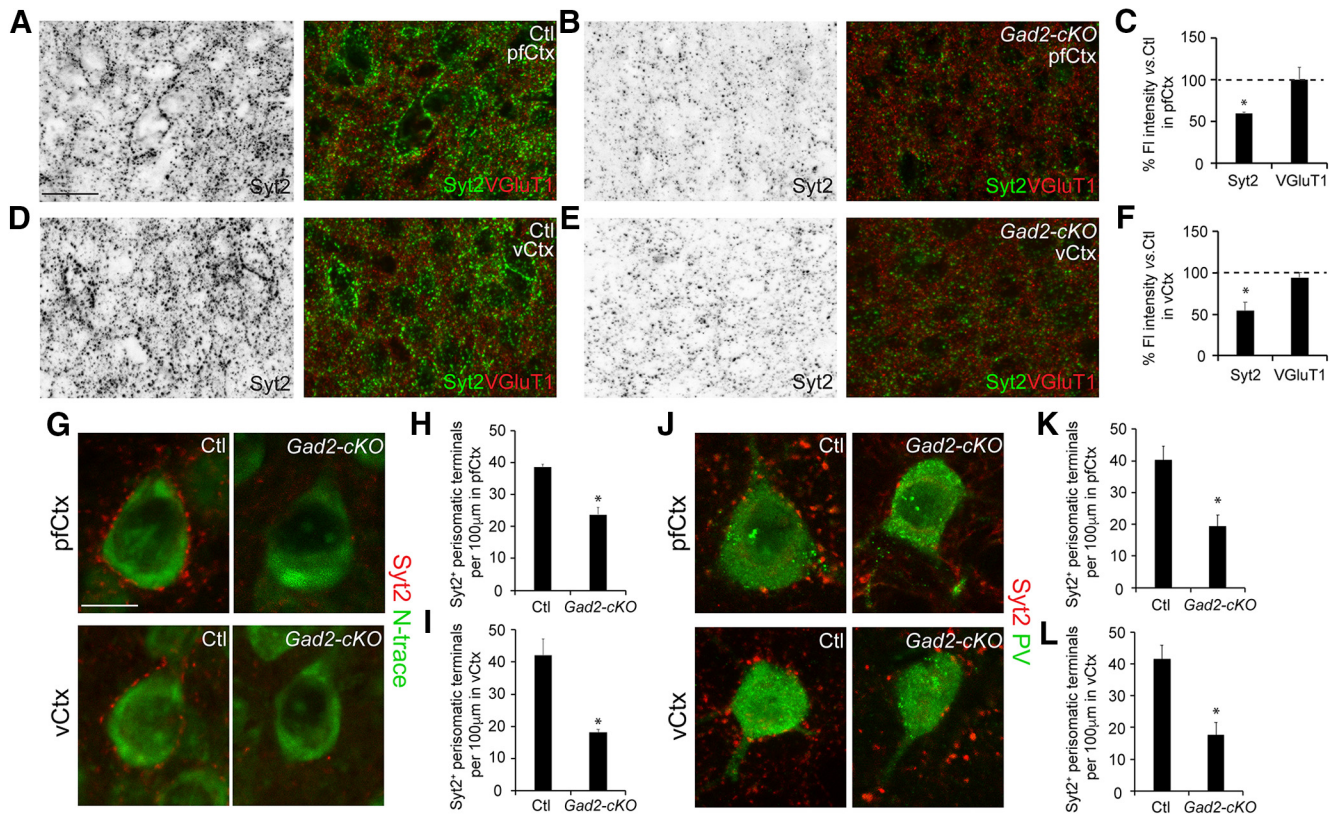
*Col19a1<sup>fl/fl</sup>::Gad2-Cre<sup>+</sup>* mutant mice are born in expected Mendelian ratios, are viable and fertile, and are indistinguishable from littermate controls as adults. Loss of Collagen XIX from *Gad2*-expressing cells in P14 *Col19a1<sup>fl/fl</sup>::Gad2-Cre<sup>+</sup>* mutants resulted in reduced Syt2<sup>+</sup> perisomatic nerve terminals, as described above in *Col19a1<sup>fl/fl</sup>::Nes-Cre<sup>+</sup>* mutants (Fig. 3). Together, studies in both *Col19a1<sup>fl/fl</sup>::Nes-Cre<sup>+</sup>* and *Col19a1<sup>fl/fl</sup>::Gad2-Cre<sup>+</sup>* demonstrate that interneuron-derived Collagen XIX is necessary for the formation of PV<sup>+</sup> inhibitory circuits in the developing neocortex.

### Loss of brain-derived collagen XIX results in phenotypes associated with dysfunctional inhibitory neocortical circuits

The loss of PV<sup>+</sup> perisomatic synapses in cerebral cortex (because of the loss of Collagen XIX or a number of other genetic and environmental factors) leads to spontaneous seizures, enhanced susceptibility to drug-induced seizures, and the acquisition of a number of behaviors associated with neurodevelopmental disorders, such as schizophrenia (Schwaller et al., 2004; Krook-Magnuson et al., 2013; Rossignol et al., 2013; Wöhr et al., 2015; Su et al., 2016). To assess seizure susceptibility in *Col19a1<sup>fl/fl</sup>::Nes-Cre<sup>+</sup>* and *Col19a1<sup>fl/fl</sup>::Gad2-Cre<sup>+</sup>* mutants (and controls), we administered low doses of the GABA antagonist PTZ and monitored seizure activity for 16 min. At low doses (40 mg/

kg), PTZ induced mild motor seizures in control mice (Fig. 4A,B). Identical doses of PTZ resulted in significantly enhanced seizure susceptibility in both *Col19a1<sup>fl/fl</sup>::Nes-Cre<sup>+</sup>* and *Col19a1<sup>fl/fl</sup>::Gad2-Cre<sup>+</sup>* mutants, in terms of both increased seizure scores (i.e., seizure severity) and faster onset of motor seizures (Fig. 4A,B).

Next, to assess whether either mutant exhibits altered behavior associated with schizophrenia, we assessed mutant performance in a nest-building assay (which models the negative symptoms associated with schizophrenia; Belforte et al., 2010; Albrecht and Stork, 2012; Pedersen et al., 2014) and in a preference for social novelty task (which models social affiliation and memory; Moy et al., 2004; Fig. 4C,H,I). Previously, we showed that *Col19a1<sup>-/-</sup>* mice exhibit impaired performance in both of these tasks (Su et al., 2016). Here, our results indicate impaired nest-building activity in both *Col19a1<sup>fl/fl</sup>::Nes-Cre<sup>+</sup>* and *Col19a1<sup>fl/fl</sup>::Gad2-Cre<sup>+</sup>* mutants (Fig. 4D–G). Moreover, while control mice spend significantly more time investigating novel conspecific mice (compared with the time they spend investigating familiar mice), both conditional mutants spend equal time investigating novel and familiar conspecifics (Fig. 4J–M). Thus, loss of either brain- or interneuron-derived Collagen XIX leads to the acquisition of abnormal behaviors.

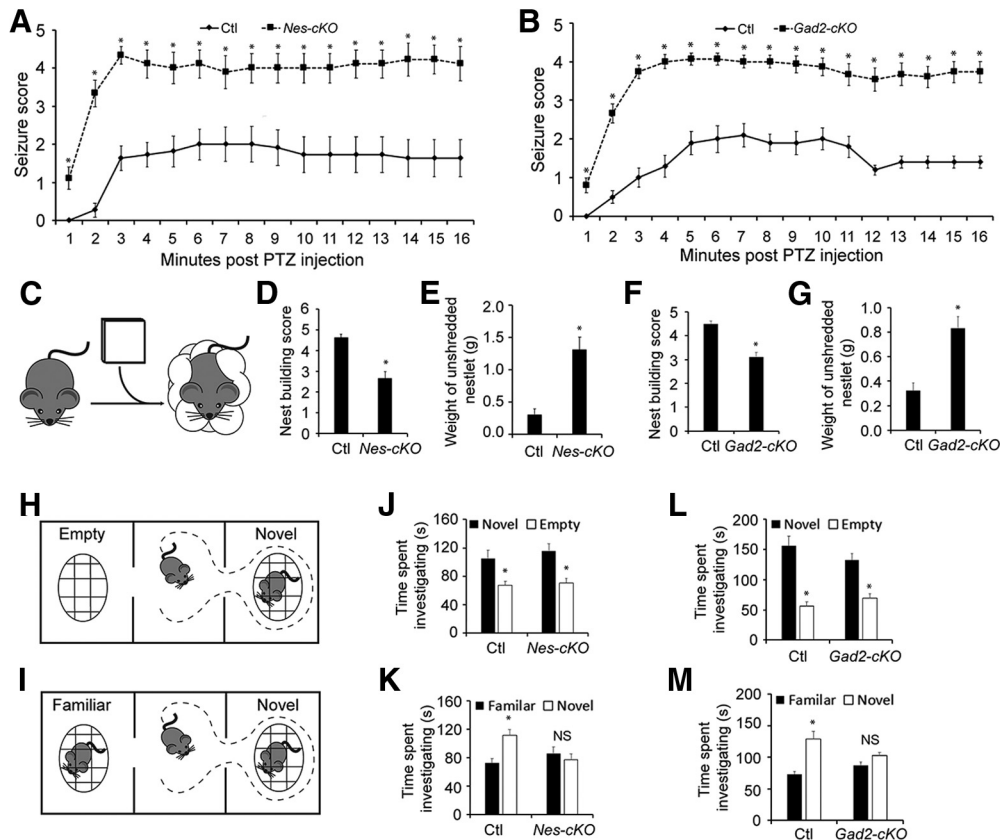


**Figure 3.** Loss of GABAergic neuron-derived Collagen XIX leads to impaired inhibitory perisomatic synapse formation. **A, B**, Immunostaining for Syt2 and VGLUT1 in layer V of prefrontal cortex (pfCtx) in P14 *Col19a1<sup>fl/fl</sup>::Gad2-Cre* (*Gad2-cKO*) mutants and littermate controls (Ctl). **C**, Mean fluorescent intensity of Syt2 and VGLUT1 in layer V of pfCtx in P14 *Gad2-cKO* and Ctl mice. Dashed line represents Ctl values. Data represent the mean  $\pm$  SD;  $n = 3$ . \*Differs from Ctl by  $p = 0.001$  by Student's  $t$  test. **D, E**, Immunostaining for Syt2 and VGLUT1 in layer V of visual cortex (vCtx) in P14 *Gad2-cKO* and Ctl mice. **F**, Mean fluorescent intensity of Syt2 and VGLUT1 in layer V of vCtx in P14 *Gad2-cKO* and Ctl mice. Dashed line represents Ctl values. Data represent the mean  $\pm$  SD;  $n = 3$ . \*Differs from Ctl by  $p = 0.003$  by Student's  $t$  test. **G**, Immunostained Syt2<sup>+</sup> perisomatic synapses were analyzed on NeuroTrace (N-trace)-labeled neurons in single optical sections from layer V of pfCtx and vCtx in P14 *Gad2-cKO* and Ctl mice. **H, I**, Reduced numbers of Syt2<sup>+</sup> perisomatic synapses (per unit length of cell soma) on N-trace-labeled neurons in pfCtx and vCtx of P14 *Gad2-cKO* mutants. Data represent the mean  $\pm$  SD;  $n = 3$ . \*Differs from Ctl by  $p = 0.0009$  (pfCtx),  $p = 0.003$  (vCtx) by Student's  $t$  test. **J**, Immunostained Syt2<sup>+</sup> perisomatic synapses were analyzed on PV-immunolabeled neurons in single optical sections from layer V of pfCtx and vCtx in P14 *Gad2-cKO* and Ctl mice. **K, L**, Reduced numbers of Syt2<sup>+</sup> perisomatic synapses (per unit length of cell soma) on PV<sup>+</sup> neurons in pfCtx and vCtx of P14 *Gad2-cKO* mutants. Data represent the mean  $\pm$  SD;  $n = 3$ . \*Differs from Ctl by  $p = 0.006$  (pfCtx),  $p = 0.004$  (vCtx) by Student's  $t$  test. Scale bars: (in **A, B, D, E**) 20  $\mu$ m; (in **G, J**) 10  $\mu$ m.

### SST<sup>+</sup> interneurons in cerebral cortex generate *Col19a1*

While *Col19a1* mRNA is generated by cortical interneurons, little, if any, is generated by PV<sup>+</sup> interneurons (Su et al., 2016). Unbiased transcriptomic studies recently have suggested that it is instead generated by SST<sup>+</sup> cells in neocortex. To test whether telencephalic SST<sup>+</sup> interneurons do in fact generate *Col19a1*, we generated riboprobes against both *Col19a1* and *Sst* mRNAs and performed *in situ* hybridization (Fig. 5A,B). We also performed *in situ* hybridization with *Col19a1* riboprobes on brain slices from *Sst-Cre::Rosa-Stop-tdT* transgenic mice, in which SST<sup>+</sup> interneurons are fluorescently labeled with tdT (Fig. 5C,D). In both sets of experiments, we observed the expression of *Col19a1* mRNA by SST<sup>+</sup> interneurons in both supragranular and infragranular layers of the developing neocortex (Fig. 5A–F). These results suggest that the majority of brain-derived Collagen XIX is produced by SST<sup>+</sup> interneurons. To test this hypothesis, we assessed *Col19a1* mRNA expression in *Sst-Cre::Rosa-Stop-DTA* mice, in which diphtheria toxin A (DTA) is selectively expressed in SST<sup>+</sup> neurons leading to the ablation of these cells (Ivanova et al., 2005; Tuncdemir et al., 2016). As expected, qRT-PCR revealed a significant reduction in *Sst* mRNA in *Sst-Cre::Rosa-Stop-DTA* mutants (Fig. 5G). Thus, SST<sup>+</sup> interneurons appear to be a predominant source of *Col19a1* mRNA in the developing mouse neocortex.

Clustering of cell types by single-cell transcriptomic analysis has revealed the presence of at least two (if not more) transcriptionally distinct subsets of GABAergic SST<sup>+</sup> interneurons in mouse cortex (Hrvatín et al., 2018; Mayer et al., 2018). In these studies, *Spon1* (which encodes F-Spondin) and *Col25a1* (which encodes Collagen XXV) are generated by distinct subtypes of SST<sup>+</sup> interneurons (with *Spon1* mRNA being enriched in type I SST<sup>+</sup> interneurons and *Col25a1* mRNA being enriched in type II SST<sup>+</sup> interneurons; Hrvatín et al., 2018; Mayer et al., 2018). To test whether *Col19a1* is generated by a select subtype of SST<sup>+</sup> interneurons, we generated riboprobes against *Spon1* and *Col25a1*. *In situ* hybridization in *Sst-Cre::Rosa-Stop-tdT* transgenic mice revealed that *Spon1* and *Col25a1* are generated in distinct cortical layers: *Spon1*<sup>+</sup>/*Sst*<sup>+</sup> cells are distributed in infragranular cortical layers, and *Col25a1*<sup>+</sup>/*Sst*<sup>+</sup> cells are distributed in supragranular cortical layers (Fig. 5I–L). This suggests laminar specificity of type I and type II SST<sup>+</sup> neurons. Interestingly, while *in situ* hybridization revealed that both *Spon1* and *Col25a1* are generated in SST<sup>+</sup> interneurons (labeled in *Sst-Cre::Rosa-Stop-tdT* transgenic mice; Fig. 5I–L), we found that *Spon1* mRNA, but not *Col25a1* mRNA, is significantly reduced in the neocortex of *Sst-Cre::Rosa-Stop-DTA* mutant mice (Fig. 5H). As an aside, this result, coupled with the persistence of some *Sst* mRNA in these mutants (Fig. 5G), suggests that perhaps only a subset of *Sst*<sup>+</sup> neurons is ablated in *Sst-Cre::Rosa-*



**Figure 4.** Mice lacking brain- or GABAergic neuron-derived Collagen XIX are more susceptible to drug-induced seizures and exhibit altered behavior. **A, B**, Seizure scores for Ctl, *Nes-cKO*, and *Gad2-cKO* mutants were recorded for 16 min after the administration of PTZ. Data represent the mean  $\pm$  SEM; in **A**,  $n = 11$  Ctl and  $n = 9$  *Nes-cKO*; in **B**,  $n = 10$  Ctl and  $n = 15$  *Gad2-cKO*. \*Differs from Ctl by  $p < 0.01$  by Student's  $t$  test. **C**, Schematic representation of nest-building behavior. **D–G**, Nests were scored manually after 12 h (**D, F**) or by weighing unused nestlets after 12 h (**E, G**). Data represent the mean  $\pm$  SEM;  $p = 0.0009$  for weight and  $p = 0.0005$  for score by Student's  $t$  test; in **D** and **E**,  $n = 8$  Ctl and  $n = 6$  *Nes-cKO*; in **F** and **G**,  $n = 13$  Ctl and  $n = 9$  *Gad2-cKO*. \*Differs from Ctl by  $p = 0.0002$  for weight and  $p = 0.0002$  for score by Student's  $t$  test. **H, I**, Schematic of sociability (**H**) and social memory (**I**) assays. **J–M**, *Nes-cKO*, *Gad2-cKO*, and Ctl mice spent significantly more time investigating novel conspecifics versus empty chambers (**J, L**); while Ctl mice also spent significantly more time investigating novel conspecifics versus familiar conspecifics (**K, M**), the same was not true for *Nes-cKO* and *Gad2-cKO* mutants. Data represent the mean  $\pm$  SEM; in **J** and **K**,  $n = 17$  Ctl and  $n = 16$  *Nes-cKO*; in **L** and **M**,  $n = 17$  Ctl and  $n = 22$  *Gad2-cKO*. \*Differs from empty or familiar conspecific by  $p = 0.006$  (for Ctl in **J**),  $p = 0.0009$  (for *Nes-cKO* in **J**),  $p = 0.001$  (for Ctl in **K**),  $p = 0.0009$  (for *Nes-cKO* in **K**),  $p = 0.00005$  (for Ctl in **L**),  $p = 0.00002$  (for *Nes-cKO* in **L**),  $p = 0.0002$  (for Ctl in **M**) by Student's  $t$  test. NS, Not significantly different.

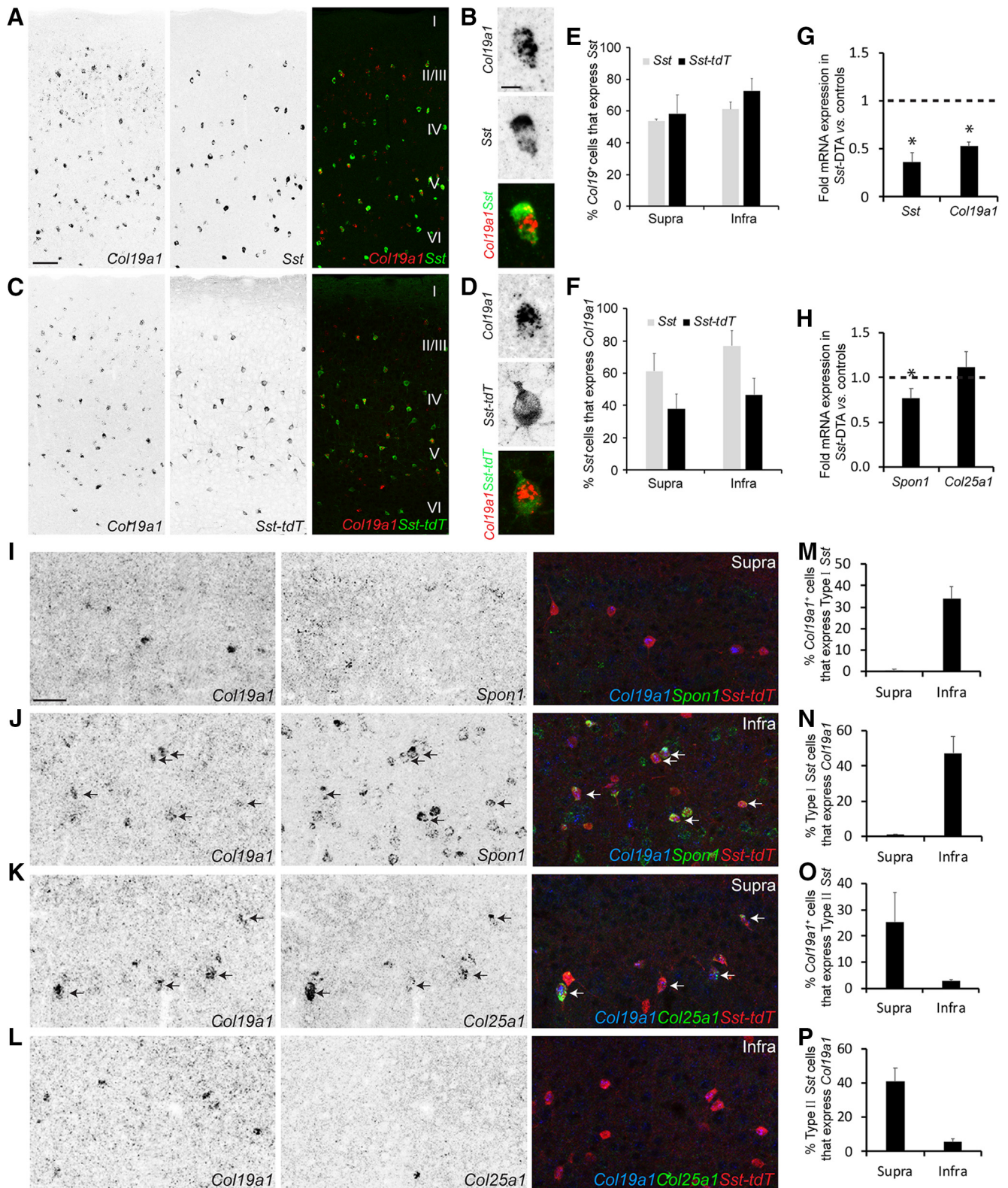
*Stop-DTA* mice. Finally, combining *in situ* hybridization for *Col19a1* and either *Spon1* and *Col25a1* in *Sst-Cre::Rosa-Stop-tdT* transgenic mice revealed that both type I and type II SST<sup>+</sup> interneurons generate *Col19a1* in neocortex (Fig. 5I–P).

### SST-derived Collagen XIX is necessary for perisomatic inhibitory synapse formation

Next, to specifically test whether SST-derived Collagen XIX is necessary for the formation of PV<sup>+</sup> perisomatic synapses, we generated *Col19a1<sup>fl/fl</sup>::Sst-Cre<sup>+</sup>* mutants. These mutants are born in expected Mendelian ratios, are viable and fertile, and exhibit significant reduction in *Col19a1* mRNA expression in neocortex ( $0.59 \pm 0.04$ -fold *Col19a1* mRNA expression in *Col19a1<sup>fl/fl</sup>::Sst-Cre<sup>+</sup>* neocortex vs control;  $p = 0.003$  by Student's  $t$  test). *Col19a1<sup>fl/fl</sup>::Sst-Cre<sup>+</sup>* mutants exhibit similar structural and behavioral phenotypes described above for *Col19a1<sup>fl/fl</sup>::Nes-Cre<sup>+</sup>* and *Col19a1<sup>fl/fl</sup>::Gad2-Cre<sup>+</sup>* mutants (Figs. 2–4) and previously for *Col19a1<sup>-/-</sup>* mutants (Su et al., 2016; Figs. 6–8). This includes a reduced density of both Syt2<sup>+</sup> and GAD67<sup>+</sup> inhibitory nerve terminals in layer V of the developing neocortex (Figs. 6A–F, 7) and a significant reduction in Syt2<sup>+</sup> perisomatic synapses onto neocortical pyramidal neurons and PV<sup>+</sup> interneurons (Fig. 6G–L). As was the case for all other mutant alleles of *Col19a1* examined to date, no differences in VGluT1<sup>+</sup> excitatory nerve

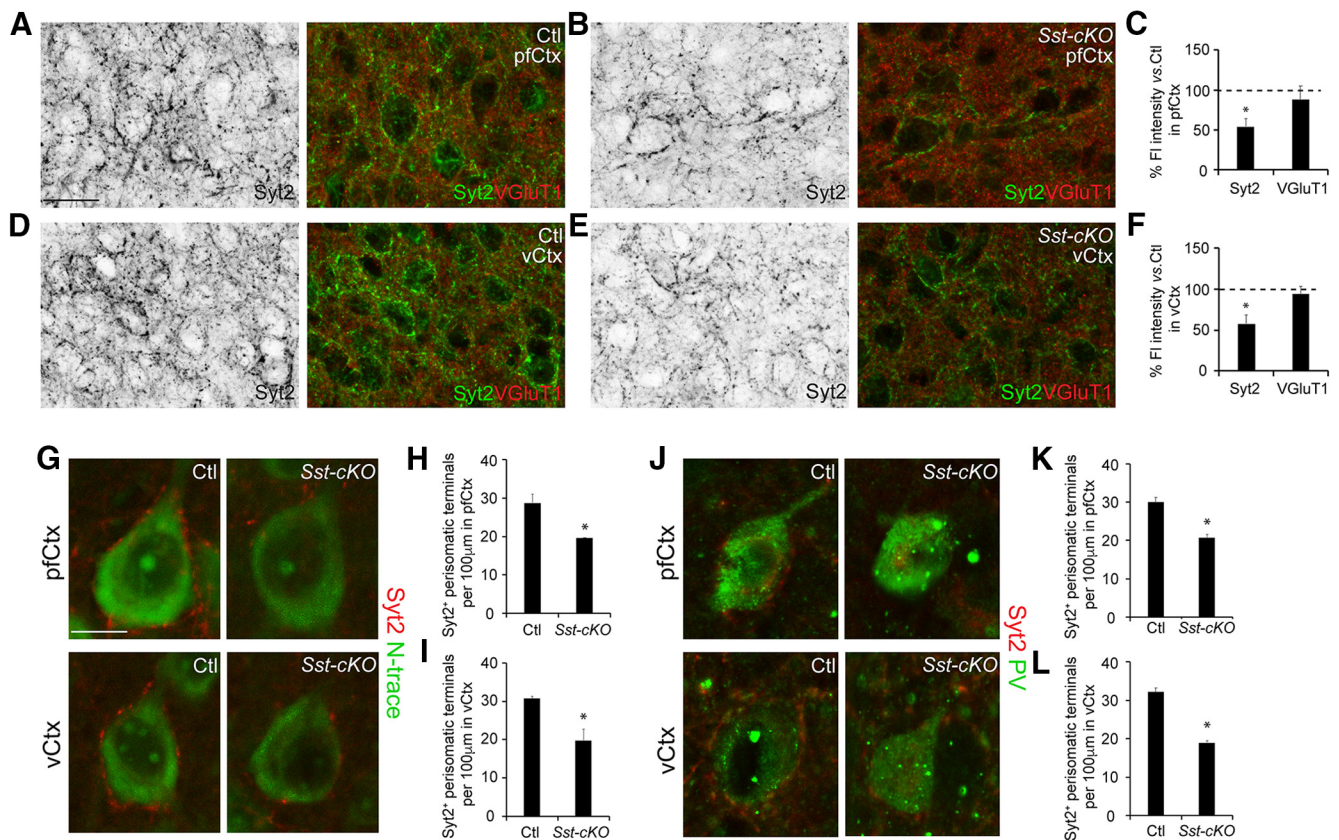
terminals were observed in *Col19a1<sup>fl/fl</sup>::Sst-Cre<sup>+</sup>* mutants compared with littermate controls (Fig. 6A–F). Defects in PV<sup>+</sup>/Syt2<sup>+</sup> perisomatic synapse assembly in the absence of SST-derived Collagen XIX do not merely reflect a delay in neocortical development since similar reductions in these inhibitory synapses were observed in adult *Col19a1<sup>fl/fl</sup>::Sst-Cre<sup>+</sup>* mutants (Fig. 8A–I). Moreover, these adult mutants exhibited significantly elevated susceptibility to PTZ-induced seizures (Fig. 8R) and impaired performance in nest-building and social preference tasks (Fig. 8S–V). Importantly, all of these changes in PV<sup>+</sup>/Syt2<sup>+</sup> perisomatic synapses and altered behaviors do not arise from reduced numbers of PV<sup>+</sup> or SST<sup>+</sup> interneurons in the absence of SST-derived Collagen XIX (Fig. 8J–Q). Together, all of these results suggest that SST-derived Collagen XIX acts in a paracrine fashion for the assembly of PV<sup>+</sup> perisomatic synapses.

Finally, while SST-derived Collagen XIX is required for the proper assembly of perisomatic inhibitory synapses, these analyses did not rule out roles for other specific types of cortical interneurons in this process. To assess roles of other defined populations of neocortical interneurons, we crossed *Col19a1<sup>fl/fl</sup>* mice to other Cre driver lines that exhibit subtype-specific expression in neocortical interneurons. Although few PV<sup>+</sup> interneurons generate Collagen XIX (Su et al., 2010, 2016), we assessed the role of limited PV-derived Collagen XIX by crossing



**Figure 5.** Collagen XIX is generated by neocortical SST<sup>+</sup> neurons. **A–D**, *In situ* hybridization for *Col19a1* mRNA in SST<sup>+</sup> neurons in P14 neocortex. SST<sup>+</sup> neurons labeled by *in situ* hybridization (**A**, **B**) or through genetic labeling (**C**, **D**) in *Sst-Cre::Rosa-Stop-tdT* mice (*Sst-tdT*) mice. Cortical layers are labeled in **A** and **C**. High-magnification images of *Col19a1* mRNA in SST<sup>+</sup> neurons are shown in **B** and **D**. **E**, Quantification of the percentage of *Col19a1*<sup>+</sup> cells expressing *Sst* mRNA (in wild-type mice) or tdT (in *Sst-tdT* mice) in supra- or infra-granular (Supra) or infra-granular (Infra) layers of neocortex. Bars depict the mean  $\pm$  SD. **F**, Quantification of the percentage of SST cells (identified by *Sst* mRNA or tdT expression in *Sst-tdT* mice) that express *Col19a1* mRNA in neocortex. Bars depict the mean  $\pm$  SD. **G**, **H**, qRT-PCR analysis of *Sst*, *Col19a1*, *Col25a1*, and *Spon1* mRNA expression in neocortex of P7 control (dashed line) or *Sst-Cre::Rosa-Stop-DTA* mice. Bars depict the mean  $\pm$  SD. \* $p = 0.001$  for *Sst* (**G**),  $p = 0.0003$  for *Col19a1* (**G**),  $p = 0.04$  for *Spon1* (**H**) compared with control by Student's *t* test ( $n = 3$ ). **I–L**, *In situ* hybridization for *Col19a1* and either *Col25a1* or *Spon1* in the supra-granular (Supra) or infra-granular (Infra) layers of neocortex in P22 *Sst-Cre::Rosa-Stop-tdT* (*Sst-tdT*) mice. **M–P**, Quantitation of the coexpression of *Col19a1* in *Spon1*<sup>+</sup>/*Sst*<sup>+</sup> (type I) cells and *Col25a1*<sup>+</sup>/*Sst*<sup>+</sup> (type II) cells in the supra-granular (Supra) or infra-granular (Infra) layers of neocortex. Bars depict the mean  $\pm$  SD. Scale bars: (in **A**, **A**, **C**, 100  $\mu$ m; (in **B**, **B**, **D**, 10  $\mu$ m; (in **I**–**L**, 50  $\mu$ m).





**Figure 6.** Loss of SST<sup>+</sup> neuron-derived Collagen XIX leads to impaired inhibitory perisomatic synapse formation. **A, B**, Immunostaining for Syt2 and VGLUT1 in layer V of prefrontal cortex (pfCtx) in P14 *Col19a1<sup>fl/fl</sup>::Sst-Cre* (*Sst-cKO*) mutants and littermate controls (Ctl). **C**, Mean fluorescent intensity of Syt2 and VGLUT1 in layer V of pfCtx in P14 *Sst-cKO* and Ctl. Dashed line represents Ctl values. Data represent the mean  $\pm$  SD;  $n = 3$ . \*Differs from Ctl by  $p = 0.006$  by Student's  $t$  test. **D, E**, Immunostaining for Syt2 and VGLUT1 in layer V of visual cortex (vCtx) in P14 *Sst-cKO* and Ctl mice. **F**, Mean fluorescent intensity of Syt2 and VGLUT1 in layer V of vCtx in P14 *Sst-cKO* and Ctl mice. Dashed line represents Ctl values. Data represent the mean  $\pm$  SD;  $n = 3$ . \*Differs from Ctl by  $p = 0.007$  by Student's  $t$  test. **G**, Immunostained Syt2<sup>+</sup> perisomatic synapses were analyzed on NeuroTrace (N-trace)-labeled neurons in single optical sections from layer V of pfCtx and vCtx in P14 *Sst-cKO* and Ctl. **H, I**, Reduced numbers of Syt2<sup>+</sup> perisomatic synapses (per unit length of cell soma) on N-trace-labeled neurons in pfCtx and vCtx of P14 *Sst-cKO* mutants. Data represent the mean  $\pm$  SD;  $n = 3$ . \*Differs from Ctl by  $p = 0.005$  (pfCtx),  $p = 0.007$  (vCtx) by Student's  $t$  test. **J**, Immunostained Syt2<sup>+</sup> perisomatic synapses were analyzed on PV-immunolabeled neurons in single optical sections from layer V of pfCtx and vCtx in P14 *Sst-cKO* and Ctl mice. **K, L**, Reduced numbers of Syt2<sup>+</sup> perisomatic synapses (per unit length of cell soma) on PV<sup>+</sup> neurons in pfCtx (**K**) and vCtx (**L**) of P14 *Sst-cKO* mutants. Data represent the mean  $\pm$  SD;  $n = 3$ . \*Differs from Ctl by  $p = 0.001$  (pfCtx),  $p = 0.0007$  (vCtx) by Student's  $t$  test. Scale bars: (in **A, B, D, E**, 20  $\mu$ m; (in **G, J**, 10  $\mu$ m).

*Col19a1<sup>fl/fl</sup>* mice to PV-Cre mice. To inhibit the production of Collagen XIX from other subtypes of interneurons, we crossed our floxed allele to *Crh-Cre* mice. Cre is generated in a large subset of SST<sup>-</sup>/PV<sup>-</sup> interneurons in *Crh-Cre* mice (Taniguchi et al., 2011). The developmental assembly of inhibitory nerve terminals in these mutants was examined using approaches identical to those described above for all other conditional mutants. We observed no alterations in Syt2<sup>+</sup> inhibitory nerve terminals in for *Col19a1<sup>fl/fl</sup>::PV-Cre*<sup>+</sup> and *Col19a1<sup>fl/fl</sup>::Crh-Cre*<sup>+</sup> mutants (Fig. 9), suggesting that Collagen XIX is not required from these classes of interneurons for the assembly of perisomatic synapses.

## Discussion

### Early-born, cortical SST<sup>+</sup> interneurons orchestrate cortical circuit development

Despite sharing a common embryonic origin with PV<sup>+</sup> interneurons (Xu et al., 2004), SST<sup>+</sup> interneurons represent the earliest interneuron population to tangentially migrate into infragranular cortical layers and integrate into circuits (Miyoshi and Fishell, 2011; Tuncdemir et al., 2016; Lim et al., 2018). These early-born SST<sup>+</sup> interneurons not only help to shape patterns of synchronous activity in the developing cortex, but they also help

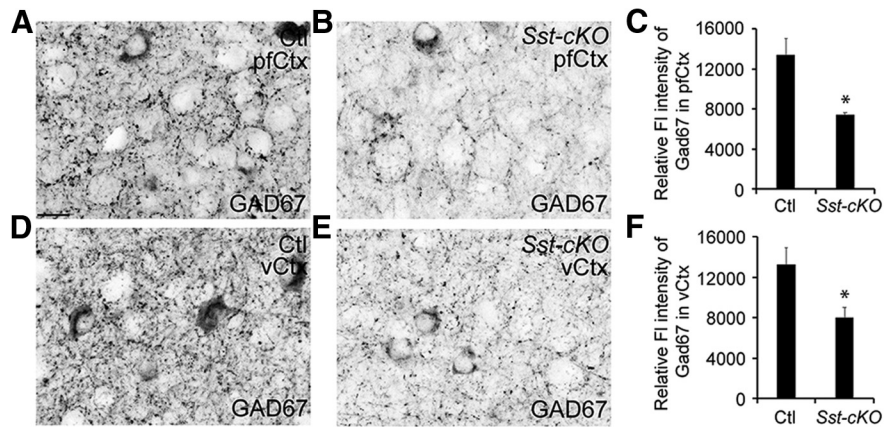
to shape the development of cortical connectivity (Bonifazi et al., 2009; Picardo et al., 2011; Tuncdemir et al., 2016; Wang et al., 2019). While genetic ablation of early-born SST<sup>+</sup> interneurons has demonstrated critical roles for these cells in orchestrating the formation and maturation of GABAergic circuits in the infragranular layers of neocortex (Tuncdemir et al., 2016; Wang et al., 2019), the mechanism underlying the paracrine action of these interneurons has remained unclear. One possibility is that the release of SST (which itself is a neuromodulator) or GABA from these early-born interneurons may have trophic, tropic, or synaptogenic effects on developing GABAergic circuits. Both SST and GABA have well established roles in neural development (Gonzalez et al., 1992; Ferriero et al., 1994; Owens and Kriegstein, 2002; Represa and Ben-Ari, 2005; Chen and Kriegstein, 2015; Liguz-Lecznar et al., 2016). Alternatively, the loss of SST<sup>+</sup> interneurons (which shape early patterns of activity in the developing cortex) may result in altered spontaneous network synchronization that impacts GABAergic circuit development (Bonifazi et al., 2009; Picardo et al., 2011). Precise patterns of activity, including GABAergic activity, during early development have well established roles in shaping neural circuit assembly and refinement (Katz and Shatz, 1996; Mohajerani et al., 2007; Huang, 2009; Griguoli and Cherubini, 2017). Interestingly,

blocking vesicular release of SST<sup>+</sup> interneurons early in development appears to have little effect on the development and maturation of infragranular GABAergic circuits (including GABAergic perisomatic synapses), suggesting that altered activity or the release of SST or GABA alone may not underlie the orchestration of cortical circuit development by early-born SST<sup>+</sup> interneurons (Tuncdemir et al., 2016; Wang et al., 2019). Data presented here support a third option for how these early-born interneurons influence GABAergic development—they generate and release synaptogenic cues that drive the assembly of GABAergic synapses in a paracrine fashion. Specifically, our data show that SST<sup>+</sup> interneurons generate Collagen XIX, a synaptogenic extracellular matrix protein capable of promoting GABAergic synapse formation (Su et al., 2016). Conditional deletion of this unconventional collagen from SST<sup>+</sup> interneurons (but not from PV<sup>+</sup> cells) is sufficient to impair the assembly of perisomatic, GABAergic synapses in the infragranular layers of neocortex. Not only does the loss of SST-derived Collagen XIX impair GABAergic synapse development, it increases seizure susceptibility and results in the acquisition of schizophrenia-related behaviors—phenotypes that are all associated with the loss or dysfunction of perisomatic, GABAergic synapses (Schwaller et al., 2004; Gonzalez-Burgos et al., 2011; Lewis et al., 2011; Paz and Huguenard, 2015). Thus, our data reveal a novel role for SST<sup>+</sup> interneurons in the production of paracrine factors that drive GABAergic synapse formation in the developing cortex.

While it remains unclear why early-born SST<sup>+</sup> interneurons assume the role of master coordinator of GABAergic circuit formation in the developing cortex, it appears to be a role conserved by early-born interneurons in other brain regions. For example, SST-expressing “hub” neurons orchestrate of circuit development in the developing hippocampus (Picardo et al., 2011; Villette et al., 2016). While there appear to be significant differences between these SST<sup>+</sup> hippocampal hub neurons and neocortical SST<sup>+</sup> interneurons (Tuncdemir et al., 2016; Wang et al., 2019), SST neurons in both regions produce Collagen XIX during GABAergic circuit formation (Su et al., 2010, 2016). Collagen XIX, however, appears dispensable for GABAergic circuit formation in stratum pyramidale of CA1 and CA3, and the loss of Collagen XIX impairs perisomatic GABAergic synapse formation in subiculum (Su et al., 2010). Thus, the role of SST-derived Collagen XIX in triggering GABAergic synapse formation is not a cortex-specific phenomenon. It is noteworthy though that Collagen XIX is generated in only a few brain regions (Su et al., 2010, 2016) making it unlikely to contribute to GABAergic circuit formation in most subcortical regions containing SST<sup>+</sup> interneurons.

### Paracrine roles for synaptogenic cues in the developing brain

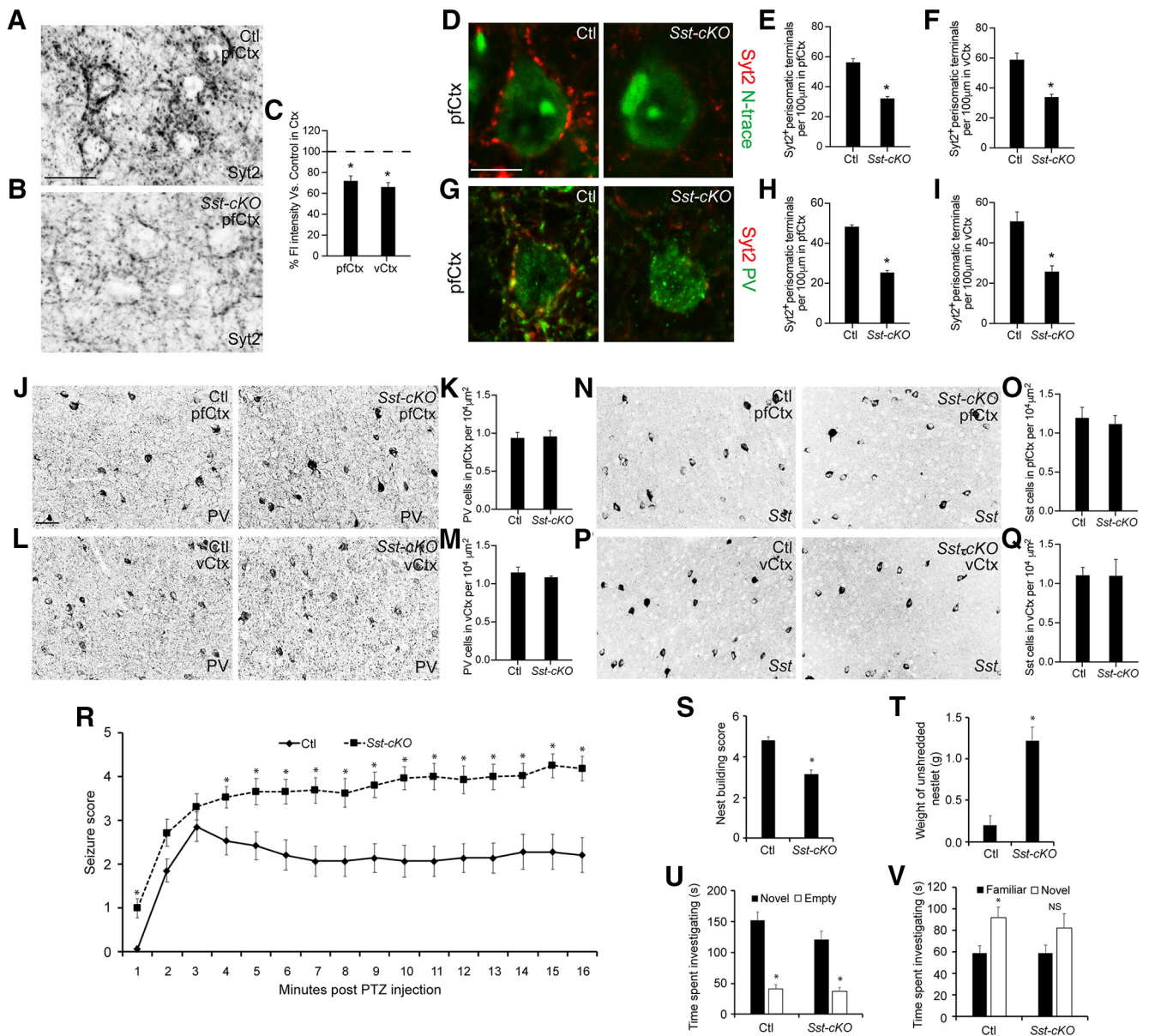
The coordinated transformation of presynaptic and postsynaptic elements into functioning synapses has long been interpreted to suggest a role for the precise exchange of developmentally relevant synaptogenic cues from each synaptic partner. Such signals would not only induce presynaptic and postsynaptic differentiation, but they would also direct the precise alignment of



**Figure 7.** Loss of SST<sup>+</sup> neuron-derived Collagen XIX leads to impaired GAD67<sup>+</sup> inhibitory nerve terminal formation in neocortex. **A, B**, Immunostaining for GAD67 in layer V of prefrontal cortex (pfCtx) in P14 *Col19a1<sup>fl/fl</sup>::Sst-Cre* (*Sst-cKO*) mutants and littermate controls (Ctl). **C**, Mean fluorescent intensity of GAD67 in layer V of pfCtx in P14 *Sst-cKO* and Ctl mice. Data represent the mean  $\pm$  SD;  $n = 3$ . \*Differs from Ctl by  $p = 0.007$  by Student's *t* test. **D, E**, Immunostaining for GAD67 in layer V of visual cortex (vCtx) in P14 *Sst-cKO* and Ctl. **F**, Mean fluorescent intensity of GAD67 in layer V of vCtx in P14 *Sst-cKO* and Ctl mice. Data represent the mean  $\pm$  SD;  $n = 3$ . \*Differs from Ctl by  $p = 0.02$  by Student's *t* test. Scale bar, 20  $\mu$ m.

neurotransmitter release sites (i.e., active zones) with neurotransmitter receptor clusters on the postsynaptic membrane (Fox and Umemori, 2006; Biederer et al., 2017). Studies over the past 2 decades have identified a number of cell adhesion molecules that localize to presynaptic or postsynaptic membranes, bind trans-synaptically to adhere these closely apposed synaptic membranes, and induce the assembly of machinery for neurotransmitter release and reception. An incomplete list of such synaptic adhesion molecules at central synapses includes neuroligins, leucine-rich repeat transmembrane proteins, synaptic cell adhesion molecules, cadherins, EphBs and ephrins, integrins, and Ig superfamily members (Craig et al., 2006; Shen and Scheiffele, 2010; Missler et al., 2012; Jang et al., 2017). As synaptic organizing factors were first identified, it became evident that not all of these cues were embedded in presynaptic or postsynaptic membranes, as neuron-derived secreted cues were discovered that regulate synaptogenesis (including morphogens, growth factors, neurotrophins, and extracellular matrix proteins; Hall et al., 2000; Umemori et al., 2004; Matsuda et al., 2010; Terauchi et al., 2010; Uemura et al., 2010; Kalinovsky et al., 2011; Ito-Ishida et al., 2012; Su et al., 2012; Ferrer-Ferrer and Dityatev, 2018). Moreover, extracellular proteases in or near the synaptic cleft are capable of cleaving the extracellular domains of synaptic adhesion molecules, liberating soluble protein fragments capable of influencing synaptogenesis (Suzuki et al., 2012; Toth et al., 2013). The realization that secreted cues or soluble, shed ectodomains within the extracellular space of the developing brain could impart precise and specific information for inducing synaptogenesis opened the possibility that these synaptogenic cues could be derived from sources other than the synaptic partners and could therefore act in a paracrine fashion (Ferrer-Ferrer and Dityatev, 2018).

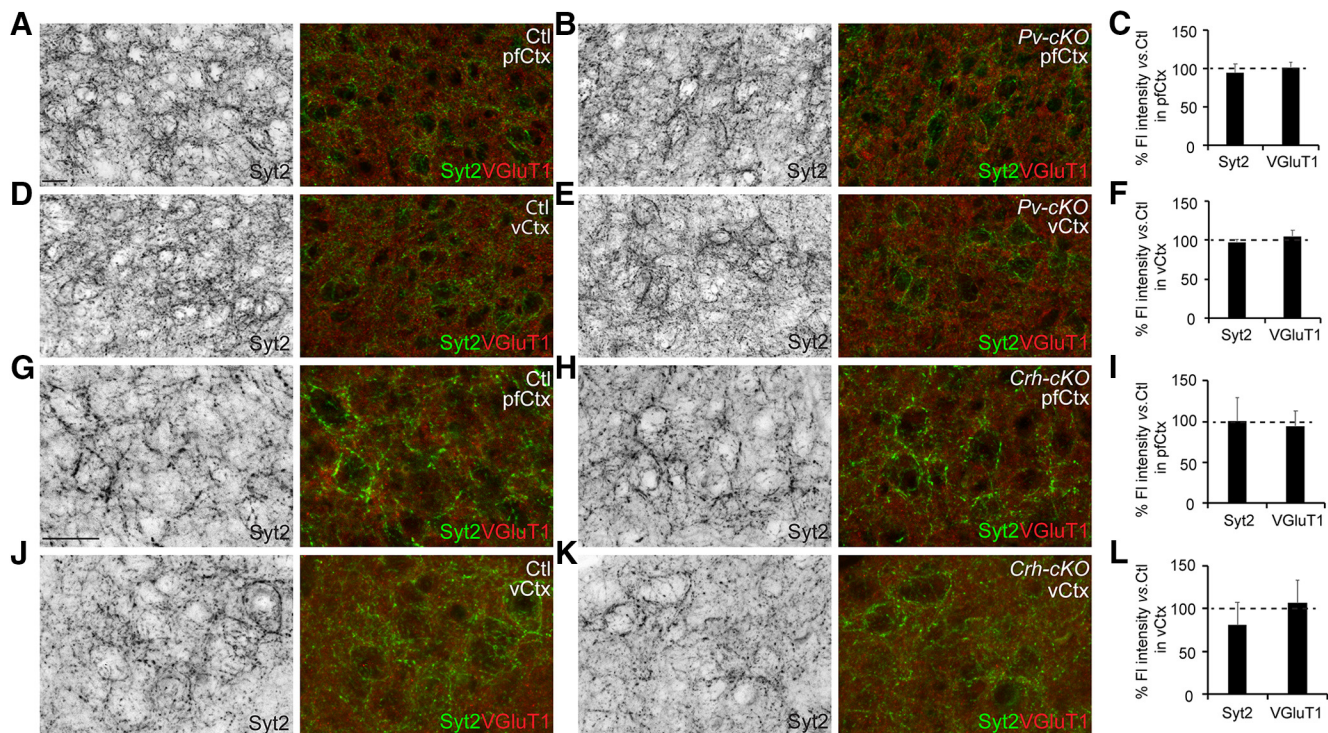
To date, glial-derived factors are the best example of synaptogenic cues that act in paracrine fashion (Eroglu and Barres, 2010; Allen and Eroglu, 2017). Astrocytes extend processes to contact presynaptic and postsynaptic elements in the brain (Bushong et al., 2002) and, thus, occupy prime real estate to deliver developmentally relevant cues to influence presynaptic or postsynaptic development, maturation, and function. Many astrocyte-derived synaptogenic cues are ECM proteins, such as thrombospondins, hevin, glypicans, and chondroitin sulfate



**Figure 8.** Loss of SST<sup>+</sup> neuron-derived Collagen XIX leads to reduced perisomatic synapse number in adult mice and impaired behaviors. **A, B**, Immunostaining for Syt2 in layer V of prefrontal cortex (pfcx) in P100 *Col19a1<sup>fl/fl</sup>::Sst-Cre (Sst-cKO)* mutants and littermate controls (Ctl). **C**, Mean fluorescent intensity of Syt2 in layer V of pfcx and vctx in P100 *Sst-cKO* and Ctl. Data represent the mean  $\pm$  SD;  $n = 3$ . \*Differs from Ctl by  $p = 0.04$  (pfcx) and  $p = 0.0008$  (vctx) by Student's *t* test. **D**, Immunostained Syt2<sup>+</sup> perisomatic synapses were analyzed on NeuroTrace (N-trace)-labeled neurons in single optical sections from layer V of pfcx in P100 *Sst-cKO* and Ctl mice. **E, F**, Reduced numbers of Syt2<sup>+</sup> perisomatic synapses (per unit length of cell soma) on N-trace-labeled neurons in pfcx and vctx of P100 *Sst-cKO* mutants. Data represent the mean  $\pm$  SD;  $n = 3$ . \*Differs from Ctl by  $p = 0.0001$  (pfcx)  $p = 0.0008$  (vctx) by Student's *t* test. **G**, Immunostained Syt2<sup>+</sup> perisomatic synapses were analyzed on PV-immunolabeled neurons in single optical sections from layer V of pfcx in P100 *Sst-cKO* and Ctl mice. **H, I**, Reduced numbers of Syt2<sup>+</sup> perisomatic synapses (per unit length of cell soma) on PV<sup>+</sup> neurons in pfcx and vctx of P100 *Sst-cKO* mutants. Data represent the mean  $\pm$  SD;  $n = 3$ . \*Differs from Ctl by  $p = 0.000008$  (pfcx),  $p = 0.001$  (vctx) by Student's *t* test. **J, L**, IHC for PV<sup>+</sup> cells in vctx and pfcx of P100 *Sst-cKO* and Ctl mice. **K, M**, Quantification of the density of PV<sup>+</sup> cells in vctx and pfcx of P100 *Sst-cKO* and Ctl mice. Data represent the mean  $\pm$  SD;  $n = 3$ . **N, P**, ISH for Sst<sup>+</sup> cells in vctx and pfcx of P100 *Sst-cKO* and Ctl mice. **O, Q**, Quantification of the density of Sst<sup>+</sup> cells in vctx and pfcx of P100 *Sst-cKO* and Ctl mice. Data represent the mean  $\pm$  SD;  $n = 3$ . **R**, Seizure scores for Ctl and *Sst-cKO* mutants were recorded for 15 min after the administration of PTZ. Data represent the mean  $\pm$  SEM;  $n = 14$  Ctl and  $n = 16$  *Sst-cKO*. \*Differs from Ctl by  $p < 0.05$  by Student's *t* test. **S, T**, Nest-building assays. Nests were scored manually after 12 h or by weighing unused nestlets after 12 h. Data represent the mean  $\pm$  SEM;  $n = 6$  Ctl and  $n = 14$  *Sst-cKO*. \*Differs from Ctl by  $p = 0.0003$  for score,  $p = 0.002$  for weight by Student's *t* test. **U, V**, Sociability (**U**) and social memory assays (**V**). *Sst-cKO* and Ctl spent significantly more time investigating novel conspecifics versus empty chambers (**U**). While Ctl mice also spent significantly more time investigating novel conspecifics versus familiar conspecifics, the same was not true for *Sst-cKO* mutants. Data represent the mean  $\pm$  SEM;  $n = 15$  Ctl and 14 *Sst-cKO*. \*Differs from empty or familiar conspecific by  $p = 0.0000001$  (Ctl in **U**),  $p = 0.00,001$  (*Sst-cKO* in **U**), and  $p = 0.01,268$  (Ctl in **V**) by Student's *t* test. NS, Not significantly different. Scale bars: (in **A, B**, 20  $\mu$ m; (in **D, E**, 10  $\mu$ m; (in **J, L, N, P**, 50  $\mu$ m).

proteoglycans (Christopherson et al., 2005; Kucukdereli et al., 2011; Pyka et al., 2011; Allen et al., 2012). Based on the roles of these astrocyte-derived ECM proteins, it is not entirely surprising that an ECM molecule like Collagen XIX could act in a paracrine fashion to regulate the assembly of perisomatic GABAergic

synapses. What is unexpected, however, is that this synaptogenic cue is not generated by a synaptic partner or an adjacent astrocyte. Instead, it is generated by an adjacent interneuron. Thus, at a time when many studies are identifying interneuron-like roles for astrocytes in the brain (i.e., gliotransmission; Papouin et al.,



**Figure 9.** Loss of Collagen XIX from PV<sup>+</sup> or CRH<sup>+</sup> cells does not impair the formation of inhibitory nerve terminals in neocortex. **A, B, D, E**, Immunostaining for Syt2 and VGlut1 in layer V of prefrontal cortex (pfCtx) and visual cortex (vCtx) in P14 *Col19a1<sup>fl/fl</sup>::PV-Cre* (*PV-cKO*) mutants and littermate controls (Ctls). **C, F**, Mean fluorescent intensity of Syt2 and VGlut1 in layer V of pfCtx and vCtx in P14 *PV-cKO* and Ctl mice. Dashed line represents Ctl values. Data represent the mean  $\pm$  SD;  $n = 3$ . **G, H, J, K**, Immunostaining for Syt2 and VGlut1 in layer V of pfCtx and vCtx in P14 *Col19a1<sup>fl/fl</sup>::Crh-Cre* (*Crh-cKO*) mutants and littermate controls (Ctls). **I, L**, Mean fluorescent intensity of Syt2 and VGlut1 in layer V of pfCtx and vCtx in P14 *Crh-cKO* and Ctl. Dashed line represents Ctl values. Data represent the mean  $\pm$  SD;  $n = 3$ . Scale bar, **A**, 20  $\mu$ m.

2017; Fiacco and McCarthy, 2018; Savtchouk and Volterra, 2018), we have identified an astrocyte-like role for SST<sup>+</sup> interneurons in orchestrating GABAergic synapse formation.

#### Matricryptins derived from collagen XIX regulate synapse formation

Collagen XIX is an unconventional, nonfibrillar collagen whose expression is largely restricted to the brain postnatally (Sumiyoshi et al., 1997). Like many nonfibrillar collagens, Collagen XIX harbors a C-terminal noncollagenous domain that can be proteolytically shed as a matricryptin—a bioactive fragment of an ECM protein that exhibits distinct functions from the full-length matrix protein from which it was liberated (Ramont et al., 2007; Ricard-Blum and Salza, 2014). Outside of the nervous system, matricryptins derived from collagens, tenascins, perlecan, and osteopontin influence cell behavior by binding and signaling through integrin receptors (Petitclerc et al., 2000; Kalluri, 2003; Yokosaki et al., 2005; Borza et al., 2006; Saito et al., 2007; Poluzzi et al., 2014; Oudart et al., 2016; Ricard-Blum and Vallet, 2016). Within the nervous system, collagen-derived matricryptins exhibit conserved roles in orchestrating axon outgrowth, synapse formation, and synaptic plasticity in both vertebrate and invertebrate organisms (Ackley et al., 2001; Fox et al., 2007; Fox, 2008; Meyer and Moussian, 2009; Hilario et al., 2010; Latvanlehto et al., 2010; Su et al., 2012, 2016; Wang et al., 2014). Although not all of the mechanisms underlying matricryptin function in the nervous system have been fully elucidated, many induce their action by binding and signaling through integrin receptors, which themselves have long established roles in neural development and synaptogenesis (Park and Goda, 2016; Ferrer-Ferrer and Dityatev, 2018; Lilja and Ivaska, 2018). For example, endostatin, the matricryptin shed

from Collagen XVIII, induces the assembly of cerebellar synapses by signaling through  $\alpha 3 \beta 1$  integrins (Su et al., 2012). In the case of Collagen XIX, a small C-terminal matricryptin (termed NC1[XIX]) is shed that is capable of binding and activating a number of aspartate-glycine-arginine (RGD)-dependent integrins (Oudart et al., 2015, 2016; Su et al., 2016). In the developing neocortex, PV<sup>+</sup> interneurons generate these same RGD-dependent integrins (Su et al., 2016), suggesting that they are able to respond to the synaptogenic influence of NC1[XIX]. *In vitro* experiments support this possibility, as NC1[XIX] triggers GABAergic synapse formation by binding and signaling through  $\alpha 5 \beta 1$  integrins (Su et al., 2016). Taken with the results demonstrated here, this strongly suggests that SST<sup>+</sup> interneurons orchestrate perisomatic GABAergic synapse formation through the release of a synaptogenic, integrin-binding matricryptin.

It is important to point out that Collagen XIX is not the only synaptogenic factor identified that contributes to the assembly and maturation of perisomatic GABAergic synapses in the developing neocortex. Neural cell adhesion molecule, neuroligin 2, L1, Slitrk3, and CXCL12 all contribute to the formation of this synapse (Guan and Maness, 2010; Takahashi et al., 2012; Woo et al., 2013; Liang et al., 2015; Maro et al., 2015; Wu et al., 2017). Why might so many cues be necessary for the assembly of this synapse? Synapse assembly is a multistep process in which priming factors first prepare synaptic partners to be competent to form a synapse, adhesion molecules then promote the interaction of presynaptic and postsynaptic membranes, inductive factors orchestrate the assembly of presynaptic and postsynaptic machinery, and destabilizing/stabilizing factors promote synaptic refinement or maturation (Waites et al., 2005). *In vitro* analyses

suggest that Collagen XIX-derived matricryptins act early in this sequential process as priming factors that promote PV<sup>+</sup> interneurons to transcribe, translate, and traffic presynaptic components to developing nerve terminals (Su et al., 2016). Many priming factors need not act locally at a synapse, but instead appear to act more diffusely, making specific neurons more competent to generate nerve terminals or become integrated into circuits (Waites et al., 2005). As such, SST<sup>+</sup> interneurons may not need to supply Collagen XIX (or NC1[XIX]) directly at sites of developing perisomatic GABAergic synapses, but instead can release it globally into the expansive extracellular space of the developing neocortex (Lehmenkühler et al., 1993; Korogod et al., 2015) or at sites of transient connectivity between SST<sup>+</sup> and PV<sup>+</sup> interneurons (Tuncdemir et al., 2016) for it to prime the assembly of feedforward inhibitory circuits.

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